

Review

# The traditional Chinese medicine *Cordyceps sinensis* and its effects on apoptotic homeostasis

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## Abstract

*Cordyceps sinensis* is a medicinal fungus of Traditional Chinese Medicine. While there are a wide range of reported uses of *Cordyceps sinensis* in the literature, the reports that extracts of this fungus may alter apoptotic homeostasis are most intriguing. However, there are significant challenges regarding research surrounding *Cordyceps sinensis*, such as the difficulty identifying the various species of *Cordyceps* and the many conflicting reports of pharmacological function in the literature. In this review we outline what is known about the ability of *Cordyceps sinensis* to alter apoptotic homeostasis, attempt to reconcile the differences in reported function, identify the challenges surrounding future *Cordyceps sinensis* research, and delineate options for overcoming these critical hurdles.

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**Keywords:** *Cordyceps sinensis*; Apoptosis; Traditional medicine; Plant; Fungus

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## 1. Introduction

The past 20 years has seen a phenomenal growth in the interest in and use of complementary or alternative medicine (CAM), and in the United States 42% of people utilize some form of CAM (Eisenberg et al., 1998). Of the total CAM market, herbal and specialty dietary supplements command a substantial portion with 29% of men and 36% of women reporting current use (Gunther et al., 2004). In 1994 the herbal supplement market boomed, partly as a result of the passage of the Dietary Supplement Health and Education Act (Center for Food Safety and Applied Nutrition, 2004), driving supplement sales up 70% from 1994 to 1997 (Radimer et al., 2000) and leading to peak sales of US\$ 3.3 billion in 1999 (Harnack et al., 2001). This interest in natural compounds is certainly valid as these natural products undoubtedly contain biologically active components: plant-based pharmaceuticals have resulted in approximately one-half of the anti-cancer drugs developed since 1960 (Kim and Park, 2002) and have led to over 100 other successful pharmaceuticals (Farnsworth, 1994). While there is a wide range of available herbal supplements, one of the most interesting supplements is the not yet well-characterized *Cordyceps sinensis* (Berk.) Sacc.

The *Cordyceps sinensis* fungus first gained worldwide attention when it was revealed that several Chinese runners who broke world records in 1993 had included this fungus as part of their training program (Hollobaugh, 1993; Steinkraus and Whitfield, 1994; Starr et al., 1993). In the ten years since the initial reports, *Cordyceps sinensis* has received tremendous attention from the public. Purported effects of the fungus suggested a wide range of biological functions such as use as an aphrodisiac (Bhattarai, 1993), analgesic (Koyama et al., 1997) immune modulator (Gong et al., 2000), and free radical scavenger (Yamaguchi et al., 2000a). However, these effects have not been well analyzed.

Recently, aseptic mycelial cultivation has resulted in establishing a number of *Cordyceps sinensis* derivative cultures. The two most well studied of the cultures, referred to by the anamorph names *Paecilomyces hepiali* (strain CS-4) and *Cephalosporium sinensis*, may present the opportunity to produce a *Cordyceps sinensis* derivative product in a sustainable fashion. While these strains undoubtedly support ecologically sustainable use of *Cordyceps sinensis*, the actual

similarities between wild *Cordyceps sinensis* and the cultures are not clear.

## 2. *Cordyceps sinensis* background and ethnomedical use

*Cordyceps sinensis* is endemic to alpine habitats (3600–5000 m in elevation) on the Tibetan plateau in southwestern China (Fig. 1), where it is a parasite on larvae of moths (Lepidoptera) in the genera *Hepialus* and *Thitarodes* (Kinjo and Zang, 2001). After a host larval infection with either meiotic or mitotic spores, the fungus multiplies in the host by yeast-like budding, eventually killing the host; the fungus then grows in the form of threadlike hyphae. Following overwintering, the fungus ruptures the host body, forming a sexual sporulating structure (a perithecial stroma) that is connected to the dead larva below ground and grows upward to emerge above the soil surface. It is this stroma, either with or without the host larva, which is traditionally used for medicinal purposes (Fig. 2).



Fig. 1. Home range of *Cordyceps sinensis*.



Fig. 2. Fruiting bodies of *Cordyceps sinensis* and dried caterpillars.

### 2.1. Phylogenetic analysis of the genus *Cordyceps*

*Cordyceps* is a genus of perithecial ascomycetes (Phylum Ascomycota) classified in the Clavicipitaceae, a family supported by molecular phylogenetic analyses as a monophyletic group derived from the order Hypocreales (Artjariyasripong et al., 2001; Rehner and Samuels, 1995; Spatafora and Blackwell, 1993; Suh et al., 1998). *Cordyceps* species are parasites of insects or fungi, often exhibiting a high degree of host specificity; as a result of this host specificity, the anamorphic forms of some species (e.g., *Beauveria bassiana*) are widely used as insect biocontrol agents (Huang et al., 2002). According to molecular phylogenetic analyses, *Cordyceps* does not represent a single evolutionary lineage; instead, *Cordyceps* appears to represent several lineages within the Clavicipitaceae (Ito and Hirano, 1997; Artjariyasripong et al., 2001; Sung et al., 2001; Zare et al., 2000). Similarly, the *Cordyceps* species associated with Lepidopteran hosts do not represent a monophyletic group (Nikoh and Fukatsu, 2000; Park et al., 2001; Sung et al., 2001). Furthermore, there appears to be a high degree of genetic variation within the *sinensis* species (Chen and Hseu, 1999). These levels of variation create a significant challenge in verifying samples for analysis of *Cordyceps sinensis*.

### 2.2. Ethnomedical use of *Cordyceps sinensis*

*Cordyceps sinensis* has a long history of medicinal use in China. This fungus is thought to have been discovered 2000 years ago (Liu et al., 2001) and its use was documented formally in the Qing dynasty *Bencao Congxin* (New Compilation of Materia Medica) in 1757. However, *Cordyceps sinensis* is a unique traditional medicine in that there exists little primary ethnomedical data describing medical use in the literature. Current ethnomedical reports on the uses of *Cordyceps sinensis* are limited to the application as a general tonic in China (Huang et al., 1981; Jiang, 1991; Hanssen and Schadler, 1982) and as an aphrodisiac in Nepal (Bhattarai, 1992a, 1993, 1992b, 1994, 1989). In

contrast to the ethnomedical data, the literature surrounding the biological effects of *Cordyceps sinensis* is diverse (reviewed in Zhu et al., 1998a, 1998b). However, some of the most intriguing reports regarding the biological functions of *Cordyceps sinensis* center on its ability to alter apoptotic homeostasis.

## 3. Apoptotic homeostasis and disease states

### 3.1. Review of apoptosis

Apoptosis, or programmed cell death, is an essential event in organism development (Hidalgo and Ffrench-Constant, 2003; Vaux and Korsmeyer, 1999) and homeostasis (Kucharczak et al., 2003, Cory and Adams, 2002); however, it is becoming clear that numerous disorders such as stroke (Zheng et al., 2003), myocardial infarction (Krijnen et al., 2002), and HIV (Buenz and Badley, in press) incorporate apoptosis in their etiology and pathogenesis. There are numerous events that can induce a cell to undergo apoptosis (Nagata, 1997) and since the implication of apoptosis in various disease states as an effector mechanism, the ability to inhibit apoptosis has emerged as an important potential therapy. Interestingly, while inducing apoptosis has already proven an efficient method to treat cancer (Hu and Kavanagh, 2003), the ability to inhibit apoptosis in a clinical setting is just starting to be explored (Liston et al., 2003).

The potential for a cell to undergo apoptosis exists in a balance between endogenous factors characteristic of apoptosis induction, such as Bax (Degli Esposti and Dive, 2003; Scorrano and Korsmeyer, 2003), and factors characteristic of apoptosis inhibition, such as Bcl-2 (Cory et al., 2003; Gross et al., 1999). Once a cell receives sufficient pro-apoptotic stimuli, or lack of anti-apoptotic stimuli, the effector caspases, a family of cysteine aspartate proteases, are activated (Fischer et al., 2003). Cells undergoing apoptosis experience a cascade of events outlined in Fig. 3 that ultimately result in phenotypic changes such as phosphotydylserine expression on the outer cell leaflet (Maderna and Godson, 2003), nuclear condensation, and DNA fragmentation (Nagata et al., 2003) shown in Fig. 4.

### 3.2. Traditional medicines and apoptosis regulation

Many traditional medicines (e.g., *Nelumbo nucifera* (Jung et al., 2003) and *Bacopa monnieri* (Russo et al., 2003)), are reported to scavenge reactive oxygen species and the role of reactive oxygen species in the apoptotic pathway is of particular interest regarding herbal supplements. Reactive oxygen species and their regulatory molecules are important components of the immune system and cell function, such as superoxide radicals generated by activated neutrophils as a pathogen defense mechanism (Babior, 1978), and cell signaling (Ichiki et al., 2003). However, there are also reports of altered oxygen-based free radical levels in disease states. These

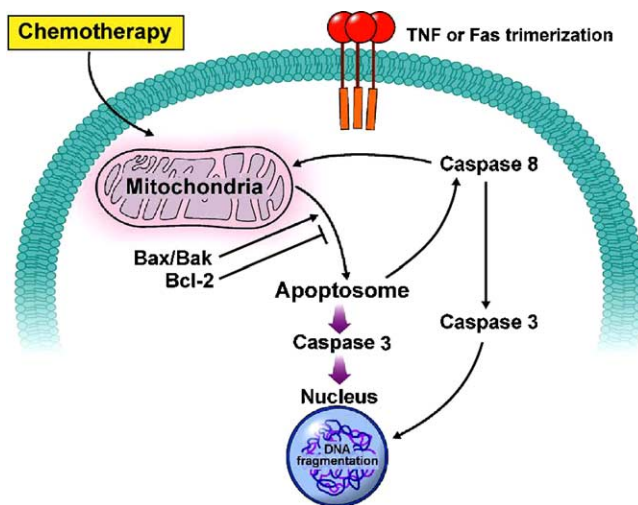


Fig. 3. Fas/CD95 apoptotic pathway. Death receptor trimerization induces a signaling cascade that ultimately results in chromosomal condensation, DNA fragmentation, and cellular apoptosis.

altered levels of reactive oxygen species in disease states such as cancer (Behrend et al., 2003) and stroke (Sugawara and Chan, 2003) have resulted in hypotheses that mitigation of excessive reactive oxygen species could be therapeutically important. While both the exact role of reactive oxygen species in disease states and whether they are a primary insult or a downstream result of the disease states have yet to be determined. It appears that the regulation of reactive oxygen species as therapy will be of significant interest in the future.

Undoubtedly there are herbal supplements that influence apoptotic homeostasis. For example, an isolated compound of the traditional medicine European feverfew (*Chrysanthemum parthenium*) has been shown to decrease susceptibility to apoptotic stimuli through down regulation of the Fas receptor and the Fas ligand agonist through inhibition of NF- $\kappa$ B (Li-

Weber et al., 2002). However, the ability of *Cordyceps sinensis* to alter the apoptotic pathway is not so straightforward. Currently, there are reports of *Cordyceps sinensis* extracts both inhibiting (Table 1) and inducing apoptosis (Table 2). Most of these reports reflect a phenomenon level of observation, such as treatment with *Cordyceps sinensis* resulting in decreased caspase-3 activity (Shahed et al., 2001). While these reports do not examine the mechanism of action, they do facilitate a necessary foundation to allow examination of the molecular mechanism.

#### 4. *Cordyceps sinensis* inhibits apoptosis

The reports of clinical trials suggest that *Cordyceps sinensis* potentially contains agents that may inhibit apoptosis (reviewed in Zhu et al., 1998a). These clinical results have driven work to assess the ability of *Cordyceps sinensis* to inhibit apoptosis in vitro; however, the results of these studies are conflicting. It has been reported that *Cordyceps sinensis* can scavenge reactive oxygen species (Zhang et al., 1995) by inhibiting malondialdehyde formation by the peroxynitrite generator SIN-1 (Yamaguchi et al., 2000a). These results have been confirmed through in vitro xanthine oxidase, hemolysis, and lipid peroxidation assays (Li et al., 2001). Furthermore, an isolated extract of *Cordyceps sinensis* H1-A has been shown to inhibit apoptosis induced by dimethyl sulfoxide (DMSO), which is known to induce apoptosis through permeabilizing the cell membrane and upregulating nitric oxide synthase (Trubiani et al., 2003). However, extracts of *Cordyceps sinensis* were unsuccessful in inhibiting hydrogen peroxide-induced apoptosis (Buenz et al., 2004), a reactive oxygen species model (Fauconneau et al., 2002).

Alternately, *Cordyceps sinensis* has also been reported to down-regulate apoptotic genes and modulate apoptosis in a

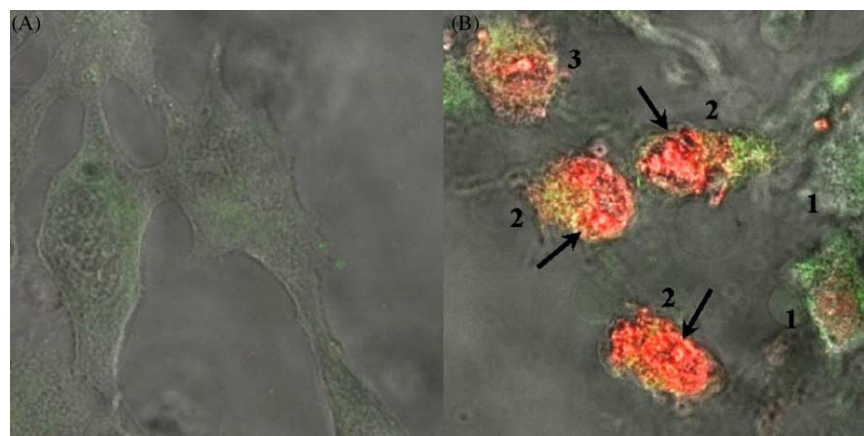


Fig. 4. HeLa cells stained with Annexin V-FITC (green) and propidium iodide (red). (A) Control cells show little phosphatidylserine expression (green) on the outer leaflet, no nuclear propidium iodide intercalation (red), and typical cellular morphology. (B) Cells treated with 30  $\mu$ M starurosporine for 4 h are induced to undergo apoptosis. (1) Early stage apoptotic cells show increased phosphatidylserine expression on the outer leaflet and slight morphological condensation; however, the nuclear membrane is still intact and thus the cells do not stain propidium iodide positive. (2) In later stages of apoptosis, the integrity of the nuclear membrane is compromised and the DNA stains propidium iodide positive. Nuclear condensation and blebbing are also evident (arrows). (3) In the final stages of apoptosis, the cellular and nuclear membranes are totally compromised and the cell appears necrotic.

Table 1  
Reported antiapoptotic effects of *Cordyceps sinensis*

Reported function	Model	Concentration	Reference
Anticytotoxic activity	Mouse	150 mg/kg	Yu et al. (1993)
Antioxidant activity	Mouse	50 mg/kg	Yamaguchi et al. (2000b)
Antiproliferation activity	Cell culture	100 µg/mL	Zhao Long and Xiao Xia (2000)
Cell proliferation inhibition	Cell culture	10 µg/mL	Chen et al. (1997)
Cell proliferation inhibition	Human adult	71 µg/mL	Kuo et al. (1996)
Cell proliferation inhibition	Human adult	40 µg/mL	Lin et al. (1999)
Cell proliferation inhibition	Mouse	1.0 % of diet	Lin et al. (1999)
Gene expression inhibition	Rat	0.5 mL per animal	Shahed et al. (2001)
Hemolysis inhibitory activity	Cell culture	1.5 mg/mL	Li et al. (2001)
Lipid peroxide formation inhibition	Cell culture	5.0 mg/mL	Li et al. (2001)
Natural killer cell inhibition	Human adult	12.9 µg/mL	Kuo et al. (1996)
Radical scavenging effect	Cell culture	5.0 mg/mL	Shahed et al. (2001)
Radical scavenging effect	Cell culture	0.08 mg/mL	Li et al. (2001)
Tumor necrosis factor inhibition	Human adult	2.7 µg/mL	Kuo et al. (1996)

rat kidney ischemia reperfusion model. Shahed et al. (2001) showed a significant decrease in Fas, Fas ligand, and Tumor Necrosis Factor- $\alpha$  (TNF- $\alpha$ ) expression along with decreased caspase-3 activity. Similarly, *Cordyceps sinensis* has been reported to inhibit TNF- $\alpha$  expression (Kuo et al., 1996). However, when apoptosis was initiated through CH-11, a Fas agonist antibody (Alderson et al., 1994), aqueous and alcohol extracts of *Cordyceps sinensis* were unable to rescue cells induced through Fas receptor ligation (Buenz et al., 2004).

Furthermore, it has been shown that in certain cell types, inhibition of proliferation or cell cycle arrest results in cells becoming resistant to apoptosis (Chaturvedi et al., 1999). In turn, the reports that *Cordyceps sinensis* inhibits proliferation of leukemic U937 cells (Chen et al., 1997) and glomerular mesangial cells (Zhao Long and Xiao Xia, 2000; Lin et al., 1999) may result by conferring a relatively apoptotic resistant state to cells. While the mechanisms of this inhibition have yet to be characterized, it is reasonable to propose that the alteration may potentially involve p53 (Fridman and Lowe, 2003) or NF- $\kappa$ B (Karin et al., 2004).

### 5. *Cordyceps sinensis* induces apoptosis

The ability to induce apoptosis has been identified and utilized in successful cancer chemotherapeutics (Hu and Kavanagh, 2003); Table 2 outlines the literature suggesting or stating the ability of *Cordyceps sinensis* to induce apoptosis.

Table 2  
Reported apoptotic effects of *Cordyceps sinensis*

Reported function	Model	Dose	Reference
Antitumor activity	Mouse	Not stated	Zang et al. (1985)
Antitumor activity	Mouse	5.0 g/kg	Xu et al. (1992)
Cell proliferation stimulation	Cell culture	10 µg/mL	Chen et al. (1997)
Cytotoxic activity	Cell culture	2.0 µg/mL	Kuo et al. (1994)
Cytotoxic activity	Cell culture	500 µg/mL	Sato (1989)
Cytotoxic activity	Cell culture	10 µg/mL	Nakamura et al. (1999)
Metastasis inhibition	Mouse	100 mg/kg	Nakamura et al. (1999)

Recently, Yang et al. (Yang et al., 2003) described the ability of the previously isolated 410 kDa polysaccharide fraction of *Cordyceps sinensis* termed H1-A (Yang et al., 1999) to induce apoptosis through inhibiting phosphorylation of Bcl-2 and Bcl-xL. These anti-apoptotic Bcl-2 family members are known to sequester cytosolic pro-apoptotic proteins such as Bax. As addressed above, this fraction also inhibited apoptosis induced by dimethyl sulfoxide (DMSO) (Yang et al., 2003). However the report consisting of the inhibition of Bcl-2 and Bcl-xL and subsequent inhibition of apoptosis by DMSO (Yang et al., 2003) seems counterintuitive. Thus, further work is necessary to clarify the physiologic role of the H1-A extract.

Finally, there are reports of direct cytotoxic activity (Nakamura et al., 1999; Kuo et al., 1994; Sato, 1989). However, these studies report inhibition at the phenomena level and do not address specific mechanisms. Yet it is interesting that cordycepin, a compound originally isolated from the *Cordyceps sinensis* relative *Cordyceps militaris* (Cunningham et al., 1950), is known to exert cytotoxic effects through nucleic acid methylation (Kredich, 1980). While isolation of this single active compound may set the stage for work to identify molecular mechanisms of action, the actual presence of cordycepin in *Cordyceps sinensis* has been difficult to confirm. Cordycepin has been shown to be present in *Cordyceps sinensis* through nuclear magnetic resonance (Chen and Chu, 1996); however, other groups have not been able to detect this compound (Shiao et al., 1994).

## 6. The challenge of identifying *Cordyceps sinensis*

The most significant challenge working with *Cordyceps sinensis* is the lack of a well-defined mechanism to identify sample material. Although *Cordyceps* species have been included in a number of recent molecular phylogenetic analyses (Artjariyasripong et al., 2001; Ito and Hirano, 1997; Lumbsch et al., 2000; Nikoh and Fukatsu, 2000; Nikoh and Fukatsu, 2001; Obornik et al., 2001; Park et al., 2001; Spatafora and Blackwell, 1993; Suh et al., 2001; Suh et al., 1998; Sung et al., 2001; Zare et al., 2000), most of these studies have either not included *Cordyceps sinensis*, or have included *Cordyceps sinensis* without enough other taxa to allow phylogenetic resolution to be achieved among *Cordyceps sinensis* and other closely related *Cordyceps* species. Most DNA-based studies that include *Cordyceps sinensis* have examined genetic differentiation at the population level rather than at the species level (Chen et al., 2001; Kinjo and Zang, 2001; Liu et al., 2001). Using a neighbor-joining analysis of ribosomal DNA internal transcribed spacer (rDNA-ITS) sequences, Park et al. (Park et al., 2001) determined *Cordyceps sinensis* to be closely related to *Cordyceps ophioglossoides*, a parasite of “false truffle” fungi in the genus *Elaphomyces*. Based on this evidence, as well as the placement of a *Hirsutella* anamorph (though with low bootstrap support) within a larger phylogenetic clade that includes *Cordyceps ophioglossoides* (Sung et al., 2001), a possible phylogenetic placement for *Cordyceps sinensis* is within a basal clade in the Clavicipitaceae that contains both entomopathogenic and fungicolous fungi. However, direct molecular evidence for the proper phylogenetic placement of *Cordyceps sinensis* in relation to a polyphyletic *Cordyceps* is currently lacking, and *Cordyceps* and the family Clavicipitaceae are greatly in need of systematic revision.

Many ascomycetes, including species of *Cordyceps*, have both an asexual (anamorphic) and sexual (teleomorphic) form. *Cordyceps* species have been shown in mycological culture studies to be associated with a number of anamorphic genera including *Paecilomyces*, *Beauvaria*, *Metarhizium*, *Verticillium*, and *Tolyposcladium* (Hodge et al., 1996; Huang et al., 2002; Nikoh and Fukatsu, 2000; Nikoh and Fukatsu, 2001; Obornik et al., 2001; Sung et al., 2001; Zare et al., 2000), and phylogenetically related to several species of yeast-like endosymbionts of insects (Suh et al., 2001). Determining the anamorphic state of *Cordyceps sinensis* has posed difficulty for researchers; as a result, 22 names in 13 anamorph genera have previously been associated with *Cordyceps sinensis* (Jiang and Yao, 2002). However, recent molecular evidence (Chen et al., 2001; Chen et al., 2002; Liu et al., 2001) and generation of the anamorphic state from germinated *Cordyceps sinensis* ascospores (Liu et al., 2001) supports *Hirsutella sinensis* Liu et al. as being the correct anamorph of *Cordyceps sinensis*. Many anamorphic fungi, including *Hirsutella sinensis*, can be cultured under laboratory conditions. Because *Cordyceps sinensis* is considered

to be declining in the wild due to overharvest (Liu et al., 2003), culture of the anamorphic state may therefore be a means of producing material for research and medicinal preparations in the face of a shortage of teleomorph material.

Several *Cordyceps* species have been described that appear morphologically similar to *Cordyceps sinensis*, including *Cordyceps nepalensis* M. Zang & Kinjo, *Cordyceps multiaxialis* M. Zang & Kinjo, *Cordyceps gansuensis* Zhang et al., and *Cordyceps crassisporea* Zang et al. However, these species may simply represent morphological variants of *Cordyceps sinensis* (Kinjo and Zang, 2001; Liu et al., 2001). Whether these morphotypes correspond to differences in pharmacological activity has not been determined. Other *Cordyceps* species (e.g., *Cordyceps militaris*) have been shown to have some of the same medicinal properties as *Cordyceps sinensis* (Wu et al., 2000), but appear to be less highly regarded by consumers and practitioners.

## 7. Strain typing and relations to product quality

The lack of molecular evidence for proper phylogenetic placement of *Cordyceps sinensis* precludes establishing a consensus strain of *Cordyceps sinensis*. Similarly, conducting reliable clinical trials and ascertaining the quality of commercial herbal and other natural products depends upon accurately identifying source materials. A number of factors could contribute to poor quality of *Cordyceps sinensis* products, including intentional substitution of other *Cordyceps* species, substitution of counterfeit material, or unintentional misidentification of field-collected *Cordyceps*. In addition, as demand for *Cordyceps sinensis* products grows and the supply of wild material declines, mycelium of the asexual (anamorphic) stage grown under artificial culture conditions is increasingly used in medicinal products. Among the source materials found to have been sold under the name Chongcao in commercial markets are other *Cordyceps* species (e.g., *Cordyceps militaris*, *Cordyceps liangshanensis*, *Cordyceps gunnii*, *Cordyceps hawkesii*, and *Cordyceps ramosa*), anamorphic fungi including *Hirsutella sinensis*, the anamorph of *Cordyceps sinensis*, as well as *Paecilomyces sinensis* and *Tolyposcladium* sp., and rhizomes from the plant species *Stachys geobombycis* and *Stachys sieboldii* (Chen et al., 2002; Cheng et al., 1998). Even within *Cordyceps sinensis*, differences in pharmacological activity have been noted between strains (Kinjo and Zang, 2001).

Because commercial products generally contain dried, powdered material, identifying source material using morphological characters is normally impossible. DNA fingerprinting methods offer a dependable means of identifying source materials from fresh specimens and commercial preparations (Zerega et al., 2002). Such methods are able to distinguish isolates at the species, or even strain, level depending upon the specific method used (e.g., (Terashima

et al., 2002)). The effectiveness of DNA fingerprinting for authenticating source material has been demonstrated for a number of pharmacologically useful plants, including skullcap (*Scutellaria* spp.; (Hosokawa et al., 2000)), *Echinacea* (Nieri et al., 2003), black cohosh (*Actaea racemosa*; (Zerega et al., 2002)), *Lycium* (Zhang et al., 2001), ginseng (*Panax ginseng*; (Hon et al., 2003, Ngan et al., 1999)), *Withania* (Negi et al., 2000), St. John's wort (*Hypericum perforatum*; (Mayo and Langridge, 2003)), opium poppy (*Papaver somniferum*; (Saunders et al., 2001)), coca (*Erythroxylum* spp.; (Johnson et al., 2003)), and yarrow (*Achillea millefolium*; (Wallner et al., 1996)). DNA fingerprinting has been successfully used in fungi to distinguish strains of human pathogenic fungi (Buffington et al., 1994; Mcewen et al., 2000), plant pathogens (Barnes et al., 2001; Chen et al., 2000; Morris et al., 2000; Schmidt et al., 2003; Zeller et al., 2003), food spoilage agents (Kure et al., 2003), biocontrol agents (Avis et al., 2001; Hermosa et al., 2001), and edible mushrooms (Terashima et al., 2002).

Previous genetic analyses of multiple *Cordyceps sinensis* populations have examined ribosomal DNA (rDNA) sequence diversity (Chen and Hseu, 1999; Chen et al., 2001; Chen et al., 2002; Kinjo and Zang, 2001; Liu et al., 2001) and patterns of genetic variability exhibited by randomly amplified polymorphic DNA (RAPD) markers (Chen et al., 1999; Cheng et al., 1998). Variability in ribosomal ITS sequences and 18S ribosomal DNA restriction fragment length polymorphism (RFLP) patterns has been shown to be informative for differentiating *Cordyceps sinensis* from other *Cordyceps* species and from market counterfeits (Chen et al., 1999; Chen et al., 2001; Chen et al., 2002; Kinjo and Zang, 2001; Liu et al., 2001), and for generating *Cordyceps sinensis*-specific DNA probes (Chen et al., 2002); however, rDNA sequence variation is too low to allow accurate genotyping at the strain level (Chen et al., 2001; Kinjo and Zang, 2001; Liu et al., 2001). Randomly amplified polymorphic DNA markers are able not only to distinguish *Cordyceps sinensis* from other *Cordyceps* species, but to distinguish individual *Cordyceps sinensis* populations (Chen and Hseu, 1999; Cheng et al., 1998). More recently developed DNA fingerprinting methods such as amplified fragment length polymorphisms (AFLP; Vos et al., 1995) uncover more genetic polymorphism over a larger part of the genome than do ribosomal DNA sequences. Additionally, the results of AFLP fingerprinting have been shown to have a higher degree of repeatability than those of RAPD fingerprinting (Ranamukhaarachchi et al., 2000; Jones et al., 1997; Barker et al., 1999). AFLP fingerprints have thus far not been obtained for *Cordyceps sinensis*, and represent a potentially useful tool for characterizing *Cordyceps sinensis* samples. Further population-level genetic characterization is extremely important for *Cordyceps sinensis* material in order to determine the geographic origins of source material, select standardized strains for clinical experiments, distinguish anamorph cultures from fungal contaminants, and facilitate conservation of genetic diversity in natural populations.

## 8. The future of apoptosis regulation through *Cordyceps sinensis*

There are three prominent factors that may contribute to the discrepancies in reports regarding the ability of *Cordyceps sinensis* to inhibit apoptosis. First, as there is no consensus strain, it is possible that certain populations contain different biologically active compounds. Second, there is the potential that *Cordyceps sinensis* extracts contain a pro-drug and there is a necessary metabolism step in order to generate the biologically active form of the drug. Third, as there are multiple methods of extraction utilized in the literature such as ethanol (Xu et al., 1992), methanol (Kuo et al., 1994), alkaline extract (Kihō et al., 1996), hot water (Manabe et al., 1996), and cold water (Chen et al., 1997), different constituents may be assayed depending on extraction method. However, these challenges are certainly not intractable. The *Cordyceps sinensis* consensus strain could be established through AFLP-based DNA fingerprinting as has been done for other medicinal plants. Similarly, the issue regarding an active metabolite could be addressed through either establishing a liposome system containing enzymes known to be important in drug metabolism such as members of the P450 family (De Graaf et al., 2002) or, alternately, it may be possible to treat the *Cordyceps sinensis* extract with a liver homogenate to generate the active metabolites of the extracts (De Graaf et al., 2002).

Furthermore, the research surrounding *Cordyceps sinensis* is lacking molecular studies. Rather, there are numerous reports of treating model systems with *Cordyceps sinensis* and measuring outcome. However, establishing a consensus strain is an essential foundation to defining molecular mechanisms. Currently the use of voucher specimens and marker compounds does allow identification of species phenotypically and biochemically, respectively. However, a principle marker compound for *Cordyceps sinensis* is cordycepic acid (mannitol-D) and while the presence of cordycepic acid is indicative of *Cordyceps sinensis*, the inclusion of a single marker compound does not necessarily guarantee the presence of other potentially active compounds. Thus, developing an AFLP based DNA fingerprinting program would allow positive identification of not just *Cordyceps sinensis*, but also identification of sub-populations of the species.

*Cordyceps sinensis* has been used as a traditional medicine throughout history and, undoubtedly, as investigation into this fungus continues, more active components with potential therapeutic value will be isolated.

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## References

- Alderson, M.R., Tough, T.W., Braddy, S., Davis-Smith, T., Roux, E., Schooley, K., Miller, R.E., Lynch, D.H., 1994. Regulation of apoptosis and T cell activation by Fas-specific mAb. *International Immunology* 6, 1799–1806.
- Artjariyasirong, S., Mitchell, J.I., Hywel-Jones, N.L., Jones, E.B.G., 2001. Relationship of the genus *Cordyceps* and related genera, based on parsimony and spectral analysis of partial 18S and 28S ribosomal gene sequences. *Mycoscience* 42, 503–517.
- Avis, T.J., Caron, S.J., Boekhout, T., Hamelin, R.C., Belanger, R.R., 2001. Molecular and physiological analysis of the powdery mildew antagonist *Pseudozyma flocculosa* and related fungi. *Phytopathology* 91, 249–254.
- Babior, B.M., 1978. Oxygen-dependent microbial killing by phagocytes (first of two parts). *New England Journal of Medicine* 298, 659–668.
- Barker, J.H.A., Matthes, M., Arnold, G.M., Edwards, K.J., Ahman, I., Larsson, S., Karp, A., 1999. Characterisation of genetic diversity in potential biomass willows (*Salix* spp.) by RAPD and AFLP analyses. *Genome/National Research Council Canada* 42, 173–183.
- Barnes, I., Gaur, A., Burgess, T., Roux, J., Wingfield, B.D., Wingfield, M.J., 2001. Microsatellite markers reflect intra-specific relationships between isolates of the vascular wilt pathogen *Ceratocystis fimbriata*. *Molecular Plant Pathology* 2, 319–325.
- Behrend, L., Henderson, G., Zwacka, R.M., 2003. Reactive oxygen species in oncogenic transformation. *Biochemical Society Transactions* 31, 1441–1444.
- Bhattarai, N.K., 1989. Traditional phytotherapy among the Sherpas of Helambu, central Nepal. *Journal of Ethnopharmacology* 27, 45–54.
- Bhattarai, N.K., 1992a. Folk use of plants in veterinary medicine in central Nepal. *Fitoterapia* 63, 497–506.
- Bhattarai, N.K., 1992b. Medical ethnobotany in the Karnali Zone, Nepal. *Economic Botany* 46, 257–261.
- Bhattarai, N.K., 1993. Folk herbal medicines of Dolakha District, Nepal. *Fitoterapia* 66, 387–395.
- Bhattarai, N.K., 1994. Folk herbal remedies for gynaecological complaints in central Nepal. *International Journal of Pharmacognosy* 32, 13–26.
- Buenz, E., Badley, A., in press. Impact of mitochondrial regulation of apoptosis on the pathogenesis of HIV-1 induced immunodeficiency. *Mitochondrion*. In press.
- Buenz, E.J., Weaver, J.G., Bauer, B.A., Chalpin, S.D., Badley, A.D., 2004. *Cordyceps sinensis* extracts do not prevent Fas-receptor and hydrogen peroxide-induced T-cell apoptosis. *Journal of Ethnopharmacology* 90, 57–62.
- Buffington, J., Reporter, R., Lasker, B.A., Mcneil, M.M., Lanson, J.M., Ross, L.A., Mascola, L., Jarvis, W.R., 1994. Investigation of an epidemic of invasive aspergillosis: utility of molecular typing with the use of random amplified polymorphic DNA probes. *The Pediatric Infectious Disease Journal* 13, 386–393.
- Center for Food Safety and Applied Nutrition, Dietary Supplement Health and Education Act of 1994. 2004.
- Chaturvedi, V., Qin, J.Z., Denning, M.F., Choubey, D., Diaz, M.O., Nickoloff, B.J., 1999. Apoptosis in proliferating, senescent, and immortalized keratinocytes. *The Journal of Biological Chemistry* 274, 23358–23367.
- Chen, C.-S., Hseu, R.-S., 1999. Differentiation of *Cordyceps sinensis* (Berk.) Sacc. specimens using restriction fragment length polymorphism of 18S rRNA gene. *Journal of the Chinese Agricultural Chemical Society* 37, 533–545.
- Chen, S.Z., Chu, J.Z., 1996. NMR and IR studies on the characterization of cordycepin and 2'-deoxyadenosine. *Zhongguo kang sheng su zhi* (Chinese Journal of Antibiotics) 21, 9–12.
- Chen, W.-D., Grau, C.R., Adee, E.A., Meng, X.-Q., 2000. A molecular marker identifying subspecific populations of the soybean brown stem rot pathogen, *Phialophora gregata*. *Phytopathology* 90, 875–883.
- Chen, Y.J., Shiao, M.S., Lee, S.S., Wang, S.Y., 1997. Effect of *Cordyceps sinensis* on the proliferation and differentiation of human leukemic U937 cells. *Life Sciences* 60, 2349–2359.
- Chen, Y.-J., Zhang, Y.-P., Yang, Y.-X., Yang, D.-R., 1999. Genetic diversity and taxonomic implication of *Cordyceps sinensis* as revealed by RAPD markers. *Biochemical Genetics* 37, 201–213.
- Chen, Y.-Q., Wang, N., Qu, L.-H., Li, T.-H., Zhang, W.-M., 2001. Determination of the anamorph of *Cordyceps sinensis* inferred from the analysis of the ribosomal DNA internal transcribed spacers and 5.8 S rDNA. *Biochemical Systematics and Ecology* 29, 597–607.
- Chen, Y.-Q., Wang, N., Zhou, H., Qu, L.-H., 2002. Differentiation of medicinal *Cordyceps* species by rDNA ITS sequence analysis. *Planta Medica* 68, 635–639.
- Cheng, K.-T., Su, C.-H., Chang, H.-C., Huang, J.-Y., 1998. Differentiation of genuines and counterfeits of *Cordyceps* species using random amplified polymorphic DNA. *Planta Medica* 64, 451–453.
- Cory, S., Adams, J.M., 2002. The Bcl2 family: regulators of the cellular life-or-death switch. *Nature Reviews Cancer* 2, 647–656.
- Cory, S., Huang, D., Adams, J., 2003. The Bcl-2 family: roles in cell survival and oncogenesis. *Oncogene* 22, 8590–8607.
- Cunningham, K.G., Manson, W., Spring, F.S., Hutchinson, S.A., 1950. Cordycepin, a metabolic product isolated from cultures of *Cordyceps militaris* (Linn.) Link. *Nature* 166, 949.
- De Graaf, I.A., Van Meijeren, C.E., Pektas, F., Koster, H.J., 2002. Comparison of in vitro preparations for semi-quantitative prediction of in vivo drug metabolism. *Drug Metabolism and Disposition* 30, 1129–1136.
- Degli Esposti, M., Dive, C., 2003. Mitochondrial membrane permeabilisation by Bax/Bak. *Biochemical and Biophysical Research Communications* 304, 455–461.
- Eisenberg, D.M., Miller, F.H., Curto, D.A., Kaptchuk, T.J., Brennan, T.A., 1998. Trends in alternative medicine use in the United States, 1990–1997: results of a follow-up national survey [comment]. *JAMA: The Journal of the American Medical Association* 280, 1610–1615.
- Farnsworth, N., 1994. Ethnopharmacology and drug development. *Ethnobotany and the search for new drugs*. Ciba Foundation Symposium 185, 42–59.
- Fauconneau, B., Petegnief, V., Sanfeliu, C., Piriou, A., Planas, A.M., 2002. Induction of heat shock proteins (HSPs) by sodium arsenite in cultured astrocytes and reduction of hydrogen peroxide-induced cell death. *Journal of Neurochemistry* 83, 1338–1348.
- Fischer, U., Janicke, R.U., Schulze-Osthoff, K., 2003. Many cuts to ruin: a comprehensive update of caspase substrates. *Cell Death and Differentiation* 10, 76–100.
- Fridman, J.S., Lowe, S.W., 2003. Control of apoptosis by p53. *Oncogene* 22, 9030–9040.
- Gong, X.J., Ji, H., Lu, S.G., Li, S.P., Li, P., 2000. Effects of polysaccharides from cultured *Cordyceps sinensis* on murine immunologic function. *Zhongguo Yaoke Daxue Xuebao* 31, 53–55.
- Gross, A., McDonnell, J.M., Korsmeyer, S.J., 1999. BCL-2 family members and the mitochondria in apoptosis. *Genes and Development* 13, 1899–1911.
- Gunther, S., Patterson, R.E., Kristal, A.R., Stratton, K.L., White, E., 2004. Demographic and health-related correlates of herbal and specialty supplement use. *Journal of the American Dietetic Association* 104, 27–34.
- Hanssen, H.P., Schadler, M., 1982. Mushrooms as folk remedy from Chinese medicine. *Deutsche Apotheker-Zeitung* 122, 1844–1848.
- Harnack, L.J., Rydell, S.A., Stang, J., 2001. Prevalence of use of herbal products by adults in the Minneapolis/St Paul, Minn, metropolitan area. *Mayo Clinic Proceedings* 76, 688–694.

- Hermosa, M.R., Grondona, I., Diaz-Minguez, J.M., Iturriaga, E.A., Monte, E., 2001. Development of a strain-specific SCAR marker for the detection of *Trichoderma atroviride* 11, a biological control agent against soilborne fungal plant pathogens. *Current Genetics* 38, 343–350.
- Hidalgo, A., Ffrench-Constant, C., 2003. The control of cell number during central nervous system development in flies and mice. *Mechanisms of Development* 120, 1311–1325.
- Hodge, K.T., Krasnoff, S.B., Humber, R.A., 1996. *Tolypocladium inflatum* is the anamorph of *Cordyceps subsessilis*. *Mycologia* 88, 715–719.
- Hollobaugh, J., 1993. A giant awakens. *Track & Field News* 46.
- Hon, C.C., Chow, Y.C., Zeng, F.Y., Leung, F.C.C., 2003. Genetic authentication of ginseng and other traditional Chinese medicine. *Acta Pharmacologica Sinica* 24, 841–846.
- Hosokawa, K., Minami, M., Kawahara, K., Nakamura, I., Shibata, T., 2000. Discrimination among three species of medicinal *Scutellaria* plants using RAPD markers. *Planta Medica* 66, 270–272.
- Hu, W., Kavanagh, J.J., 2003. Anticancer therapy targeting the apoptotic pathway. *The Lancet Oncology* 4, 721–729.
- Huang, B., Li, C.-R., Li, Z.-G., Fan, M.-Z., Li, Z.-Z., 2002. Molecular identification of the teleomorph of *Beauveria bassiana*. *Mycotaxon* 81, 229–236.
- Huang, H.T., Chou, S.H., Ho, H.L., 1981. Comparison of chemical constituents between *Cordyceps hawkesii* and *Cordyceps sinensis*. *Chinese Pharmaceutical Bulletin* 16, 53.
- Ichiki, T., Tokunou, T., Fukuyama, K., Iino, N., Masuda, S., Takeshita, A., 2003. Cyclic AMP response element-binding protein mediates reactive oxygen species-induced c-fos expression. *Hypertension* 42, 177–183.
- Ito, Y., Hirano, T., 1997. The determination of the partial 18S ribosomal DNA sequences of *Cordyceps* species. *Letters in Applied Microbiology* 25, 239–242.
- Jiang, S.J., 1991. Immunomodulating effects of cordyceps sinensis. *International Journal of Oriental Medicine* 16, 128–130.
- Jiang, Y., Yao, Y.-J., 2002. Names related to *Cordyceps sinensis* anamorph. *Mycotaxon* 84, 245–254.
- Johnson, E.L., Saunders, J.A., Mischke, S., Helling, C.S., Emche, S.D., 2003. Identification of *Erythroxyllum* taxa by AFLP DNA analysis. *Phytochemistry* 64, 187–197.
- Jones, C.J., Edwards, K.J., Castaglione, S., Winfield, M.O., Sala, F., Van De Wiel, C., Bredemeijer, G., Vosman, B., Matthes, M., Daly, A., Brettschneider, R., Bettini, P., Buiatti, M., Maestri, E., Malcevski, A., Marmioli, N., Aert, R., Volckaert, G., Rueda, J., Linacero, R., Vazquez, A., Karp, A., 1997. Reproducibility testing of RAPD, AFLP and SSR markers in plants by a network of European laboratories. *Molecular Breeding* 3, 381–390.
- Jung, H.A., Kim, J.E., Chung, H.Y., Choi, J.S., 2003. Antioxidant principles of *Nelumbo nucifera* stamens. *Archives of Pharmacol Research* 26, 279–285.
- Karin, M., Yamamoto, Y., Wang, Q.M., 2004. The IKK NF-kappa B system: a treasure trove for drug development. *Nature Reviews. Drug Discovery* 3, 17–26.
- Kiho, T., Yamane, A., Hui, J., Usui, S., Ukai, S., 1996. Polysaccharides in fungi. XXXVI. Hypoglycemic activity of polysaccharide (CS-F30) from the cultural mycelium of *Cordyceps sinensis* and its effect on glucose metabolism in mouse liver. *Biological & Pharmaceutical Bulletin* 19, 294–296.
- Kim, J., Park, E.J., 2002. Cytotoxic anticancer candidates from natural resources. *Current Medicinal Chemistry. Anti-Cancer Agents* 2, 485–537.
- Kinjo, N., Zang, M., 2001. Morphological and phylogenetic studies on *Cordyceps sinensis* distributed in southwestern China. *Mycoscience* 42, 567–574.
- Koyama, K., Imaizumi, T., Akiba, M., Kinoshita, K., Takahashi, K., Suzuki, A., Yano, S., Horie, S., Watanabe, K., Naoi, Y., 1997. Antinociceptive components of ganoderma lucidum. *Planta Medica* 63, 224–227.
- Kredich, N.M., 1980. Inhibition of nucleic acid methylation by cordycepin. In vivo synthesis of S-3'-deoxyadenosylmethionine by WI-L2 human lymphoblasts. *The Journal of Biological Chemistry* 255, 7380–7385.
- Krijnen, P.A., Nijmeijer, R., Meijer, C.J., Visser, C.A., Hack, C.E., Niessen, H.W., 2002. Apoptosis in myocardial ischaemia and infarction. *Journal of Clinical Pathology* 55, 801–811.
- Kucharczak, J., Simmons, M., Fan, Y., Gelinas, C., 2003. To be, or not to be: NF-kappaB is the answer—role of Rel/NF-kappaB in the regulation of apoptosis. *Oncogene* 22, 8961–8982.
- Kuo, Y.C., Lin, C.Y., Tsai, W.J., Wu, C.L., Chen, C.F., Shiao, M.S., 1994. Growth inhibitors against tumor cells in *Cordyceps sinensis* other than cordycepin and polysaccharides. *Cancer Investigation* 12, 611–615.
- Kuo, Y.C., Tsai, W.J., Shiao, M.S., Chen, C.F., Lin, C.Y., 1996. *Cordyceps sinensis* as an immunomodulatory agent. *American Journal of Chinese Medicine* 24, 111–125.
- Kure, C.F., Skaar, I., Holst-Jensen, A., Abeln, E.C.A., 2003. The use of AFLP to relate cheese-contaminating *Penicillium* strains to specific points in the production plants. *International Journal of Food Microbiology* 83, 195–204.
- Li, S.P., Li, P., Dong, T.T.X., Tsim, K.W.K., 2001. Anti-oxidation activity of different types of natural *Cordyceps sinensis* and cultured *Cordyceps mycelia*. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology* 8, 207–212.
- Lin, C.Y., Ku, F.M., Kuo, Y.C., Chen, C.F., Chen, W.P., Chen, A., Shiao, M.S., 1999. Inhibition of activated human mesangial cell proliferation by the natural product of cordyceps sinensis (HI-1): an implication for treatment of IGA mesangial nephropathy. *The Journal of Laboratory and Clinical Medicine* 133, 55–63.
- Liston, P., Fong, W., Korneluk, R., 2003. The inhibitors of apoptosis: there is more to life than Bcl2. *Oncogene* 22, 8568–8580.
- Liu, P.-G., Wang, X.-H., Yu, F.-Q., Zheng, H.-D., Chen, J., 2003. Key taxa of larger members in higher fungi of biodiversity from China. *Acta Botanica Yunnanica* 25, 285–296.
- Liu, Z.-Y., Yao, Y.-J., Liang, Z.-Q., Liu, A.-Y., Pegler, D.N., Chase, M.W., 2001. Molecular evidence for the anamorph-teleomorph connection in *Cordyceps sinensis*. *Mycological Research* 105, 827–832.
- Li-Weber, M., Giaisi, M., Baumann, S., Treiber, M.K., Krammer, P.H., 2002. The anti-inflammatory sesquiterpene lactone parthenolide suppresses CD95-mediated activation-induced-cell-death in T-cells. *Cell Death and Differentiation* 9, 1256–1265.
- Lumsch, H.T., Lindemuth, R., Schmitt, I., 2000. Evolution of filamentous ascomycetes inferred from LSU rDNA sequence data. *Plant Biology* 2, 525–529.
- Maderna, P., Godson, C., 2003. Phagocytosis of apoptotic cells and the resolution of inflammation. *Biochimica et Biophysica Acta* 1639, 141–151.
- Manabe, N., Sugimoto, M., Azuma, Y., Taketomo, N., Yamashita, A., Tsuboi, H., Tsunoo, A., Kinjo, N., Huang, N.L., Miyamoto, H., 1996. Effects of the mycelial extract of cultured *Cordyceps sinensis* on in vivo hepatic energy metabolism in the mouse. *Japanese Journal of Pharmacology* 70, 85–88.
- Mayo, G.M., Langridge, P., 2003. Modes of reproduction in Australian populations of *Hypericum perforatum* L. (St. John's wort) revealed by DNA fingerprinting and cytological methods. *Genome/National Research Council Canada* 46, 573–579.
- Mcewen, J.G., Taylor, J.W., Carter, D., Xu, J., Felipe, M.S.S., Vilgalys, R., Mitchell, T.G., Kasuga, T., White, T., Bui, T., Soares, C.M.A., 2000. Molecular typing of pathogenic fungi. *Medical Mycology: Official publication of the International Society for Human and Animal Mycology* 38, 189–197.
- Morris, P.F., Connolly, M.S., St. Clair, D.A., 2000. Genetic diversity of *Alternaria alternata* isolated from tomato in California assessed using RAPDs. *Mycological Research* 104, 286–292.
- Nagata, S., 1997. Apoptosis by death factor. *Cell* 88, 355–365.
- Nagata, S., Nagase, H., Kawane, K., Mukae, N., Fukuyama, H., 2003. Degradation of chromosomal DNA during apoptosis. *Cell Death and Differentiation* 10, 108–116.

- Nakamura, K., Yamaguchi, Y., Kagota, S., Kwon, Y.M., Shinzuka, K., Kunitomo, M., 1999. Inhibitory effect of *Cordyceps sinensis* on spontaneous liver metastasis of Lewis lung carcinoma and B16 melanoma cells in syngeneic mice. *Japanese Journal of Pharmacology* 79, 335–341.
- Negi, M.S., Singh, A., Lakshmi Kumaran, M., 2000. Genetic variation and relationship among and within *Withania* species as revealed by AFLP markers. *Genome/National Research Council Canada* 43, 975–980.
- Ngan, F., Shaw, P., But, P., Wang, J., 1999. Molecular authentication of *Panax* species. *Phytochemistry* 50, 787–791.
- Nieri, P., Adinolfi, B., Morelli, I., Breschi, M.C., Simoni, G., Martinotti, E., 2003. Genetic characterization of the three medicinal *Echinacea* species using RAPD analysis. *Planta Medica* 69, 685–686.
- Nikoh, N., Fukatsu, T., 2000. Interkingdom host jumping underground: phylogenetic analysis of entomoparasitic fungi of the genus *Cordyceps*. *Molecular Biology and Evolution* 17, 629–638.
- Nikoh, N., Fukatsu, T., 2001. Evolutionary dynamics of multiple group I introns in nuclear ribosomal RNA genes of endoparasitic fungi of the genus *Cordyceps*. *Molecular Biology and Evolution* 18, 1631–1642.
- Obornik, M., Jirku, M., Dolezel, D., 2001. Phylogeny of mitosporic entomopathogenic fungi: is the genus *Paecilomyces* polyphyletic? *Canadian Journal of Microbiology* 47, 813–819.
- Park, J.-E., Kim, G.-Y., Park, H.-S., Nam, B.-H., An, W.-G., Cha, J.-H., Lee, T.-H., Lee, J.-D., 2001. Phylogenetic analysis of caterpillar fungi by comparing ITS 1-5.8S-ITS 2 ribosomal DNA sequences. *Mycobiology* 29, 121–131.
- Radimer, K.L., Subar, A.F., Thompson, F.E., 2000. Nonvitamin, non-mineral dietary supplements: issues and findings from NHANES III. *Journal of the American Dietetic Association* 100, 447–454.
- Ranamukhaarachchi, D.G., Kane, M.E., Guy, C.L., Li, Q.B., 2000. Modified AFLP technique for rapid genetic characterization in plants. *BioTechniques* 29, 858–866.
- Rehner, S.A., Samuels, G.J., 1995. Molecular systematics of the Hypocreales: a teleomorph gene phylogeny and the status of their anamorphs. *Canadian Journal of Botany* 73, S816–S823.
- Russo, A., Borrelli, F., Campisi, A., Acquaviva, R., Raciti, G., Vanella, A., 2003. Nitric oxide-related toxicity in cultured astrocytes: effect of *Bacopa monniera*. *Life Sciences* 73, 1517–1526.
- Sato, A., 1989. Studies on anti-tumor activity of crude drugs. I. The effects of aqueous extracts of some crude drugs in short-term screening test. *Yakugaku Zasshi. Journal of the Pharmaceutical Society of Japan* 109, 407–423.
- Saunders, J.A., Pedroni, M.J., Penrose, L.D.J., Fist, A.J., 2001. AFLP analysis of opium poppy. *Crop Science* 41, 1596–1601.
- Schmidt, H., Ehrmann, M., Vogel, R.F., Taniwaki, M.H., Niessen, L., 2003. Molecular typing of *Aspergillus ochraceus* and construction of species specific SCAR-primers based on AFLP. *Systematic and Applied Microbiology* 26, 138–146.
- Scorrano, L., Korsmeyer, S.J., 2003. Mechanisms of cytochrome c release by proapoptotic BCL-2 family members. *Biochemical and Biophysical Research Communications* 304, 437–444.
- Shahed, A.R., Kim, S.I., Shoskes, D.A., 2001. Down-regulation of apoptotic and inflammatory genes by *Cordyceps sinensis* extract in rat kidney following ischemia/reperfusion. *Transplantation Proceedings* 33, 2986–2987.
- Shiao, M.-S., Wang, Z.-N., Lin, L.-J., Lien, J.-Y., Wang, J.-J., 1994. Profiles of nucleosides and nitrogen bases in Chinese medicinal fungus *Cordyceps sinensis* and related species. *Botanical Bulletin of Academia Sinica New Series* 35, 261–267.
- Spatafora, J.W., Blackwell, M., 1993. Molecular systematics of unitunicate perithecial ascomycetes: the Clavicipitales-Hypocreales connection. *Mycologia* 85, 912–922.
- Starr, M., Post, T., Huus, K., 1993. *The Games China Plays*. Newsweek, 62–63.
- Steinkraus, D.C., Whitfield, J.B., 1994. Chinese Caterpillar Fungus and World Record Runners. *American Entomologist*. Winter, 235–239.
- Sugawara, T., Chan, P.H., 2003. Reactive oxygen radicals and pathogenesis of neuronal death after cerebral ischemia. *Antioxidants & Redox Signaling* 5, 597–607.
- Suh, S.-O., Noda, H., Blackwell, M., 2001. Insect symbiosis: derivation of yeast-like endosymbionts within an entomopathogenic filamentous lineage. *Molecular Biology and Evolution* 18, 995–1000.
- Suh, S.-O., Spatafora, J.W., Ochiel, G.R.S., Evans, H.C., Blackwell, M., 1998. Molecular phylogenetic study of a termite pathogen *Cordyceps pioideus bisporusogia*. *Mycologia* 90, 611–617.
- Sung, G.-H., Spatafora, J.W., Zare, R., Hodge, K.T., Gams, W., 2001. A revision of *Verticillium* sect. *Prostrata*, II. Phylogenetic analyses of SSU and LSU nuclear rDNA sequences from anamorphs and teleomorphs of the Clavicipitaceae. *Nova Hedwigia* 72, 311–328.
- Terashima, K., Matsumoto, T., Hasebe, K., Fukumasa-Nakai, Y., 2002. Genetic diversity and strain-typing in cultivated strains of *Lentinula edodes* (the shii-take mushroom) in Japan by AFLP analysis. *Mycological Research* 106, 34–39.
- Trubiani, O., Salvolini, E., Staffolani, R., Di Primio, R., Mazzanti, L., 2003. DMSO modifies structural and functional properties of RPMI-8402 cells by promoting programmed cell death. *International Journal of Immunopathology and Pharmacology* 16, 253–259.
- Vaux, D.L., Korsmeyer, S.J., 1999. Cell death in development. *Cell* 96, 245–254.
- Vos, P., Hogers, R., Bleeker, M., Reijers, M., Van De Lee, T., Hornes, M., Frijters, A., Pot, J., Peleman, J., Kulper, M., Zabeau, M., 1995. AFLP: a new technique for DNA fingerprinting. *Nucleic Acids Research* 23, 4407–4414.
- Wallner, E., Weising, K., Rompf, R., Kahl, G., Kopp, B., 1996. Oligonucleotide fingerprinting and RAPD analysis of *Achillea* species: characterization and long-term monitoring of micropropagated clones. *Plant Cell Reports* 15, 647–652.
- Wu, Z.-L., Wang, X.-X., Cheng, W.-Y., 2000. Inhibitory effect of *Cordyceps sinensis* and *Cordyceps militaris* on human glomerular mesangial cell proliferation induced by native LDL. *Cell Biochemistry and Function* 18, 93–97.
- Xu, R., Peng, X.E., Chen, G.Z., Chen, G.L., 1992. Effects of *Cordyceps sinensis* on natural killer activity and colony formation of B16 melanoma. *Chinese Medical Journal* 105, 97–101.
- Yamaguchi, Y., Kagota, S., Nakamura, K., Shinzuka, K., Kunitomo, M., 2000a. Antioxidant activity of the extracts from fruiting bodies of cultured *Cordyceps sinensis*. *Phytotherapy Research: PTR* 14, 647–649.
- Yamaguchi, Y., Kagota, S., Nakamura, K., Shinzuka, K., Kunitomo, M., 2000b. Inhibitory effects of water extracts from fruiting bodies of cultured *Cordyceps sinensis* on raised serum lipid peroxide levels and aortic cholesterol deposition in atherosclerotic mice. *Phytotherapy Research: PTR* 14, 650–652.
- Yang, L.Y., Chen, A., Kuo, Y.C., Lin, C.Y., 1999. Efficacy of a pure compound H1-A extracted from *Cordyceps sinensis* on autoimmune disease of MRL LPR/LPR mice. *The Journal of Laboratory and Clinical Medicine* 134, 492–500.
- Yang, L.Y., Huang, W.J., Hsieh, H.G., Lin, C.Y., 2003. H1-A extracted from *Cordyceps sinensis* suppresses the proliferation of human mesangial cells and promotes apoptosis, probably by inhibiting the tyrosine phosphorylation of Bcl-2 and Bcl-XL. *The Journal of Laboratory and Clinical Medicine* 141, 74–83.
- Yu, L., Chen, G.Z., Jiang, D.Z., 1993. Combined traditional Chinese and western medicine. Effect of *Cordyceps sinensis* on erythropoiesis in mouse bone marrow. *Chinese Medical Journal* 106, 313–316.
- Zang, Q., He, G., Zheng, Z., Liu, J., Wang, S., Huang, J., Du, D., Zeng, Q., Al, E., 1985. Pharmacological action of the polysaccharide from cordyceps (*Cordyceps sinensis*). *Zhong cao yao (Chinese Traditional and Herbal Drugs)* 16, 306–311.
- Zare, R., Gams, W., Culham, A., 2000. A revision of *Verticillium* sect. *Prostrata*, I. Phylogenetic studies using ITS sequences. *Nova Hedwigia* 71, 465–480.

- Zeller, K.A., Bowden, R.L., Leslie, J.F., 2003. Diversity of epidemic populations of *Gibberella zeae* from small quadrats in Kansas and North Dakota. *Phytopathology* 93, 874–880.
- Zerega, N.J.C., Mori, S., Lindqvist, C., Zheng, Q., Motley, T.J., 2002. Using amplified fragment length polymorphisms (AFLP) to identify black cohosh (*Actaea racemosa*). *Economic Botany* 56, 154–164.
- Zhang, K.Y.B., Leung, H.-W., Yeung, H.W., Wong, R.N.S., 2001. Differentiation of *Lycium barbarum* from its related *Lycium* species using random amplified polymorphic DNA. *Planta Medica* 67, 379–381.
- Zhang, Z.H.W., Liao, S., Li, J., Lei, L., Lui, J., Leng, F., Gong, W., Zhang, H., Wan, L., Wu, R., Li, S., Luo, H., Zhu, F., 1995. Clinical and laboratory studies of JinShuiBao in scavenging oxygen free radicals in elderly senescent XuZheng patients. *Journal of Administration Traditional Chinese Medicine*, 5.
- Zhao Long, W., Xiao Xia, W., Wei Ying, C., 2000. Inhibitory effect of *Cordyceps sinensis* and *Cordyceps militaris* on human glomerular mesangial cell proliferation induced by native LDL. *Cell Biochemistry and Function* 18, 93–97.
- Zheng, Z., Lee, J.E., Yenari, M.A., 2003. Stroke: molecular mechanisms and potential targets for treatment. *Current Molecular Medicine* 3, 361–372.
- Zhu, J.S., Gm, H., K.J., 1998a. The scientific rediscovery of an ancient Chinese herbal medicine: *Cordyceps sinensis*: part I. *Journal of Alternative and Complementary Medicine* (New York, NY). *The Journal of Alternative and Complementary Medicine: Research on Paradigm, Practice, and Policy* 4, 289–303.
- Zhu, J.S., Halpern, G.M., Jones, K., 1998b. The scientific rediscovery of a precious ancient Chinese herbal regimen: *Cordyceps sinensis*: part II. *Journal of Alternative and Complementary Medicine* (New York). *The Journal of Alternative and Complementary Medicine: Research on Paradigm, Practice, and Policy* 4, 429–457.