How can we differentiate species within the major evolutionary clades of Phytophthora? A focus on morphology.

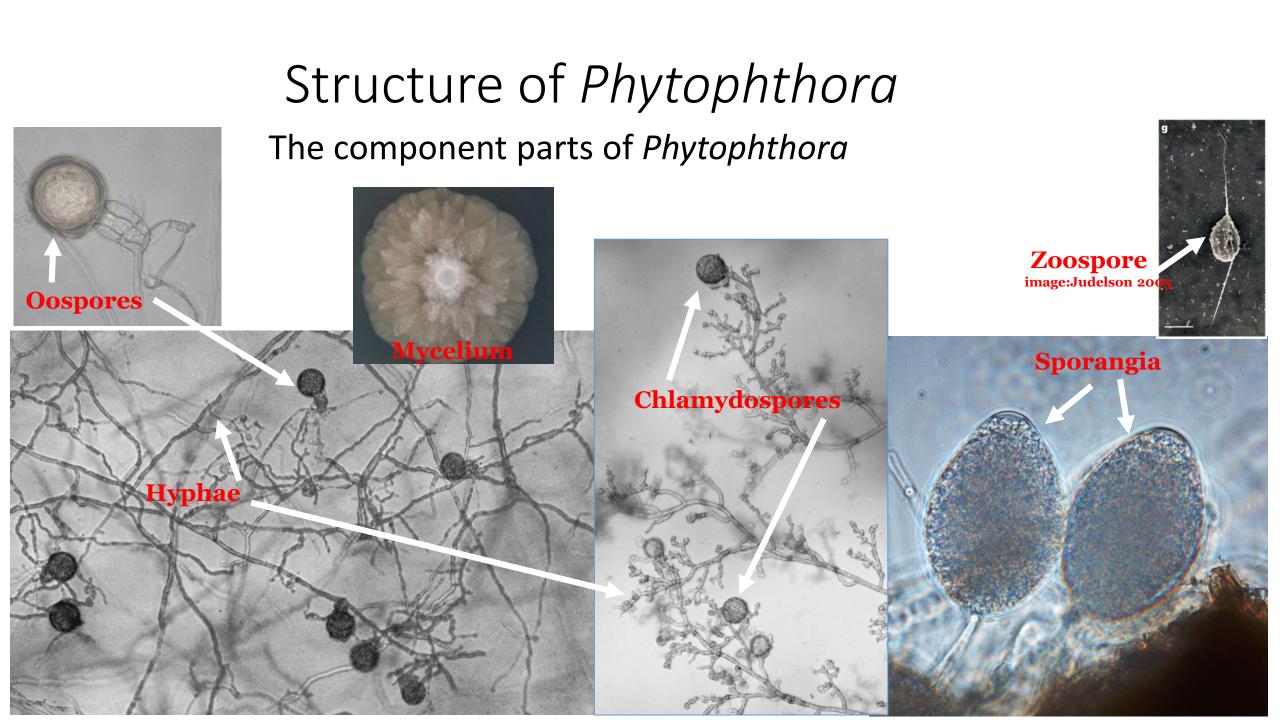
Laura Sims

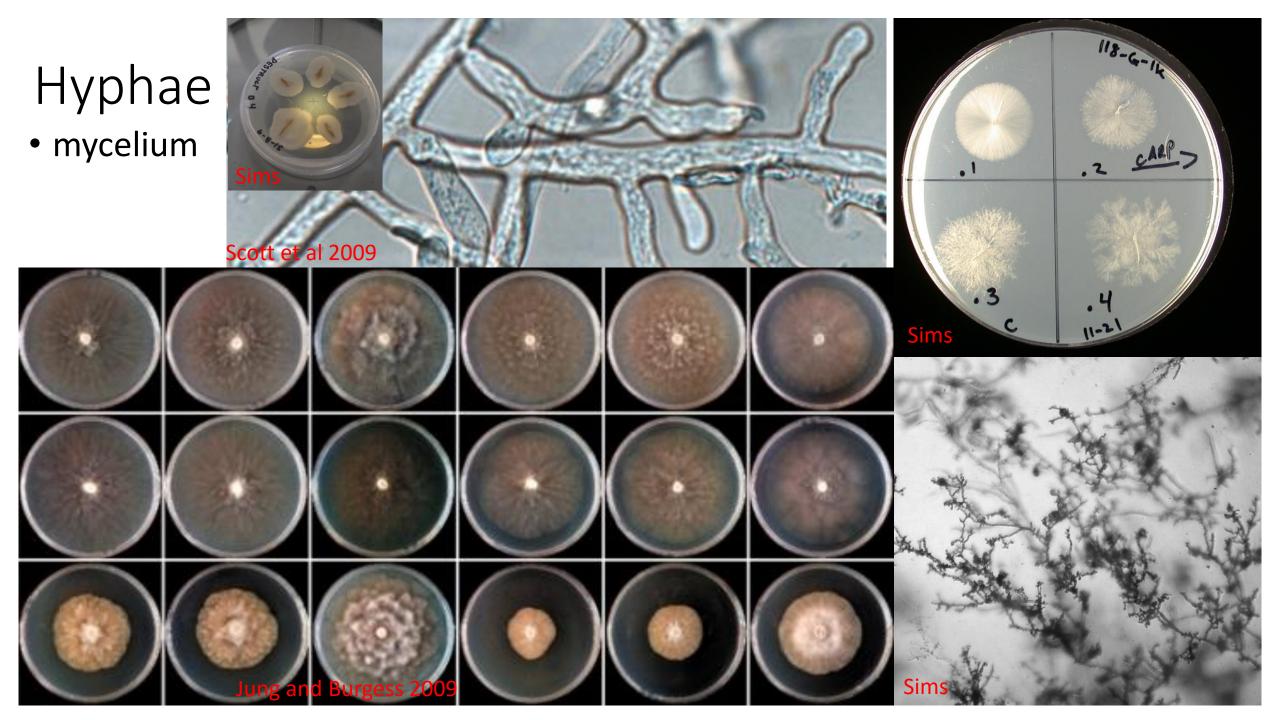
Three pathways for a complete ID

- Genetic looking at gene regions to match to a known species type
- Physiological- comparing growth rates, growth optimum temperatures, and growth limiting temperatures
- Morphological –explained herein, in practice used in some combination with genetic and physiological characters to definatively determine species

Guides to recognizing species based on morphology

- Tucker (1931) and Leonian (1934) seperately developed the early taxonomic keys for IDing *Phytophthora*
- Waterhouse developed detailed descriptions (1950s) and the most widely used keys (1963, 1978, Stamps et al. 1990) in the mid-late 20th Century
- Erwin and Ribiero (1996) produces a guidebook on Phytophthora <u>Phytophthora Diseases Worldwide</u> with morphological descriptions
- Gallegly and Hong (2008) key with some morphology and the use of SSCP DNA fingerprinting of 59 *Phytophthora* species
- Ristaino (2012) and has a modern key to species that combines 20 morphological characters with genetic sequences, and includes 59 *Phytophthora* species





Hyphal Characteristics

- ~ 90 ° angle branch system
- aseptate (lacking cell walls)
- Pinched at branch points
- Hyphae refracts light well

~ 90 ° angle branch system

easy to see on

Phytophthora

selective

medium

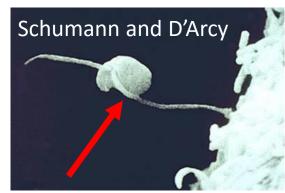
Sims

pinched

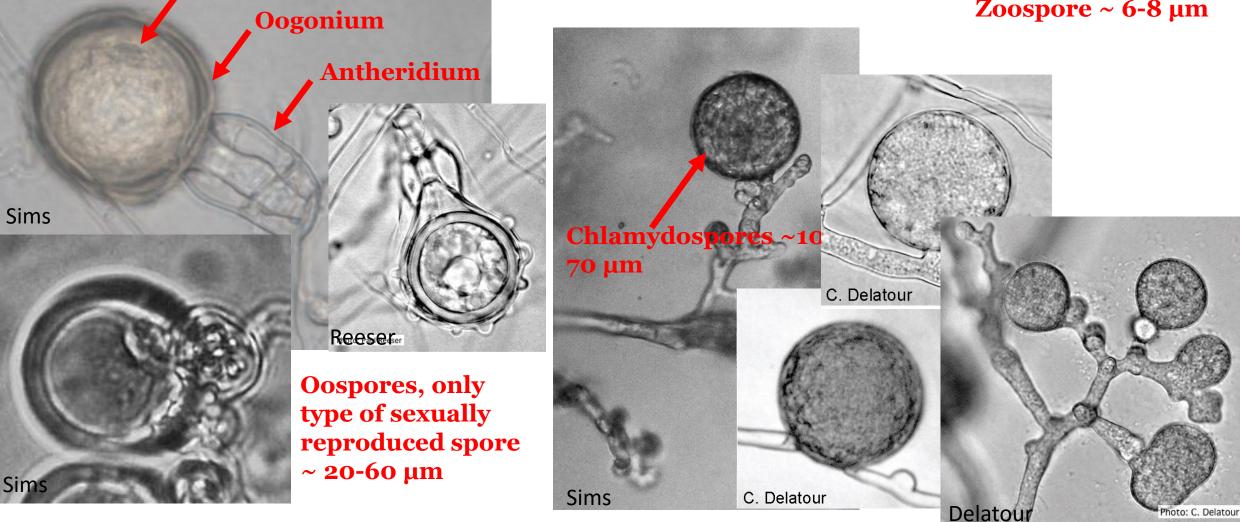
pinched

Phytophthora Spore Types

Oospore

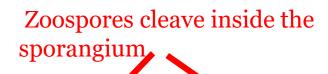


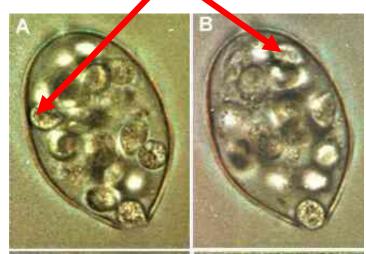
Zoospore ~ 6-8 μm

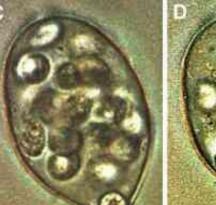


Sporangium

spore bearing structure







Jean Deacon

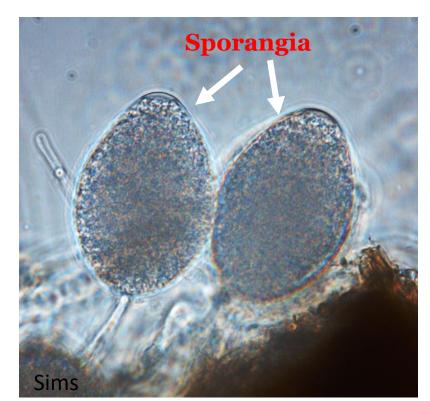
Zoospores

release

Biflagellate Zoospores are released from a sporangium



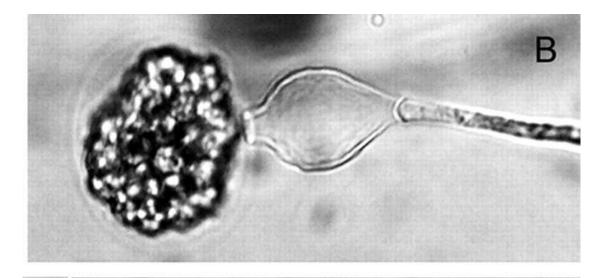
Schumann and D'Arcy SEM



Sims

Seperating closely related genera

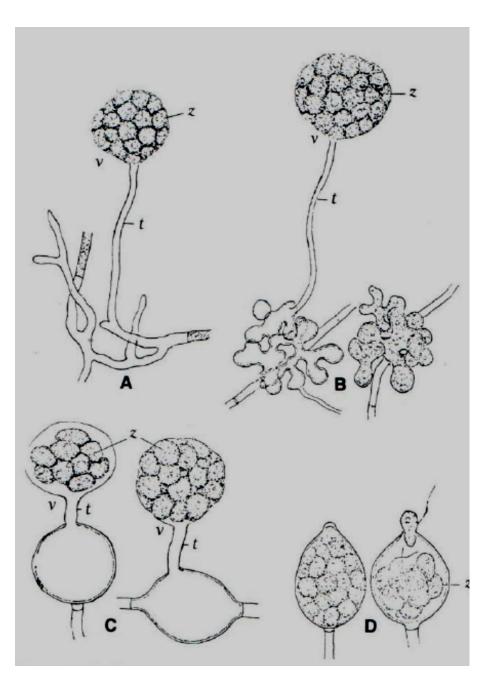
Pythium vs. Phytophthora



Mycologia 2009 Pythium delawarii

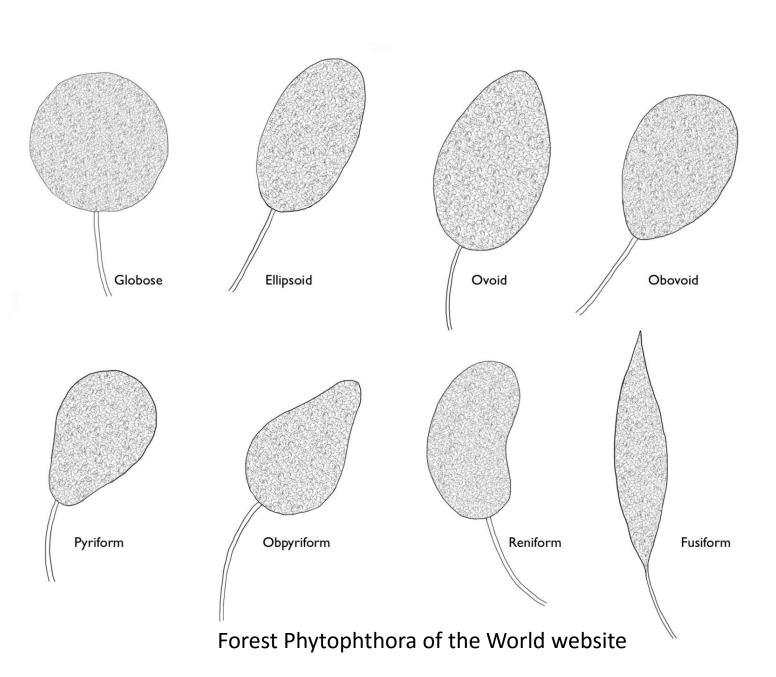


Persoonia (2009), Phytophthora plurivora



From Erwin and Ribiero 1996

Shapes of sporangia, and spore shapes



To what level does *Phytophthora* structure inform us about the genus and species?

- Hyphae- genus and species
- Mycelium- genus and species
- Sporangia- genus and species
- Chlamydospores- genus and species
- Oospores- genus and species
- Sexual state- genus and species
- Zoospores- genus

How do we use *Phytophthora* structure inform us about the genus and species?

To ID *Phytophthora* we look at the characteristics of the components **Hyphae** – hyphal swelling , aerial or appressed, regular, irregular **Mycelium-** colony pattern: petaloid, irregular, arachnoid, fluffy

Sporangia- attachment location, ability to separate from sporangiophore, papilation, proliferation type, biometric measurements and shape

Chlamydospores- wall thickness, placement on hyphae, and size

Oospores- size, shape, presence or absence, wall thickness, centeredness, oogonial ornamentation, antheridial attachment

Sexual state- thallism, sterility

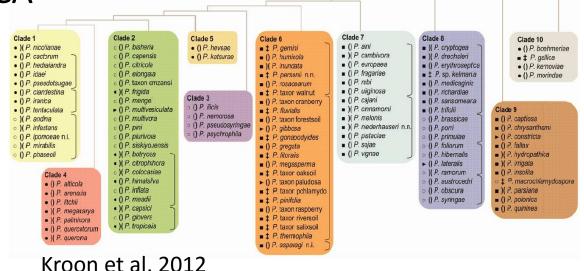
Zoospores- presence or absence and where formed

How to recognize species from your area

- Wherever you are there is a finite number of *Phytophthora* pathogens
- Familiarizing yourself (i.e. establishing a baseline) with the *Phytophthora* species from your crop-plant species of interest which provides a limit of *Phytophthora* to expect.
- Root-rotting *Phytophthora* will be more diverse on a single host than foliar or stem canker *Phytophthora*
- Consider the differences that help to distinguish closely related species
- Consider the differences that help to distinguish more distant species
- Aware of the limitations of using morphology

Evolutionary clades in *Phytophthora*

- Genus can be divide into 10 groups; Clades 1-10
- Groups that are important in wildlands in California, are in clades 1, 2, 3, 6, 7, and 8
- Go through distinguishing characters of two species from each of these clades that are important in CA

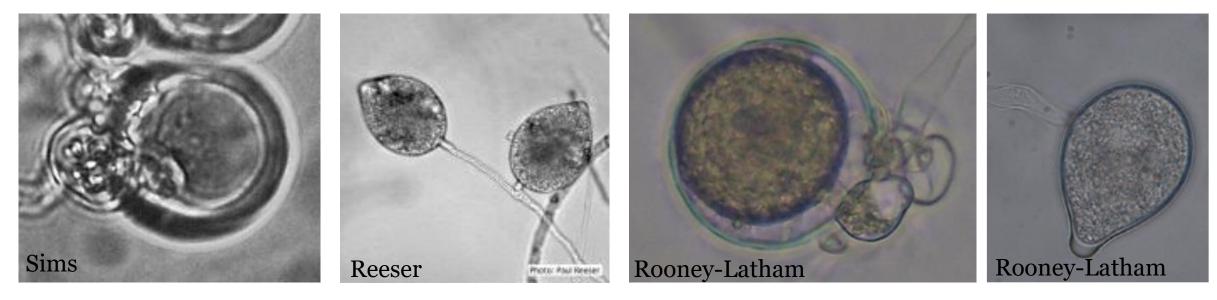


Species pairs within Evolutionary clades: Clade 1

• P. cactorum

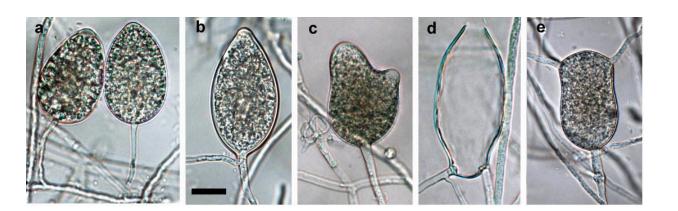
VS.

P. tentaculata



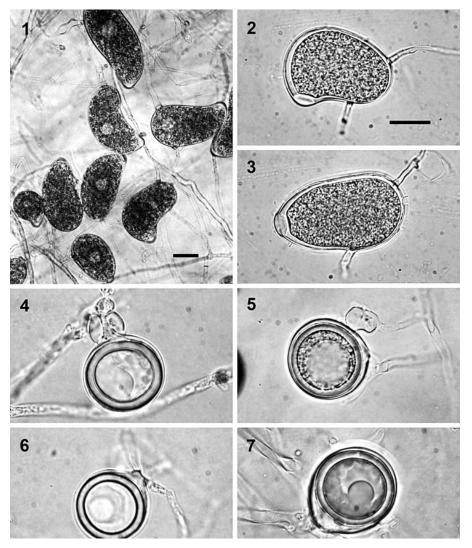
Paragynous antheridia look like knots on the edge of the oogonia because of their closeness. Only one antheridia per oogonium. Oospore are plerotic. Broadly ovoid papilate sporangia occur sympodially and are easily detachable on short pedicels. Antheridia are either amphigynous or paragynous (shown), one – three antheridia per oogonium. Oospore are aplerotic. Obpyriform sporangium with beak-like end near the papila. Sporangia occur singly and are noncaducous.

Species pairs within evolutionary clades: Clade 2 *P. multivora vs. P. siskiyouensis*



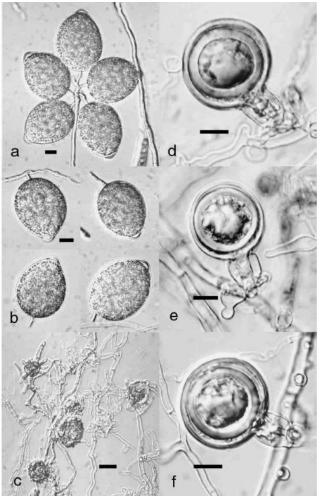


Scott et al. 2009. P. multivora. Persoonia

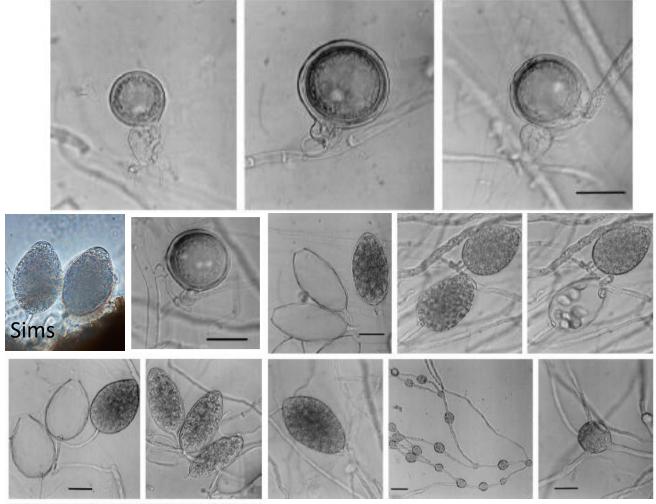


Reeser et al. 2007. P. siskiyouensis. Mycologia

Species pairs within evolutionary clades: Clade 3 • P. nemerosa vs. P. pseudosyringae



Hansen et al. 2003. P. nemerosa. Mycotaxon



Jung et al. 2003. P. pseudosyringae. Mycological Research

Species pairs within evolutionary clades: Clade 6

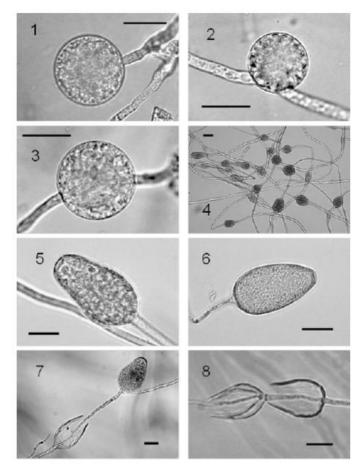
• P. gonapodyides

VS.

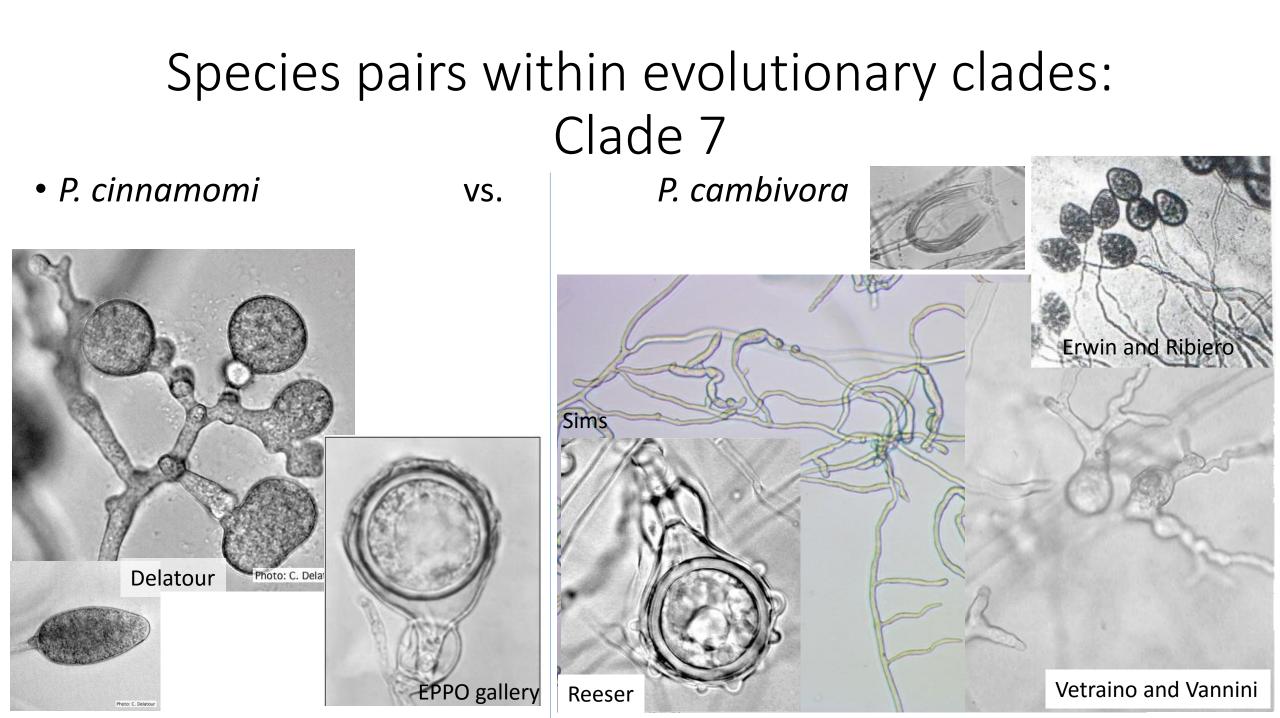
C

Erwin and Ribiero 1996

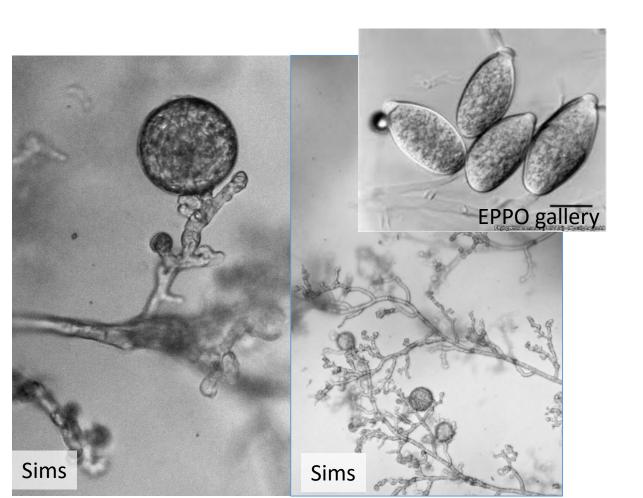
P. chlamydospora

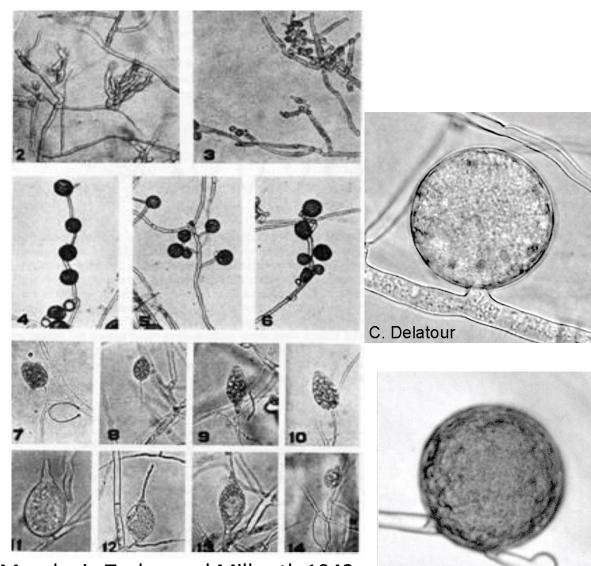


Hansen et al. 2015



Species pairs within evolutionary clades: P. ramorum Clade 8 vs. P. lateralis





C. Delatour

Mycologia Tucker and Milbrath 1942

Overview

- Morphology explained herein, in practice used in some combination with genetic and physiological characters to definatively determine species
- Used to differentiate the genus and species based on visible characteristics
- Can be used to differentiate many of the species within evolutionary clades from a particular crop plant or ecosystem due to the finite number of species that will occur in that system
- Takes time and is somewhat of an art –similar to botany
- Requires the use of *Phytophthora* selective medium and microscopy

Thank you

• Questions?

