

# Susceptibility of New Zealand flora to *Phytophthora ramorum* and pathogen sporulation potential: an approach based on the precautionary principle

D. Hüberli<sup>A,B,D</sup>, B. Lutz<sup>A</sup>, B. Voss<sup>A</sup>, M. Calver<sup>B</sup>, M. Ormsby<sup>C</sup> and M. Garbelotto<sup>A</sup>

<sup>A</sup>Department of ESPM-ES, 137 Mulford Hall, University of California, Berkeley, CA 94720, USA.

<sup>B</sup>Present address: Centre for *Phytophthora* Science and Management, School of Biological Sciences and Biotechnology, Murdoch University, Murdoch, WA 6150, Australia.

<sup>C</sup>Biosecurity New Zealand, Ministry of Agriculture and Forestry, PO Box 2526, Wellington, New Zealand.

<sup>D</sup>Corresponding author. Email: D.Huberli@murdoch.edu.au

**Abstract.** *Phytophthora ramorum*, the cause of sudden oak death in the western USA and a damaging pathogen in Europe, is a biosecurity threat of unknown magnitude to New Zealand and Australasia because of its presence in traded ornamental plants. Knowledge of potential hosts acting as carriers and of symptoms caused by the pathogen on such hosts will strengthen precautionary quarantine regulations to prevent inadvertent introductions of *P. ramorum* into the region. Also, the identification of potential hosts will permit determination of areas at risk within countries that do not have *P. ramorum*. Susceptibility of New Zealand plants, including 17 endemic and three commercial species (*Eucalyptus globulus*, *Pinus radiata* and *Acacia melanoxylon*), as well as two known *Rhododendron* cultivar hosts, was determined by analysing the size of lesions on inoculated excised leaves and branches, while infectivity was determined by counting sporangia produced on leaves. In order to identify extremely susceptible hosts, seven species were inoculated using three concentrations of zoospores ranging from low ( $1 \times 10^2$  zoospores/mL) to high ( $5 \times 10^3$  zoospores/mL). In branch inoculations, *P. radiata* and *Nothofagus fusca* were as susceptible as the *Rhododendron* cultivars. *Pseudopanax arboreus*, *Fuchsia excorticata* and one *Rhododendron* cultivar were equally susceptible in leaf inoculations. However, *F. excorticata* was the only species with 100% infected leaves, high foliar sporulation and was highly susceptible at all three zoospore concentrations. *Leptospermum scoparium* was the only asymptomatic foliar host that had high reisolations of the pathogen. *F. excorticata*, *P. radiata*, *N. fusca*, *P. arboreus* and *L. scoparium* should be added to the potential host list for *P. ramorum* and monitored for symptoms and sporulation in gardens and nurseries in the USA and Europe. As part of a precautionary strategy, these species are suitable candidates for targeted surveillance programs in high-risk incursion areas of New Zealand. Furthermore, the sympatry of foliar hosts with high infectivity and of highly susceptible stem hosts was identified: these areas may be at risk for the development of a forest epidemic.

**Additional keyword:** oomycete.

## Introduction

Increased international trade in plants and plant products heightens the risk of introduction of alien plant pathogens (Levine and D'Antonio 2003). Once introduced, these pathogens cannot normally be eradicated, due to the fact that their recognition occurs years or decades after they have already become established and caused disease over significantly large zones of infestation (e.g. oomycete pathogens such as *Phytophthora ramorum*, *P. cinnamomi*, *P. kernoviae* and *P. lateralis*; Brasier 2007; Hansen 2008). International trade regulations are designed to prevent movement of known dangerous pathogens, but are of limited use in preventing the introduction of novel pathogens. Moreover, because extensive biosecurity measures may interfere with vast and profitable international trade in plants or plant products, quarantine measures must be carefully targeted to maximise protection at

a reasonable cost. The precautionary principle can be used as a guide in situations of high plausibility regarding the environmental risk associated with a potential pathogen introduction, but characterised by high uncertainty over the possible impacts. It states:

'Where there are threats of serious or irreversible damage, lack of full scientific certainty should not be used as a reason for postponing measures to prevent environmental degradation. In the application of the precautionary principle, public and private decisions should be guided by: (i) careful evaluation to avoid, wherever practicable, serious or irreversible damage to the environment; and, (ii) an assessment of the risk-weighted consequences of various options.' (The Intergovernmental Agreement on the

Environment, May 1992, quoted in Deville and Harding 1997, p. 13).

In applying the precautionary principle, it is necessary to establish if precautionary measures are needed and, if so, what measures are appropriate. Simultaneously, measures should be taken to reduce ignorance regarding both the risk and the extent of potential impact, in an attempt to move from precautionary to preventive actions (Deville and Harding 1997; for applications of the precautionary approach see Calver *et al.* 1999; Grayson and Calver 2004; Lilith *et al.* 2006).

The precautionary principle is particularly relevant to the possibility of the introduction of *P. ramorum* to Australasia, especially New Zealand. *Phytophthora ramorum* presents a significant and costly problem where it was recently introduced to western USA and Europe from an unknown origin (Werres *et al.* 2001; Rizzo *et al.* 2002) through the nursery trade (Ivors *et al.* 2006; Mascheretti *et al.* 2008). In Californian coastal forests, over 90 species are susceptible and the severity of the disease is mostly exemplified by the high levels of mortality of susceptible oaks (*Quercus agrifolia*, *Q. kelloggii*, *Q. parvula* var. *shrevei* and *Q. chrysolepis*) and of the related *Lithocarpus densiflora* (tanoak) (Rizzo *et al.* 2002; Murphy and Rizzo 2003). The epidemic is driven largely by *Umbellularia californica* (bay laurel), a host that supports abundant foliar sporulation (Davidson *et al.* 2005). The ability of *P. ramorum* to disseminate aerially differentiates it from most forest *Phytophthora* pathogens and, together with its ability to disperse in soil and water, places it in a category of extreme potential risk in any moist temperate climate. It infects many ornamental plants routinely traded on the international market (Werres *et al.* 2001; Tooley *et al.* 2004; APHIS 2008; RAPRA 2008) and it has been reported to sporulate on some species with limited or no symptoms (Bienapfl *et al.* 2005; Denman *et al.* 2006), making it a severe quarantine risk. In New Zealand, all but a few of the 27 plant families containing known hosts of *P. ramorum* in the USA and Europe have representative genera that are either indigenous or naturalised. One of the known hosts is indigenous (*Griselinia littoralis*) and eight are naturalised (*Drimys winterii*, *Frangula purshiana*, *Fraxinus excelsior*, *Pseudotsuga menziesii*, *P. undulatum*, *Sequoia sempervirens*, *Syringa vulgaris* and *Taxus baccata*) (Allan Herbarium 2007; RAPRA 2008). Overall, we believe that the demonstrated impacts of *P. ramorum* in the USA and Europe and the likelihood of susceptibility of New Zealand native flora argue for high levels of precaution.

One prompt and effective precautionary measure that could be applied to protect New Zealand is to identify susceptible New Zealand species. Knowledge of potential hosts acting as vectors of the pathogen may bolster quarantine guidelines to help prevent further inadvertent introductions of *P. ramorum* into uninfested countries. For example, the recent European isolations of *P. ramorum* from several tree species in the field, previously identified as potential hosts from artificial inoculations (Brasier *et al.* 2004), highlights the importance in identification of potential hosts before establishment. In this paper, the susceptibility levels of a broad range of New Zealand plant taxa to *P. ramorum* are compared with those of two known susceptible *Rhododendron* cultivars, using an excised inoculation test with leaves and branches. The potential of

each host species to support sporulation and thus be infectious was examined by counting sporangia produced after inoculation of detached leaves. Finally, a subset of species comprising five New Zealand species and two 'control' *Rhododendron* cultivars was exposed to three different levels of zoospore concentrations, to determine whether inoculum dilution may affect predicted susceptibility levels.

## Methods

### *Inoculation experiments*

Two inoculation experiments using 20 different plant species from New Zealand (endemic or commercial) were conducted concurrently using excised branches (Experiment 1) and leaves (Experiment 2) in July 2005 (Table 1). Two *Rhododendron* cultivars, Cunningham's White rhododendron and White Dwarf azalea, were included as positive controls in each inoculation, bringing the total number of plant species tested to 22. The two *Rhododendron* cultivars are known hosts to *P. ramorum*, with the azalea having somewhat lower susceptibility than the rhododendron (Steve Tjosvold, pers. comm.). In May 2006, a third experiment was conducted. This involved inoculations of leaves using three different zoospore concentrations on a subset of five New Zealand species from Experiment 2, and on the same two susceptible rhododendron cultivars used previously. This experiment was designed to identify plant species that may become infected even when pathogen inoculum may be low, a condition likely to occur initially when a pathogen has just been introduced in a new environment.

### *Phytophthora ramorum isolate*

Isolate PR-52 (CBS 110537; ATCC MYC-2436) of the A2 mating-type was used in all experiments. It was isolated from a *Rhododendron* in 2000 from a nursery in California and was shown to be highly pathogenic compared with other isolates in previous inoculation studies of two hosts (Hüberli *et al.* 2006a). For branch inoculations, inoculum discs (3-mm diameter) were cut with a sterile cork borer from the margin of 14-day-old cultures grown at 20°C on vegetable-8 juice (Campbell Soup Co., Davis, CA, USA) agar (V8A). For foliar inoculations, zoospores were produced as described in Hüberli *et al.* (2003). After haemocytometer counts, zoospore concentrations were adjusted to  $2 \times 10^4$  zoospores/mL in Experiment 2 and to three concentrations ( $1 \times 10^2$ ,  $1 \times 10^3$  and  $5 \times 10^3$  zoospores/mL) in Experiment 3. Prior to contact with zoospore solutions, laboratory-ware was acid washed (5 M hydrochloric acid) for 24 h and then washed three times with deionised water to reduce zoospore attraction to these surfaces so that zoospores remained in suspension.

### *Plant material*

Plants were selected to represent common species in New Zealand, either native to the insular nation or abundantly traded and imported into that country. For Experiments 1 and 2, branches of 25–30 cm in length, with 8–10 mature healthy leaves and no lateral branches, were collected in July 2005 from a

**Table 1. Disease parameters assessed on excised branches of 18 New Zealand (NZ) plant species (indigenous, exotic and commercial) and two *Rhododendron* cultivars (known susceptibles) wound inoculated with *Phytophthora ramorum* colonised agar discs and incubated in the greenhouse for 10 days**

Species are sorted by lesion area in ascending order

Species <sup>A</sup>	Family	Plant status in NZ	Plants ( <i>n</i> )	K-mean cluster groups <sup>B</sup>	Lesion length (mm) $\pm$ s.e. <sup>C</sup>	Symptomatic ratio	Pathogen reisolation	Branches girdled
<i>Brachyglottis repanda</i>	Asteraceae (Compositae)	Indigenous	1	1	2.3	3/4	3/4	0/4
<i>Dacrydium cupressinum</i>	Podocarpaceae	Indigenous	1	1	4.0	2/4	2/4	0/4
<i>Olearia bullata</i>	Asteraceae (Compositae)	Indigenous	1	1	4.0	3/4	1/4	0/4
<i>Dacrycarpus dacrydioides</i>	Podocarpaceae	Indigenous	2	1	4.3 $\pm$ 0.8	1/4	1.5/4	0/4
<i>Sophora microphylla</i>	Fabaceae (Leguminosae)	Indigenous	1	1	4.8	2/4	2/4	0/4
<i>Leptospermum scoparium</i>	Myrtaceae <sup>D</sup>	Indigenous	1	1	5.5	2/4	2/4	0/4
<i>Corynocarpus laevigatus</i>	Corynocarpaceae	Indigenous	1	1	6.0	4/4	3/4	0/4
<i>Prumnopitys spicata</i> <sup>E</sup>	Podocarpaceae	Indigenous	3	1	6.3 $\pm$ 3.7	3.5/4	3.5/4	0.5/4
<i>Nothofagus menziesii</i>	Nothofagaceae <sup>D</sup>	Indigenous	1	1	6.5	2/4	3/4	0/4
<i>Eucalyptus globulus</i>	Myrtaceae <sup>D</sup>	Exotic/ commercial	1	1	7.0	3/4	0/4	0/4
<i>Fuchsia excorticata</i>	Onagraceae	Indigenous	2	1	7.4 $\pm$ 4.0	3.5/4	2.5/4	0/4
<i>Coprosma robusta</i>	Rubiaceae	Indigenous	1	1	8.0	2/4	3/4	0/4
<i>Agathis australis</i>	Araucariaceae	Indigenous	2	1	8.5 $\pm$ 0.5	1/4	1.5/4	0/4
<i>Pseudopanax arboreus</i>	Araliaceae	Indigenous	1	1	14.3	3/4	2/4	1/4
<i>Acacia melanoxylon</i>	Fabaceae (Leguminosae)	Exotic/ commercial	1	1	15.0	2/4	4/4	0/4
<i>Pomaderris prunifolia</i>	Rhamnaceae <sup>D</sup>	Indigenous	1	1	17.5	2/4	4/4	0/4
<i>Pinus radiata</i>	Pinaceae <sup>D</sup>	Exotic/ commercial	1	2	22.8	4/4	3/4	1/4
<i>Nothofagus fusca</i>	Nothofagaceae <sup>D</sup>	Indigenous	1	2	26.6	4/4	3/4	3/4
<i>Rhododendron</i> cultivar (White Dwarf azalea)	Ericaceae <sup>D</sup>	Exotic	2	2	24.1 $\pm$ 5.6	2.5/4	3/4	1/4
<i>Rhododendron</i> cultivar (Cunningham's White rhododendron)	Ericaceae <sup>D</sup>	Exotic	2	2	51.1 $\pm$ 2.1	4/4	3.5/4	2.5/4

<sup>A</sup>*Pittosporum eugenioides* and *Melicytus ramiflorus* were tested in leaf inoculations, but not in branch inoculations.<sup>B</sup>K-mean cluster groups determined on all four disease parameters assessed; the variables lesion length, symptomatic ratio and branches girdled were significant ( $P < 0.02$ ).<sup>C</sup>Standard error presented only for species with more than one replicate plant.<sup>D</sup>Family containing at least one known host for *P. ramorum* (RAPRA 2008).<sup>E</sup>For *Prumnopitys spicata*, one plant of three was not infected and the pathogen was not reisolated; it was not included in the data presented.

total of 20 New Zealand plant species and the two positive controls (*Rhododendron* cultivars) growing at the University of California Botanical Garden in Berkeley or on the university campus (Tables 1 and 2). Two separate species of the 20 were not tested in Experiment 1 (*Melicytus ramiflorus* and *Pittosporum eugenioides*) and 2 (*Olearia bullata* and *Sophora microphylla*). Plant material was collected from one individual in most cases, or two to three individuals when these were available. All branches were kept in cooler-boxes with their basipetal-end in jars filled with sterilised water. One day after collection, branches (average 4-mm diameter) were trimmed to a length of 20 to 25 cm, with four to five leaves remaining, for inoculations in Experiment 1. Remaining branches provided the foliar material used for foliar inoculation in Experiment 2. Prior to inoculation, leaves were rinsed in sterile deionised water to remove any debris. In May 2006, plant material for Experiment 3 was collected and prepared for foliar inoculation, as described

above. Only a subset of five species was tested in this last experiment, namely *Dacrydium cupressinum*, *Fuchsia excorticata*, *Nothofagus fusca*, *Pinus radiata* and *P. eugenioides*. The same two *Rhododendron* cultivars used in Experiments 1 and 2 were inoculated as well, and served as controls.

#### Experiment 1: branch inoculation

##### Inoculation and test conditions

To prevent excessive evaporation during the experiment, jars were sealed by wrapping Parafilm (American National Can, Chicago, USA) around their mouths, with branches protruding through a hole in the Parafilm. Using a sterile syringe needle, a wound was created through the bark, 10 cm from the acropetal end of the branch. An inoculum disc was placed mycelium side down on the outside of the wound and the inoculation point was then

**Table 2. Disease parameters assessed on excised leaves of 18 New Zealand (NZ) plant species (indigenous, exotic and commercial) and two *Rhododendron* cultivars (known susceptibles) inoculated with zoospores *Phytophthora ramorum* and incubated for 6 days at 19°C**

Species are sorted by lesion area in ascending order

Species <sup>A</sup>	Family	Plant status in NZ	Plants (n)	K-mean cluster groups <sup>B</sup>	Lesion area (mm <sup>2</sup> ) ± s.e. <sup>C</sup>	Symptomatic ratio	Pathogen reisolation	Leaves with sporangia	Sporangia/leaf ± s.e. <sup>C</sup>
<i>Acacia melanoxylon</i>	Fabaceae (Leguminosae)	Exotic/commercial	1	1	0	0/14	0/14	— <sup>D</sup>	— <sup>D</sup>
<i>Agathis australis</i>	Araucariaceae	Indigenous	2	1	0	0/14	0/14	9/14	1.5 ± 0.5
<i>Brachyglottis repanda</i>	Asteraceae (Compositae)	Indigenous	1	1	0	0	2/14	5/14	1.8
<i>Corynocarpus laevigatus</i>	Corynocarpaceae	Indigenous	1	1	0	0	2/14	12/14	6.1
<i>Dacrycarpus dacrydioides</i>	Podocarpaceae	Indigenous	2	1	0	0/14	4/14	7/14	1.9 ± 0.3
<i>Eucalyptus globulus</i>	Myrtaceae <sup>E</sup>	Exotic/commercial	1	1	0	0/14	0/14	— <sup>D</sup>	— <sup>D</sup>
<i>Leptospermum scoparium</i>	Myrtaceae <sup>E</sup>	Indigenous	1	1	0	0/14	9/14	9/14	12.0
<i>Melicytus ramiflorus</i>	Violaceae	Indigenous	1	1	0	0	3/14	13/14	4.5
<i>Nothofagus fusca</i>	Nothofagaceae <sup>E</sup>	Indigenous	1	1	0	0	4/14	9/14	3.6
<i>Nothofagus menziesii</i>	Nothofagaceae <sup>E</sup>	Indigenous	1	1	0	0	2/14	8/14	2.2
<i>Pinus radiata</i>	Pinaceae <sup>E</sup>	Exotic/commercial	1	1	0	0/7	0/7	0/7	0.0
<i>Pomaderris prunifolia</i>	Rhamnaceae <sup>E</sup>	Indigenous	1	1	0	0	5/14	6/14	1.2
<i>Prumnopitys spicata</i>	Podocarpaceae	Indigenous	3	1	0	0	0.3/14	8.7/14	7.0 ± 2.7
<i>Dacrydium cupressinum</i>	Podocarpaceae	Indigenous	1	1	10.8	3/14	12/14	10/14	2.8
<i>Pittosporum eugenioides</i>	Pittosporaceae <sup>E</sup>	Indigenous	1	1	22.5	5/14	7/14	10/14	3.8
<i>Coprosma robusta</i>	Rubiaceae	Indigenous	1	1	39.9	2/14	2/14	12/14	1.6
<i>Fuchsia excorticata</i>	Onagraceae	Indigenous	2	2	81.5 ± 6.4	14/14	5.5/14	11/14	65.3 ± 59.7
<i>Pseudopanax arboreus</i>	Araliaceae	Indigenous	1	2	216.3	2/14	3/14	12/14	3.5
<i>Rhododendron</i> cultivar (White Dwarf azalea)	Ericaceae <sup>E</sup>	Exotic	2	1	32.7 ± 4.1	7/28	17/28	9/14	9.8 ± 6.8
<i>Rhododendron</i> cultivar (Cunningham's White rhododendron)	Ericaceae <sup>E</sup>	Exotic	2	2	119.1 ± 3.9	12.5/28	10.5/28	12.8/14	23.5 ± 10.2

<sup>A</sup>*Olearia bullata* and *Sophora microphylla* were tested in branch inoculations, but not in leaf inoculations.<sup>B</sup>K-mean cluster groups determined on all five disease parameters assessed; the variables lesion area, symptomatic ratio and sporangia/leaf were significant ( $P < 0.002$  in all cases).<sup>C</sup>SE presented only for species with more than one replicate plant.<sup>D</sup>No data.<sup>E</sup>Family containing at least one known host for *P. ramorum* (RAPRA 2008).

wrapped with a layer of Parafilm on top of which a layer of silver Nashua tape (Tyco Adhesives, Massachusetts, USA) was superimposed. For each individual plant, four replicate branches were inoculated. In total, material from 27 plants was used in this experiment (including the different genotypes of some plant species), resulting in 108 inoculated branches. Since most material was available only from the botanical garden, we were restricted in the number of branches we could remove from plants and hence replicate numbers were low. One branch from each individual plant was inoculated with a sterile V8A disc as a negative control. Branches in jars were arranged randomly in a misting chamber in a greenhouse with natural light of 12-h photoperiod, ambient temperatures (averages 16–22°C) and automatic misting three times daily for 30 min. The jars were given supplemental water as needed.

#### Measurements

Ten days after inoculation, the outer bark around the inoculation site was carefully scraped off with a scalpel and the entire lesion (if present) was exposed. Total lesion length

extending above and below the site of inoculation was measured and a note was made to indicate whether the lesion had girdled the entire inoculated stem. In order to determine survival of the pathogen in inoculated stems, small pieces of tissue from the margins of the longitudinal lesion and from the site of inoculation were cut and plated onto pimarcin–ampicillin–rifampicin–PCNB agar (P<sub>10</sub>ARP) selective medium, modified to contain one-quarter of the original amount of PCNB used by Erwin and Ribeiro (1996).

#### Experiments 2 and 3: foliar inoculations

##### Inoculation and test conditions

Shallow plastic trays (40 × 60 × 5 cm) were used to hold 1 L of zoospore inoculum, which filled the tray to an approx. depth of 5 mm. Fourteen leaves of each species were dipped simultaneously tip first into the inoculum at a concentration of 2 × 10<sup>4</sup> zoospores/mL for 5 min (Experiment 2). In order to do so, leaves were sandwiched between two Perspex strips (50 × 5 cm) held together with fold-back clips, the strips were then suspended

above the inoculum so that the leaf tips were immersed. The leaves from the different plants were placed randomly across the tray to account for minor differences in zoospore distribution. In Experiment 3, three sets of 14 leaves from each of the selected species were dipped into inoculum concentrations of  $1 \times 10^2$ ,  $1 \times 10^3$  and  $5 \times 10^3$  zoospores/mL. One control leaf for each species for both experiments was dipped in sterile deionised water. After inoculation, all leaves were removed from the Perspex strips and placed, adaxial side down, into trays containing moist paper towels. The trays were incubated in a growth chamber at 19°C for 6 days in darkness.

#### Measurements

After incubation, all leaves were scanned and the size of lesions was recorded using the image analysis software ASSESS V1.01 (APS Press, St Paul, MN, USA). Presence and absence of lesions was also recorded to determine the ratio of successful inoculations. Prior to scanning leaves in Experiment 2, the region of the inoculation was gently scraped with a flat spatula from both surfaces into individual wells of a microtiter plate containing 300 µL sterile deionised water. To each well, 10 µL of phenol-cotton blue was added. The number of sporangia per well was counted using a microscope at 40× magnification. Sporangia were not counted in Experiment 3.

After surface sterilisation of leaves with 70% ethanol, two segments from the lesion margin, or the region of inoculation if lesions were absent, were plated onto P<sub>10</sub>ARP to determine the pathogen reisolation proportion out of the total leaves inoculated.

#### Statistical analysis

All zero values for lesion length and area and sporangia counts were removed from the dataset before calculations of means and the statistical analysis. All statistical analyses were done using the Statistical software V6.1 (Statsoft Inc., Tulsa, OK). We used k-means clustering for branch and foliar inoculations and each of the three zoospore concentration foliar inoculations, to determine

whether any New Zealand plant species were equally susceptible to *P. ramorum* as the known highly susceptible *Rhododendron* cultivars. K-means clustering is a variation on conventional cluster analysis, in which the investigators designate the number of groups into which the cases (in this study, the plant species) will be clustered. We specified two clusters, seeking to force a classification into two groups, using all the disease variables assessed. In this case, the results are useful if one group contains one or both of the susceptible *Rhododendron* cultivars. Any other species classified with them can be regarded as similarly susceptible and thus a potential host of *P. ramorum*. The analysis also calculates an F-statistic comparing each variable between the two groups.

## Results

### Experiment 1: branch inoculation

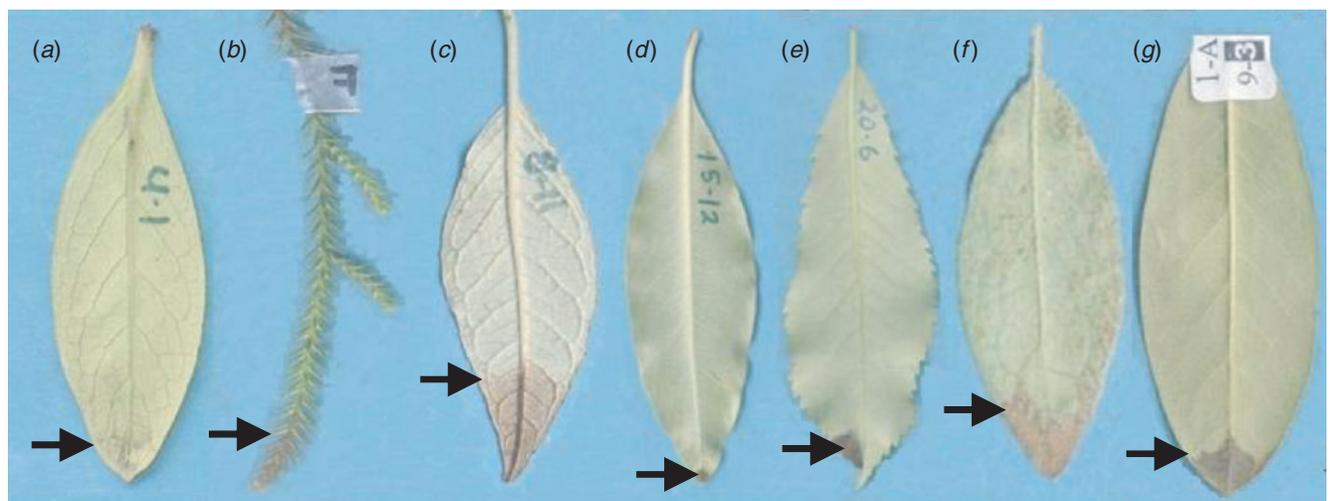
*Pinus radiata* and *N. fusca* had large lesions that were in the range of the two susceptible *Rhododendron* cultivars, and k-means clustering grouped these species together (Table 1). The remaining 16 New Zealand species had very low susceptibility to *P. ramorum*, in comparison to the susceptible *Rhododendron* cultivars, and were clustered into a second k-means clustering group (Table 1). Furthermore, the two groups were significantly ( $P < 0.02$  in all cases) different for lesion length, symptomatic ratio and branch girdling, but not for pathogen reisolation.

Girdling of branches by *P. ramorum* occurred in *Pseudopanax arboreus*, *N. fusca*, *P. radiata*, *Prumnopitys spicata* and *Rhododendron* cultivars (Table 1). Most severely affected were *N. fusca* and the *Rhododendron* cultivars.

### Foliar inoculations

#### Experiment 2: susceptibility and sporulation potential of 20 species

Five of the 18 New Zealand species formed foliar lesions when inoculated with *P. ramorum* zoospores (Table 2; Fig. 1).



**Fig. 1.** Leaves of (a) *Coprosma robusta*, (b) *Dacrydium cupressinum*, (c) *Fuchsia excorticata*, (d) *Pittosporum eugenioides*, (e) *Pseudopanax arboreus*, (f) *Rhododendron* cultivar (White Dwarf azalea) and (g) *Rhododendron* cultivar (Cunningham's White rhododendron) showing lesions (arrows) 6 days after incubation in the dark with zoospores of *Phytophthora ramorum*.

K-means clustering grouped the three species, *F. excorticata*, *P. arboreus* and Cunningham's White rhododendron, into one group. For this particular group, lesion area, symptomatic ratio and sporangia/leaf were significantly different ( $P < 0.002$ ) than equivalent values from the other species. *Fuchsia excorticata* was the only species among the three for which all leaves were infected; this species also supported the highest recorded foliar sporulation. Notably, one of the genotypes of *F. excorticata* produced twice as many sporangia as either *Rhododendron* cultivar. *P. arboreus* had the largest lesions, but only a few leaves (2 of 14) were infected and they both supported little sporulation. *Phytophthora ramorum* was recovered not only from all species with visible lesions, but also from nine New Zealand spp. that were asymptomatic (*Brachyglottis repanda*, *Corynocarpus laevigatus*, *Leptospermum scoparium*, *N. fusca*, *N. menziesii*, *Dacrycarpus dacrydioides*, *P. spicata*, *Pomaderris prunifolia*, *Melicytus ramiflorus*). In particular, *L. scoparium*, the species with the highest reisolation from asymptomatic leaves, was found to support sporulation at levels comparable to White Dwarf azalea (Table 2).

#### Experiment 3: susceptibility of seven species at three different inoculum concentrations

At the highest inoculum concentration ( $5 \times 10^3$  zoospores/mL), lesion size was significantly different among tested species ( $P < 0.05$ ) and k-means clustering grouped *F. excorticata* and White Dwarf azalea together (Fig. 2; Table 3). For the remaining four species (which had previously been identified as resistant) and Cunningham's White rhododendron, there was no detectable effect of inoculum concentration at the highest concentration. For *D. cupressinum* and *P. eugenoides*, disease incidence increased slightly at the higher inoculum levels.

Lowering the zoospore concentration below our standard inoculation concentration ( $1-2 \times 10^4$  zoospores/mL) did not change the high susceptibility of *F. excorticata* (Fig. 2). *Dacrydium cupressinum*, *P. radiata* and *P. eugenoides* always clustered together across the three zoospore concentrations. For both *Rhododendron* cultivars, susceptibility was lower when  $1 \times 10^2$  zoospores/mL was applied. In fact, they were clustered together with the other four species at the low zoospore concentration inoculation. At this concentration, lesions on *F. excorticata* were larger than those of any other species, including the two susceptible *Rhododendron* cultivars, and this species was clustered in its own group by k-means clustering analysis (Table 3).

Reisolation of *P. ramorum* increased with each successive increase in inoculum for all species except *P. radiata*, from which no reisolations were obtained (Fig. 2c). At the low inoculum concentrations, recovery was also low except for *F. excorticata*. *Phytophthora ramorum* was consistently isolated from ~40% of *F. excorticata* leaves for all three inoculations.

## Discussion

### Are precautionary measures needed?

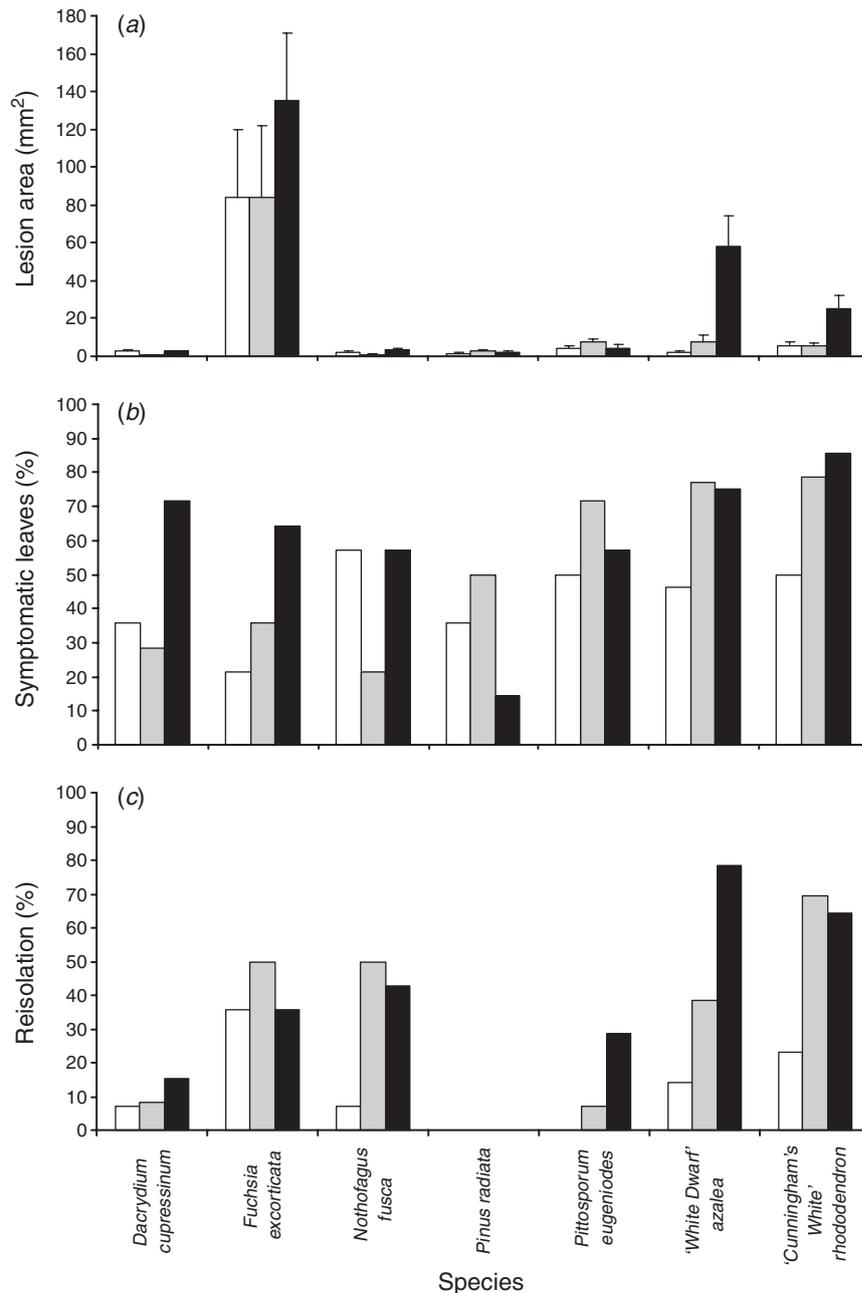
For an alien *Phytophthora* species to present a significant biosecurity problem, susceptible hosts are required, as well as favourable environmental conditions. We identified several potential hosts for *P. ramorum* from New Zealand, including

symptomatic foliar (*F. excorticata* and *P. arboreus*) and canker (*P. radiata* and *N. fusca*) hosts. Previous studies also found susceptibility of other *Fuchsia*, *Nothofagus* and *Pinus* spp. (Inman *et al.* 2002; RAPRA 2008). This expands the list of potential Australasian hosts for *P. ramorum*, highlighting the potential impacts of an incursion on the region (Hüberli *et al.* 2006b; RAPRA 2008).

Production of caducous sporangia on leaves of hosts such as *Umbellularia californica* and *Rhododendron* spp. drives the *P. ramorum* epidemic in the USA and European nurseries, respectively (Werres *et al.* 2001; Davidson *et al.* 2005). In our *in vitro* assay, at least one genotype of *F. excorticata* (the tree fuchsia) supported exceptionally high sporulation comparable to that produced by detached *U. californica* leaves (Hüberli *et al.* 2006b). *F. excorticata* is a tree (up to 15 m tall) common in lowland to lower montane forests throughout New Zealand, especially on forest margins. If inoculum dispersal is as liberal as it is for *U. californica*, also a tall forest tree, this New Zealand species could be the driving force of an epidemic similar to that observed in California. We have overlapped the occurrence of this particular species with *N. fusca*, a potential canker host (Fig. 3), to demonstrate areas that have the highest potential risk for development of an epidemic similar to sudden oak death. The area characterised by sympatry of these two species is quite extensive and highlights portions of New Zealand that are indeed at risk, were *P. ramorum* to be introduced. In California, it is the areas of sympatry between the infectious *U. californica* and the dead-end hosts (oaks or tanoaks) that have witnessed the most destructive levels of the sudden oak death epidemic (Rizzo and Garbelotto 2003). Clearly, the potential role of *F. excorticata* as a potential vector for *P. ramorum* should be investigated further because sporulation varied among the genotypes tested. It is also possible that susceptibility of hosts may vary between individuals grown in North America and individuals growing in their native land.

We reisolated *P. ramorum* from nine of 10 asymptomatic indigenous species after surface sterilisation. Sporangia were observed on all leaves, albeit in very low numbers, corroborating results from tests on UK and Mediterranean hosts (Denman *et al.* 2006). Of particular concern was *L. scoparium*, with 64% (9/14) reisolation from asymptomatic tissue. The global trade of plants and plant products places countries at extreme risk of inadvertent introductions of devastating pathogens, especially via unidentified hosts or asymptomatic hosts producing infectious inoculum. In New Zealand, the agricultural and horticultural industries rely on imported stock (Warmington *et al.* 1996). More than 13 000 vascular plants have been introduced into New Zealand, and almost 20% of these are now naturalised (Warmington *et al.* 1996). At such high importation levels, the chance of potentially infectious asymptomatic plant material entering into New Zealand is also much greater, further threatening the 2287 endemic terrestrial plants, of which 20 are listed as 'critical', 37 as 'endangered', 62 as 'vulnerable' and 79 as 'rare' (Cameron *et al.* 1995).

*Pinus radiata*, which was highly susceptible in wound branch inoculations, comprises ~89% of the planted production forest resource in New Zealand (Anon. 2006). Another planted production forest species in New Zealand is *P. menziesii*



**Fig. 2.** (a) Mean lesion area ( $\pm$ s.e.), (b) symptomatic leaves of total inoculated (%) and (c) pathogen reisolation (%) of *Phytophthora ramorum* on selective agar from excised leaves of seven plant species inoculated with three zoospore concentrations (white bars,  $1 \times 10^2$ ; grey bars,  $1 \times 10^3$ ; black bars,  $5 \times 10^3$  zoospores/mL) of *P. ramorum* after 6 days incubation at 19°C;  $n = 12-14$ .

(Douglas fir), also of the family Pinaceae, and this is a known host for *P. ramorum* (Davidson *et al.* 2003). The conifer, *D. cupressinum* (popular for quality furniture wood and an iconic indigenous species to New Zealanders), showed twig dieback in our foliar inoculation assay, with symptoms very similar to those described for *P. menziesii*. Reliable data on the susceptibility of these three species to *P. ramorum* and other alien *Phytophthora* spp. are crucial to New Zealand's timber industry,

as well as to regions with *P. ramorum* where these species may be indigenous or planted in large numbers.

Further screenings, using the methods employed here as well as other approaches (e.g. log inoculations described by Brasier and Jung 2006), are needed to identify plants and ecosystems at risk of infestation by *P. ramorum*. Caution should be used in extrapolating data to infection in natural conditions using *in vitro* tests, as such tests cannot fully reproduce natural conditions.

**Table 3. K-means clustering of five New Zealand species and two *Rhododendron* cultivars in foliar inoculations with *Phytophthora ramorum* zoospores at three different concentrations**

For each concentration, the table shows: cluster (which of two clusters the species belongs to), cluster means (the mean lesion area, symptomatic ratio and pathogen reisolation proportion) for the species in each cluster and analysis of variance statistics testing for differences in these disease variables between the two clusters at each concentration.

Species	Cluster <sup>A</sup>	Mean lesion area	Cluster means Symptomatic ratio	Pathogen reisolation proportion
<i>1 × 10<sup>2</sup> zoospores/mL</i>				
<i>Fuchsia excorticata</i>	1	84.2	21.4	35.7
<i>Dacrydium cupressinum</i>	2			
<i>Nothofagus fusca</i>	2			
<i>Pinus radiata</i>	2			
<i>Pittosporum eugenioides</i>	2			
<i>Rhododendron</i> cultivar (White Dwarf azalea)	2			
<i>Rhododendron</i> cultivar (Cunningham's White rhododendron)	2			
Mean (Cluster 2) =		2.9	45.8	8.6
Analysis of variance		$F_{(1,5)} = 2388.4; P < 0.0001$	$F_{(1,5)} = 6.9; P = 0.05$	$F_{(1,5)} = 8.0; P < 0.04$
<i>5 × 10<sup>2</sup> zoospores/mL</i>				
<i>Fuchsia excorticata</i>	1			
<i>Nothofagus fusca</i>	1			
<i>Rhododendron</i> cultivar (White Dwarf azalea)	1			
<i>Rhododendron</i> cultivar (Cunningham's White rhododendron)	1			
Mean (Cluster 1) =		24.6	53.2	51.9
<i>Dacrydium cupressinum</i>	2			
<i>Pinus radiata</i>	2			
<i>Pittosporum eugenioides</i>	2			
Mean (Cluster 2) =		3.6	50.0	5.2
Analysis of variance		$F_{(1,5)} = 0.80; P = 0.41$	$F_{(1,5)} = 0.02; P = 0.88$	$F_{(1,5)} = 35.4; P = 0.002$
<i>5 × 10<sup>3</sup> zoospores/mL</i>				
<i>Fuchsia excorticata</i>	1			
<i>Rhododendron</i> cultivar (White Dwarf azalea)	1			
Mean (Cluster 1) =		96.8	69.6	57.1
<i>Dacrydium cupressinum</i>	2			
<i>Nothofagus fusca</i>	2			
<i>Pinus radiata</i>	2			
<i>Pittosporum eugenioides</i>	2			
<i>Rhododendron</i> cultivar (Cunningham's White rhododendron)	2			
Mean (Cluster 2) =		7.3	57.1	30.2
Analysis of variance		$F_{(1,5)} = 17.1; P = 0.009$	$F_{(1,5)} = 0.38; P = 0.56$	$F_{(1,5)} = 1.5; P = 0.27$

<sup>A</sup>K-means clustering determined using lesion area, symptomatic ratio and pathogen reisolation proportion.

Although a study like the one described here is only a first step towards the identification of potential host and geographic ranges favourable to this pathogen in New Zealand, it does have the obvious advantage of avoiding any of the risks associated with importing the pathogen to New Zealand for experimentation. The number of New Zealand species and the number of genotypes per species available at U. C. Berkeley was obviously limited. Under these constraints, we propose caution regarding species that were negative in foliar and branch inoculations. Furthermore, those

species that were positive should be accepted as putative hosts pending a comprehensive study to determine whether resistance plays a role (see Tooley *et al.* 2004; Dodd *et al.* 2005; Hayden and Garbelotto 2005; Hüberli *et al.* 2006a). Nonetheless, the positive results obtained here are likely to be highly correlated with the genetic makeup of the tested species, due to the fact that all plants were from the U. C. Berkeley botanical garden and, hence, differential site effects on disease development were reduced to a minimum, as expected for a common garden experiment.

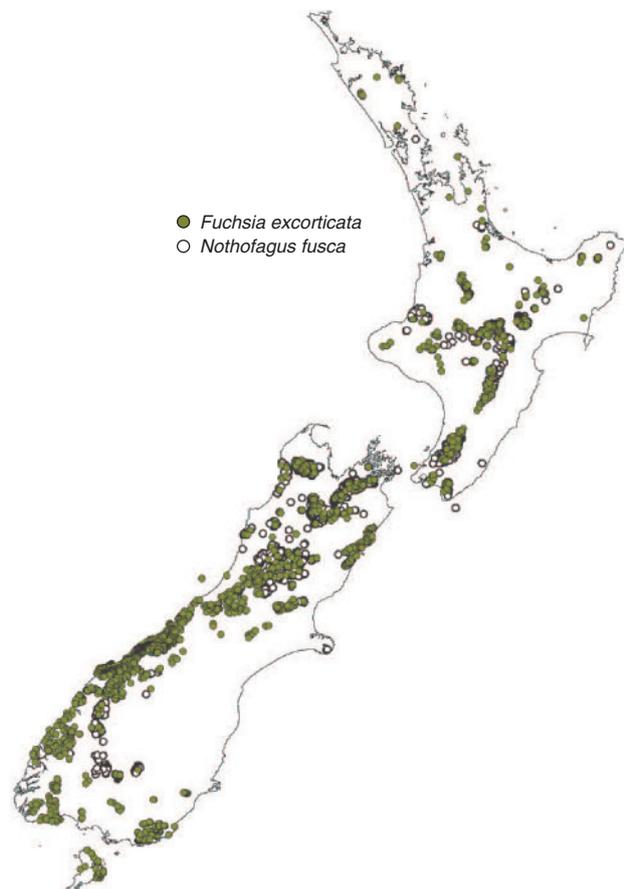


Fig. 3. Distribution of *Nothofagus fusca* and *Fuchsia excorticata* in New Zealand.

One approach we adopted to bypass the issue of artificial results caused by artificial conditions in the laboratory, was that of checking which plant species were susceptible *in vitro* when using low, but maybe more realistic, zoospore concentrations. At low inoculum concentrations, disease dropped markedly for all species including the known susceptible *Rhododendron* cultivars, but not for *F. excorticata*. Results from the low inoculum concentration inoculation thus identified *F. excorticata* as extremely susceptible. At higher inoculum, *F. excorticata* even surpassed the susceptible *Rhododendron* cultivars in susceptibility. Since disease was unrelated to inoculum load for *F. excorticata*, this potential host is probably extremely susceptible and we recommend it is used as an indicator for early detection of *P. ramorum* infection in high risk areas of New Zealand.

Previous studies found that detached foliar tests are often affected by host physiological factors, including leaf age and position in the canopy (Denman *et al.* 2005; Hansen *et al.* 2005). In our foliar assay, we only used mature leaves as these are less susceptible than juvenile leaves (Denman *et al.* 2005; Hansen *et al.* 2005). Additionally, leaves were always chosen from the sunny side of a plant, where the potential for epidermis formation was highest. Incubation in darkness in our foliar assay was

unnatural and may have contributed to lower resistance because leaves could not photosynthesise as stomatal opening was affected (see Tooley *et al.* 2004). Detached branch inoculations are widely used to assess susceptibility (Dodd *et al.* 2005). Despite the artificial nature of our inoculation assays, we showed susceptibility among species tested in excised branches and leaves. When available, whole plants exposed to natural inoculum in the field are the best substitute for detached leaves or branches, as they provide holistic plant responses.

All our inoculations used the A2 mating type isolate, Pr-52. We chose this isolate because it had a known pathogenicity from our previous studies (Hüberli *et al.* 2006a) and also for comparability to other studies that used this isolate on several hosts with differing inoculation methods (Rizzo *et al.* 2002; Tooley *et al.* 2004). Whilst quarantine regulations prohibited using the A1 isolate in our inoculations, other foliar inoculation studies using both A1 and A2 mating types found no statistical difference in aggressiveness amongst isolates (Tooley *et al.* 2004; Denman *et al.* 2005). Clear differences in aggressiveness amongst the A1 and A2 mating type isolates occurred in log inoculations (Brasier 2003) but, as far as we are aware, never in foliar inoculations.

Overall, while our experiments do not definitively establish the extent of susceptibility of the New Zealand flora to *P. ramorum*, they do show clearly the potential for the spread of any introduction, with possibly severe economic and conservation consequences. This argues for strong precautionary measures.

#### What precautionary measures should be applied?

New potential hosts were identified in this study. These should be included in monitoring programs not only in New Zealand, but in all places where *P. ramorum* is regarded as a dangerous pathogen and regulated. Given some plants display no symptoms, the presence of *P. ramorum* should be screened, not only by surveying for symptoms, but also by destructive sampling of a small number of plants to be assessed using the most sensitive available DNA-based assays (for instance Hayden *et al.* 2006 or Hughes *et al.* 2006). In addition, soil baiting of a subset of remaining plants could be used to identify the presence of the pathogen. Intensive screening of potential Australian hosts to *P. ramorum* is underway to identify hosts with possible high sporulation potential and to develop incursion risk models (Hüberli *et al.* in press).

Our current study lists potential hosts, both foliar and stem, which may place New Zealand's native forest and the commercial plantations at risk of a *P. ramorum* incursion. While few indigenous plants were tested for susceptibility to *P. ramorum*, 12% (2/17) of them showed substantial susceptibility. Areas where *F. excorticata* and *N. fusca* are sympatric were further identified as zones extremely favourable to invasion by this pathogen. Were this pathogen to be introduced in New Zealand, it is these areas that could witness the development of a forest epidemic comparable to sudden oak death in California.

More species should be screened to accurately assess the risk posed to New Zealand and the movement of these foreign plant species within USA and Europe. Until such time, in accord with

the precautionary principle, it is prudent to implement strict quarantine measures in an attempt to prevent an accidental introduction of *P. ramorum*.

### Acknowledgements

We thank T. Harnik, K. Maeda and A. Smith for technical assistance; G. Hardy, K. Howard and K. Ireland for comments and assistance with the manuscript; and two anonymous reviewers who made this paper stronger. Most plant material was kindly supplied by the University of California Botanical Garden at Berkeley. We acknowledge the use of Fig. 3, drawn from the National Vegetation Survey Database (Manaaki Whenua, Landcare Research New Zealand Ltd 2004). The project was funded by Biosecurity New Zealand, the Betty and Gordon Moore Foundation and the Pacific Southwest Research Station USDA Forest Service.

### References

- Allan Herbarium (2007) 'Ngā Tipu o Aotearoa – New Zealand Plant Names Database.' (Landcare Research: New Zealand) Available at: <http://nzflora.landcareresearch.co.nz> [Verified 27 August 2008].
- Anon. (2006) 'New Zealand forest industry facts and figures: 2005/2006.' (Ministry of Agriculture and Forestry: Wellington, New Zealand)
- APHIS (2008) 'APHIS listed of regulated hosts and plants associated with *Phytophthora ramorum*.' (Animal and Plant Health Inspection Service: Riverdale, MD) Available at [http://www.aphis.usda.gov/plant\\_health/plant\\_pest\\_info/pram/downloads/pdf\\_files/usdaprlst.pdf](http://www.aphis.usda.gov/plant_health/plant_pest_info/pram/downloads/pdf_files/usdaprlst.pdf) [Verified 27 August 2008].
- Bienapfl J, Zanzot J, Murphy S, Garbelotto M, Rizzo D (2005) Isolation of a new lineage of *Phytophthora ramorum* from asymptomatic stems and roots of a commercial lot of rhododendron in California. *Phytopathology* **95**, S9.
- Brasier C (2008) *Phytophthora ramorum* + *P. kernoviae* = international biosecurity failure. In 'Proceedings of the sudden oak death third science symposium, Santa Rosa, California, US, March 2007. Tech. Rep. PSW-GTR-214'. (Tech. Coords SJ Frankel, JT Kliejunas, KM Palmieri) pp. 133–139. (Department of Agriculture, Forest Service, Pacific Southwest Research Station: Albany, CA)
- Brasier C, Jung T (2006) Recent developments in *Phytophthora* diseases of trees and natural ecosystems in Europe. In 'Progress in research on *Phytophthora* diseases of forest trees. Proceedings of third international IUFRO working party S07.02.09 meeting at Freising, Germany'. (Eds C Brasier, T Jung, W Oßwald) pp. 5–16. (Forest Research: Farnham, UK)
- Brasier C, Denman S, Brown A, Webber J (2004) Sudden oak death (*Phytophthora ramorum*) discovered on trees in Europe. *Mycological Research* **108**, 1108–1110. doi: 10.1017/S0953756204221244
- Brasier CM (2003) Sudden oak death: *Phytophthora ramorum* exhibits transatlantic differences. *Mycological Research* **107**, 258–259. doi: 10.1017/S0953756203227660
- Calver MC, Bradley JS, Wright IW (1999) Towards scientific contributions in applying the precautionary principle: an example from southwestern Australia. *Pacific Conservation Biology* **5**, 63–72.
- Cameron EK, de Lange PJ, Given DR, Johnson PN, Ogle CC (1995) New Zealand Botanical Society threatened and local plant lists (1995 revision). *Newsletter of the New Zealand Botanical Society* **32**, 15–28.
- Davidson JM, Werres S, Garbelotto M, Hansen EM, Rizzo DM (2003) Sudden oak death and associated diseases caused by *Phytophthora ramorum*. *Plant Health Progress*. doi: 10.1094/PHP-2003-0707-01-DG
- Davidson JM, Wickland AC, Patterson HA, Falk KR, Rizzo DM (2005) Transmission of *Phytophthora ramorum* in mixed-evergreen forest in California. *Phytopathology* **95**, 587–596. doi: 10.1094/PHYTO-95-0587
- Denman S, Kirk SA, Brasier CM, Webber JF (2005) *In vitro* leaf inoculation studies as an indication of tree foliage susceptibility to *Phytophthora ramorum* in the UK. *Plant Pathology* **54**, 512–521. doi: 10.1111/j.1365-3059.2005.01243.x
- Denman S, Moralejo E, Kirk S, Orton E, Webber J (2006) Sporulation of *Phytophthora ramorum* and *P. kernoviae* on asymptomatic foliage. *Phytopathology* **96**, S28.
- Deville A, Harding R (1997) 'Applying the precautionary principle.' (The Federation Press: Sydney)
- Dodd RS, Hüberli D, Douhovnikoff V, Harnik TY, Afzal-Rafii Z, Garbelotto M (2005) Is variation in susceptibility to *Phytophthora ramorum* correlated with population genetic structure in coast live oak (*Quercus agrifolia*)? *The New Phytologist* **165**, 203–214. doi: 10.1111/j.1469-8137.2004.01200.x
- Erwin DC, Ribeiro OK (1996) '*Phytophthora* diseases worldwide.' (APS Press: St Paul, MN)
- Grayson J, Calver MC (2004) Regulation of domestic cat ownership to protect urban wildlife: a justification based on the precautionary principle. In 'Urban wildlife: more than meets the eye'. (Eds D Lunney, S Burgin) pp. 169–178. (Royal Zoological Society of New South Wales: Mosman, NSW)
- Hansen EM (2008) Alien forest pathogens: *Phytophthora* species are changing world forests. *Boreal Environment Research* **13**, 33–41.
- Hansen EM, Parke JL, Sutton W (2005) Susceptibility of Oregon forest trees and shrubs to *Phytophthora ramorum*: a comparison of artificial inoculation and natural infection. *Plant Disease* **89**, 63–70. doi: 10.1094/PD-89-0063
- Hayden K, Garbelotto M (2005) Variation in susceptibility of tanoak to sudden oak death at the population and species levels. *Phytopathology* **95**, S41.
- Hayden K, Ivors K, Wilkinson C, Garbelotto M (2006) Taqman chemistry for *Phytophthora ramorum* detection and quantification, with a comparison of diagnostic methods. *Phytopathology* **96**, 846–854. doi: 10.1094/PHYTO-96-0846
- Hüberli D, Van Sant-Glass W, Tse JG, Garbelotto M (2003) First report of foliar infection of starflower by *Phytophthora ramorum*. *Plant Disease* **87**, 599. doi: 10.1094/PDIS.2003.87.5.599B
- Hüberli D, Afzal-Rafii Z, Dodd RS, Douhovnikoff V, Harnik TY, Meshriy M, Miles L, Reuther K, Garbelotto M (2006a) Interactions of *Phytophthora ramorum* with two native Californian trees: bay laurel and coast live oak. In 'Progress in research on *Phytophthora* diseases of forest trees. Proceedings of third international IUFRO working party S07.02.09 meeting at Freising, Germany'. (Eds C Brasier, T Jung, W Oßwald) pp. 79–83. (Forest Research: Farnham, UK)
- Hüberli D, Wilkinson C, Smith MA, Meshriy M, Harnik TY, Garbelotto M (2006b) *Pittosporum undulatum* is a potential Australian host of *Phytophthora ramorum*. *Australasian Plant Disease Notes* **1**, 19–21. doi: 10.1071/DN06009
- Hüberli D, Ireland K, Smith I, Dell B, Ormsby M, Rizzo D, Garbelotto M, Hardy G (In press) Australasia is at high risk of a *Phytophthora ramorum* epidemic. In 'Progress in research on *Phytophthora* diseases of forest trees. Proceedings of fourth international IUFRO working party S07.02.09 meeting at Monterey, California'. (Eds E Goheen, S Frankel, K Palmieri) (USDA Forest Service, Pacific Southwest Research Station: Albany, CA)
- Hughes KJD, Tomlinson JA, Griffin RL, Boonham N, Inman AJ, Lane CR (2006) Development of a one-step real-time polymerase chain reaction assay for diagnosis of *Phytophthora ramorum*. *Phytopathology* **96**, 975–981. doi: 10.1094/PHYTO-96-0975
- Inman AJ, Beales PA, Lane CR, Brasier CM (2002) Comparative pathogenicity of European and American isolates of *Phytophthora ramorum* to leaves of ornamental, hedgerow and woodland under-story plants in the UK. In 'Sudden oak death I science symposium, Monterey, CA'. p. 80. (Poster abstract)

- Ivors KI, Garbelotto M, Vries IDE, Ruyter-Spira C, Hekkert BTE, Rosenzweig N, Bonants P (2006) Microsatellite markers identify three lineages of *Phytophthora ramorum* in US nurseries, yet single lineages in US forest and European nursery populations. *Molecular Ecology* **15**, 1493–1505. doi: 10.1111/j.1365-294X.2006.02864.x
- Levine JM, D'Antonio CM (2003) Forecasting biological invasion with increasing international trade. *Conservation Biology* **17**, 322–326. doi: 10.1046/j.1523-1739.2003.02038.x
- Lilith M, Calver MC, Styles I, Garkaklis MJ (2006) Protecting wildlife from predation by owned domestic cats: application of a precautionary approach to the acceptability of proposed cat regulations. *Austral Ecology* **31**, 176–189. doi: 10.1111/j.1442-9993.2006.01582.x
- Mascheretti S, Croucher PJP, Vettriano A, Prospero S, Garbelotto M (2008) Reconstruction of the sudden oak death epidemic in California through microsatellite analysis of the pathogen *Phytophthora ramorum*. *Molecular Ecology* **17**, 2755–2768. doi: 10.1111/j.1365-294X.2008.03773.x
- Murphy SK, Rizzo DM (2003) First report of *Phytophthora ramorum* on canyon live oak in California. *Plant Disease* **87**, 315. doi: 10.1094/PDIS.2003.87.3.315C
- RAPRA (2008) Database of naturally infected hosts for *Phytophthora ramorum*. In 'Risk analysis for *Phytophthora ramorum*'. Available at <http://rapra.csl.gov.uk/objectives/wp1/naturalhostsearch.cfm/> [Verified 27 August 2008].
- Rizzo DM, Garbelotto M (2003) Sudden oak death: endangering California and Oregon forest ecosystems. *Frontiers in Ecology and the Environment* **5**, 197–204.
- Rizzo DM, Garbelotto M, Davidson JM, Slaughter GW, Koike ST (2002) *Phytophthora ramorum* as the cause of extensive mortality of *Quercus* spp. and *Lithocarpus densiflorus* in California. *Plant Disease* **86**, 205–214. doi: 10.1094/PDIS.2002.86.3.205
- Tooley PW, Kyde KL, Englander L (2004) Susceptibility of selected Ericaceous ornamental host species to *Phytophthora ramorum*. *Plant Disease* **88**, 993–999. doi: 10.1094/PDIS.2004.88.9.993
- Warmington B, Cole G, King G (1996) New Zealand: country report. In 'FAO international technical conference on plant genetic resources, Leipzig, Germany'. 82 pp. Available at <http://193.43.36.103/ag/agp/AGPS/pgrfal/pdf/NEWZELAN.pdf> [Verified 10 September 2008].
- Werres S, Marwitz R, Man In 'T, Veld WA, De Cock AWAM, Bonants PJM, De Weerd M, Themann K, Ilieva E, Baayen RP (2001) *Phytophthora ramorum* sp. nov., a new pathogen on *Rhododendron* and *Viburnum*. *Mycological Research* **105**, 1155–1165. doi: 10.1016/S0953-7562(08)61986-3

Manuscript received 14 May 2007, accepted 29 July 2008