

BIOTIC DISEASES caused by:

Parasitic plants

Bacteria

Fungi

Oomycetes

Viruses

Nematodes

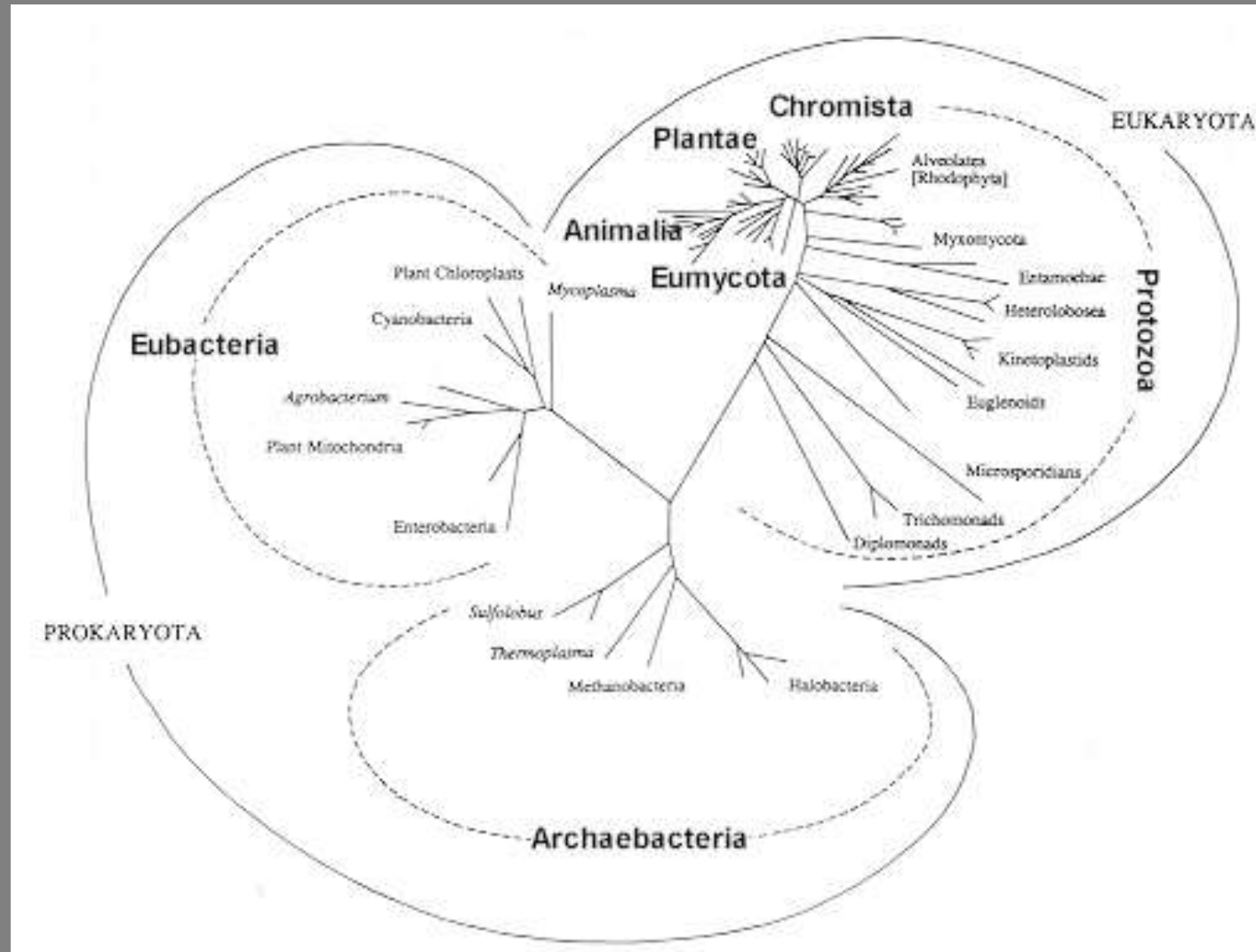
Biotic diseases can be grouped by:

- Agent causing disease (fungus, oomycete, virus, bacteria, plant)
- Host specificity of pathogen: generalist, specialist or “in between” (e.g. limited to a plant family or to conifers)
- Plant part affected (flower , fruit, leaves, branches, stems, roots, vessels, cambium)
- Scale of infection (from one tree to a whole forest)
- Age of host (juvenile vs. adult vs. mature)
- Whether plant tissue is dead or alive: biotroph, hemibiotroph, necrotroph,
- Type of hosts1: whether it affects primary (source) and secondary (sink) hosts
- Type of hosts 2: whether it has transmissive and dead-end hosts
- Type of hosts 3: whether it needs to alternate between different host
- Virulence: whether it is primary, secondary (opportunistic) or a latent pathogen that can change lifestyle
- Wound vs non-wound pathogen
- Airborne vs waterborne and/or soilborne
- Vectored by animals or not
- Pathogen’s reproductive mode: clonal, sexual , mixed

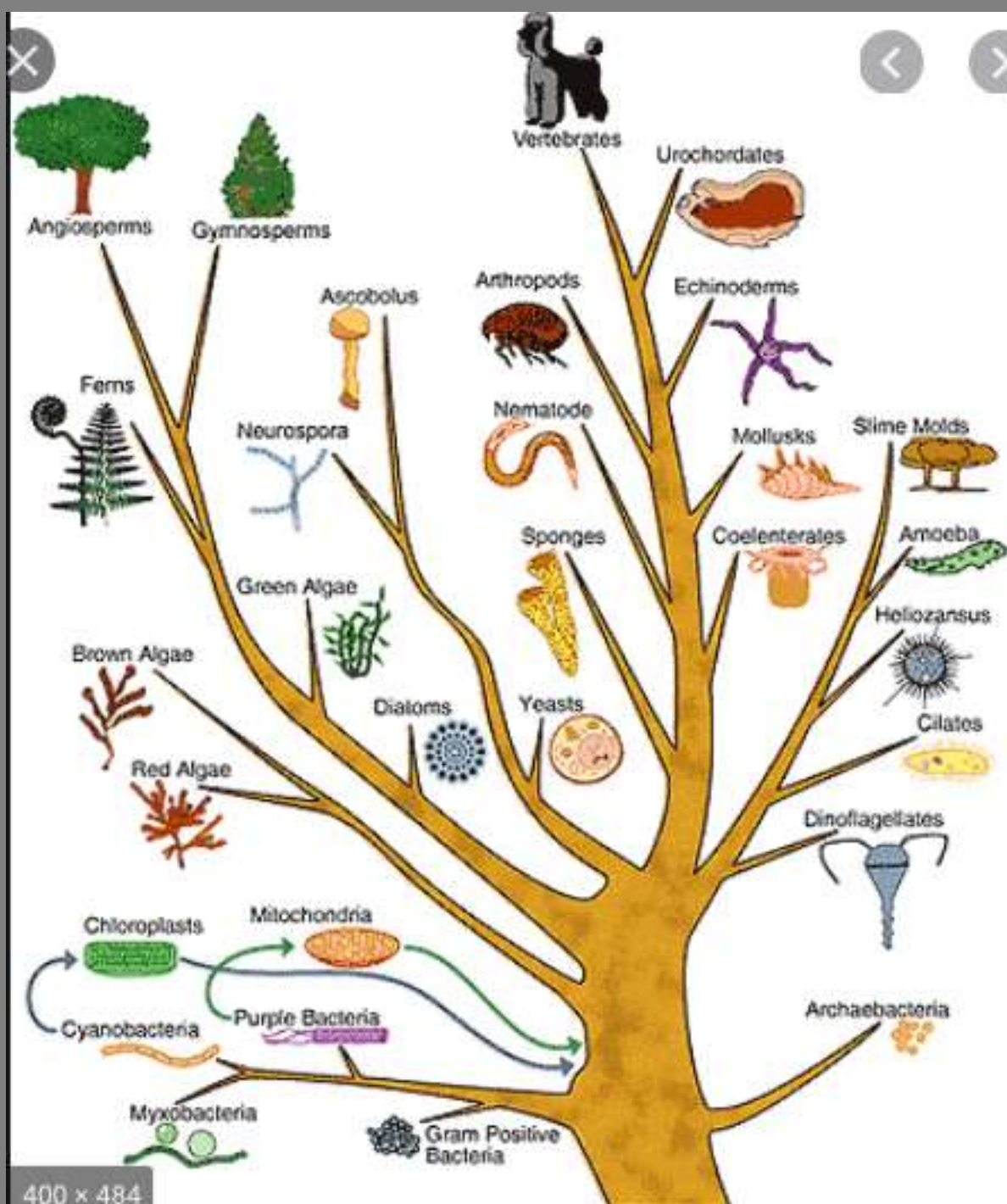
Definitions

- **Biotroph:** a plant pathogen which establishes a long term feeding relationship with the living cells of a host, without killing it as part of the infection process.
- **Hemibiotroph:** an organism that is parasitic in living tissue for some time and then continues to live in dead tissue
- **Necrotroph:** a parasitic organism that kills the living cells of its host and then feeds on the dead matter

Tree of Life, from Patterson & Sogin, 1992



CHROMISTA now referred to as STRAMINIPILA



PARASITIC PLANTS:

True Mistletoe

Dwarf Mistletoe

Dodder

Orchids

Ericaceous plants (Indian pipe)

Phorodenron villosum on oak

Leafy or true mistletoe



UGA4215043b



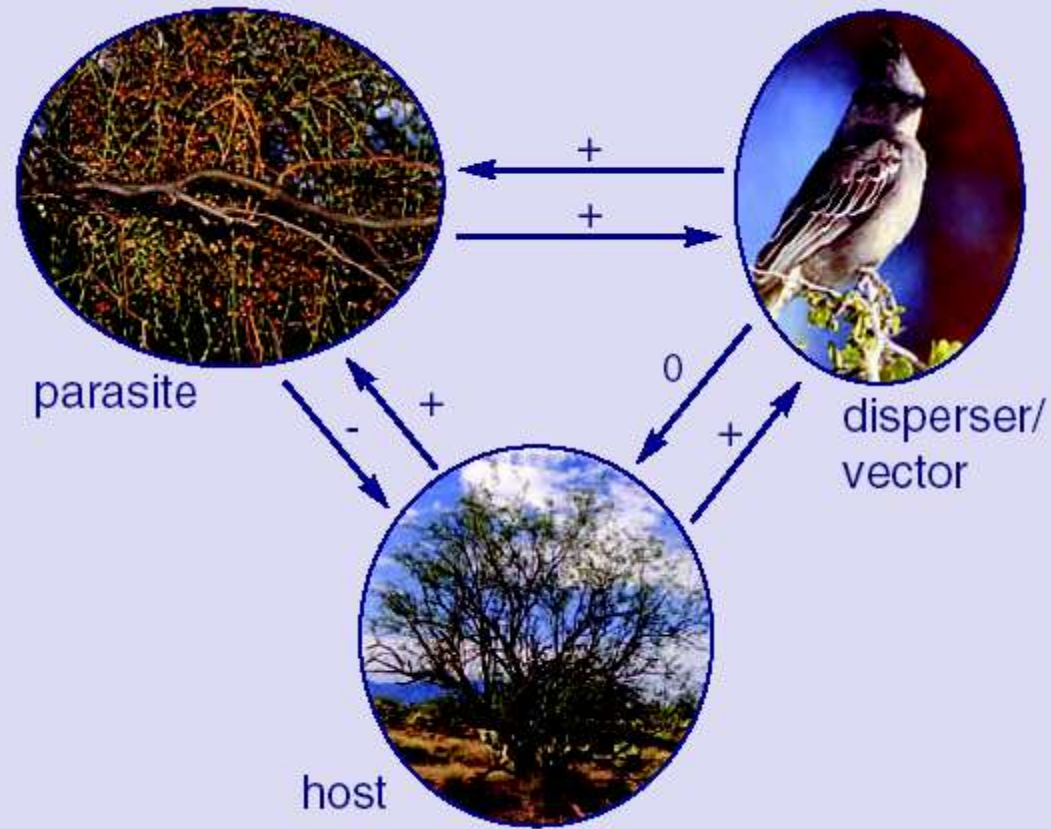


Courtesy of Thomas A. Carlo



Photos courtesy of Thomas A. Carlo

Sticky seed adhere to birds, they also can be ingested and spread in feces



Photos courtesy of Thomas A. Carlo

***Phorodendron pauciflorum*
on white fir**





Phorodendron libocedri
on incence cedar

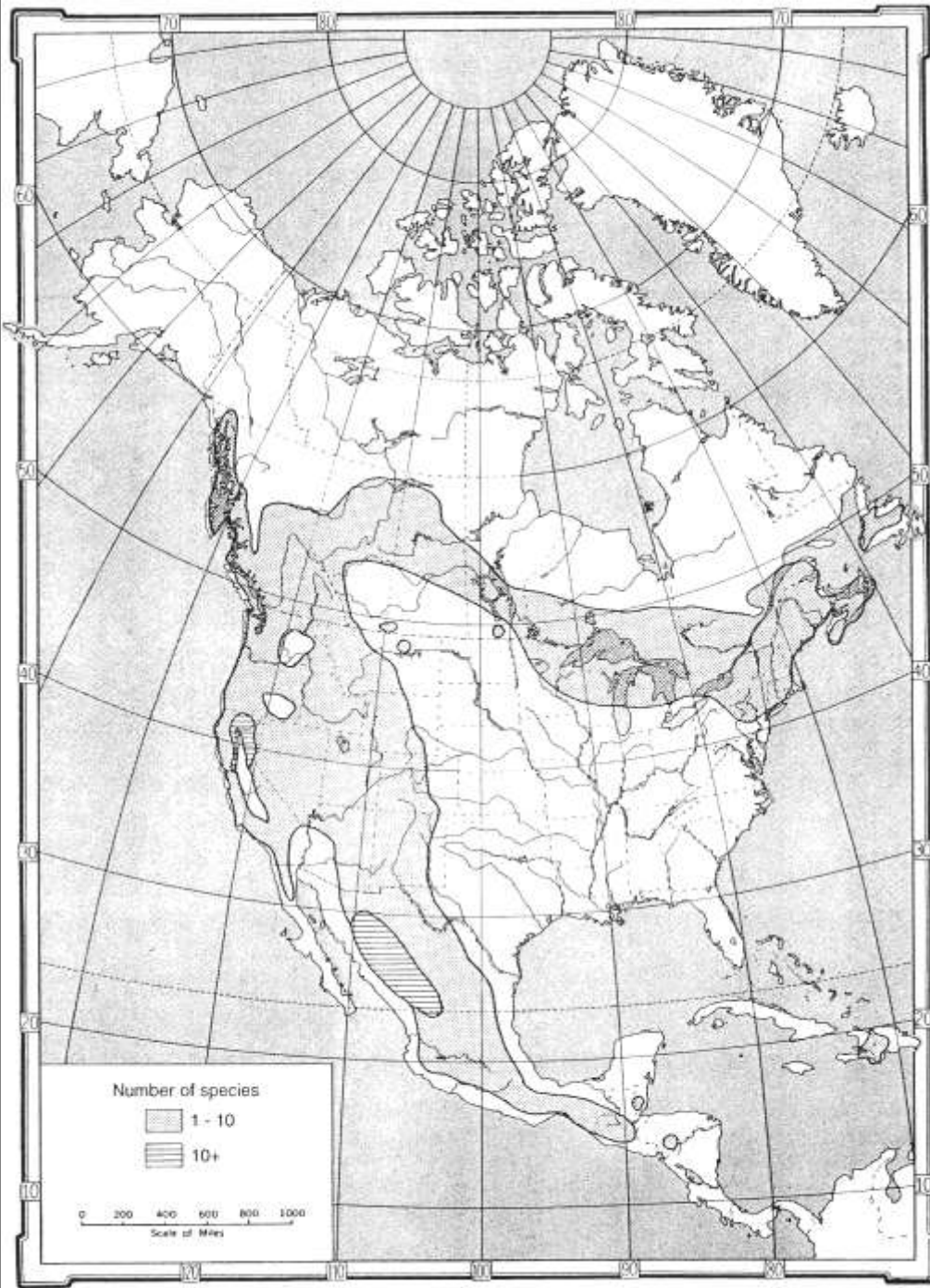






Figure 5.1 –World distribution of *Arceuthobium*. The distribution is primarily in the Northern Hemisphere, crossing the Equator only in Kenya.

Dwarf Mistletoe



Large number of species because of high host specificity and co-evolution between parasite and plant



***Arceuthobium campylopodum*
on Jeffrey pine**



***Arceuthobium douglasii*
on Douglas-fir**



Male flowers



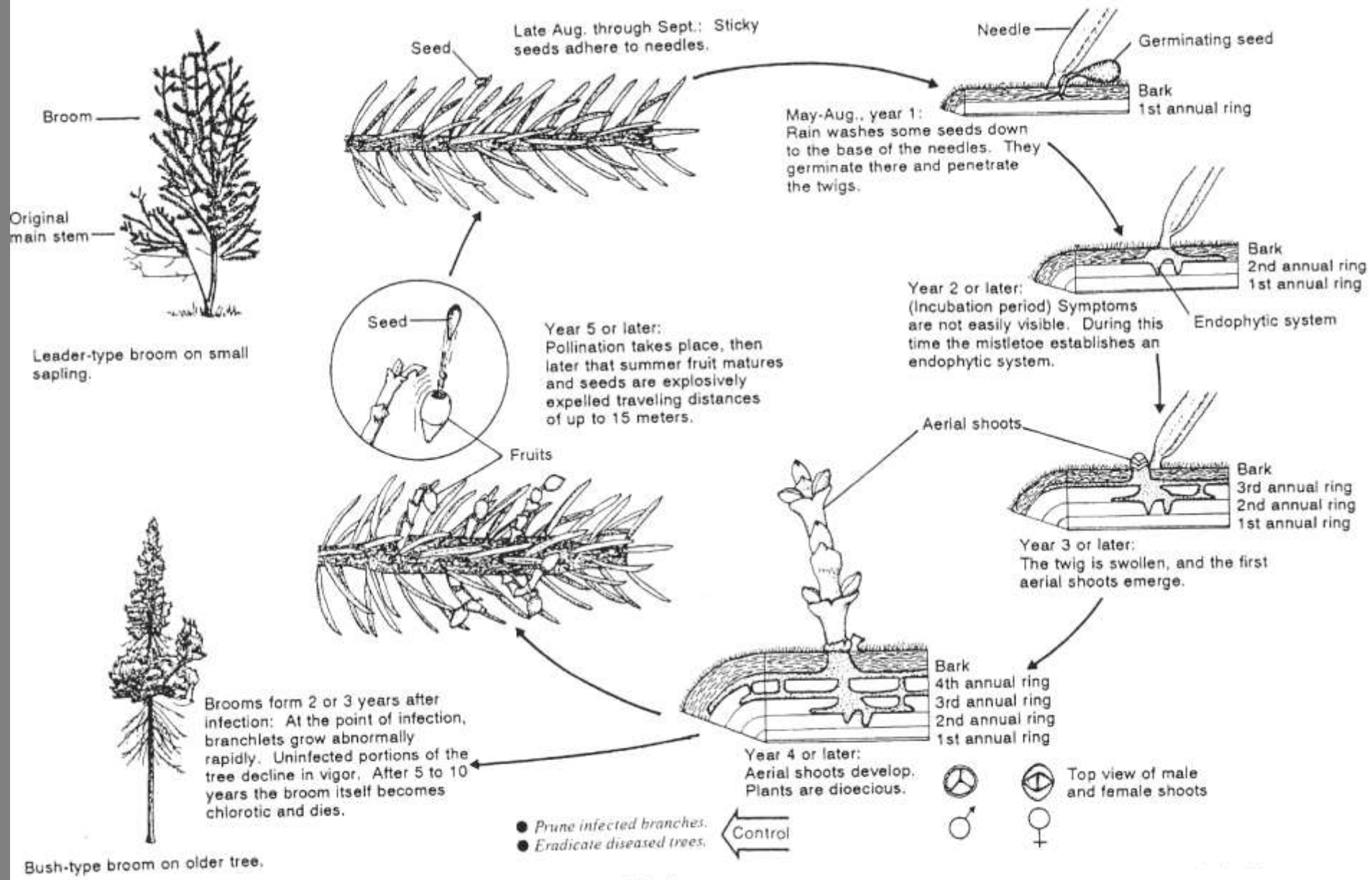
Mature female shoots - seeds



UGA4215030b

Explosive seed dispersal of *Arceuthobium*





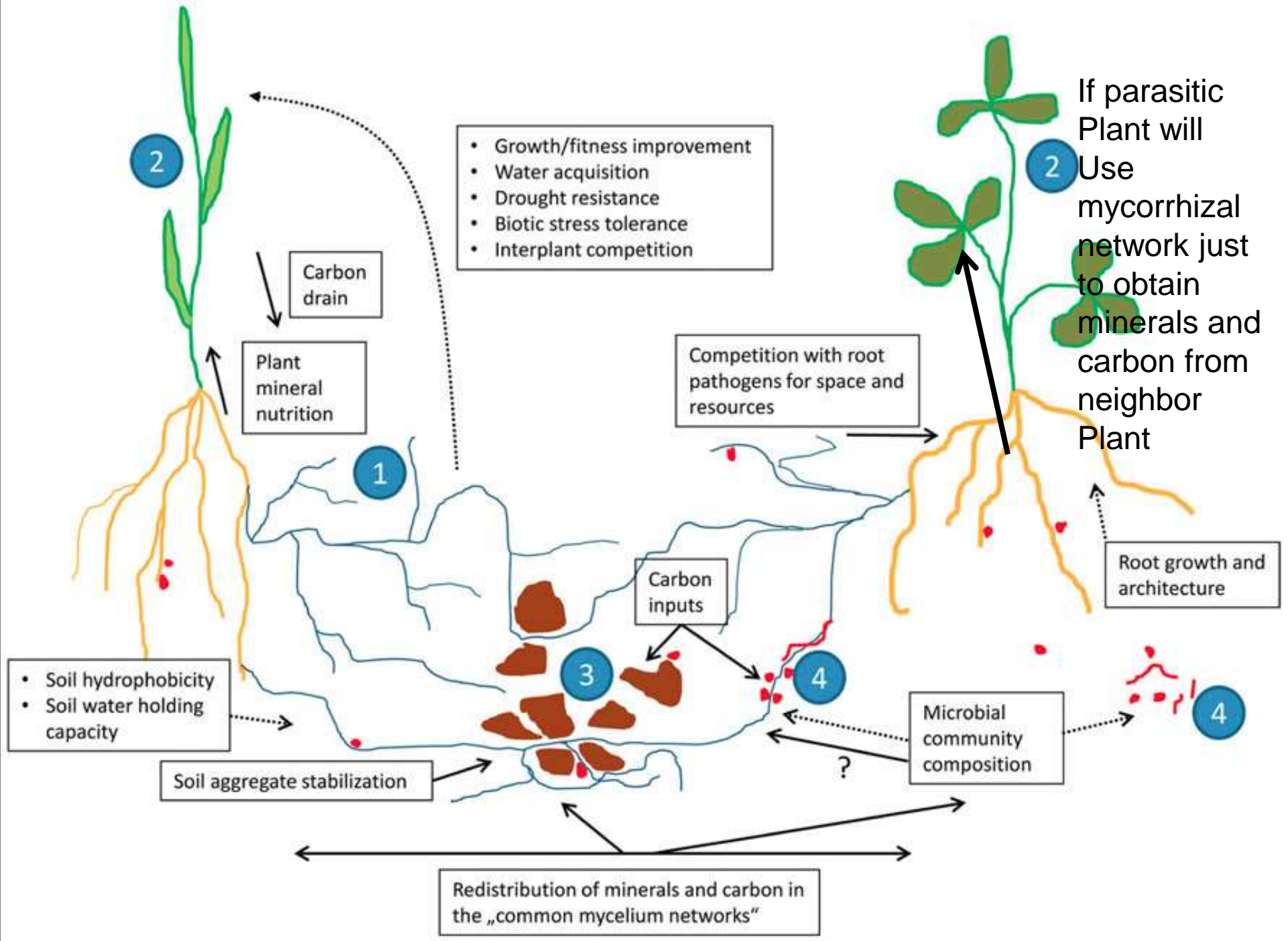
Management issues associated with dwarf mistletoe

- Multi layered forest increases infection
- Group selection also
- You can have up to 30% reduction in growth
- Larger clearcuts is the only way to mitigate problem

Parasitic orchid and ericaceous plants



Coralroot orchids



Proc. Natl. Acad. Sci. USA
Vol. 94, pp. 4510–4515, April 1997
Evolution

Independent, specialized invasions of ectomycorrhizal mutualism by two nonphotosynthetic orchids

(mycorrhiza/ecology/symbiosis/specificity/ribosomal DNA sequences)

D. LEE TAYLOR* AND THOMAS D. BRUNS

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Epiparasitic plants specialized on arbuscular mycorrhizal fungi

Martin I. Bidartondo*†, Dirk Redecker†‡, Isabelle Hijri‡, Andres Wiemken‡, Thomas D. Bruns*, Laura Domínguez§, Alicia Sérsic§, Jonathan R. Leake|| & David J. Read||

BACTERIA

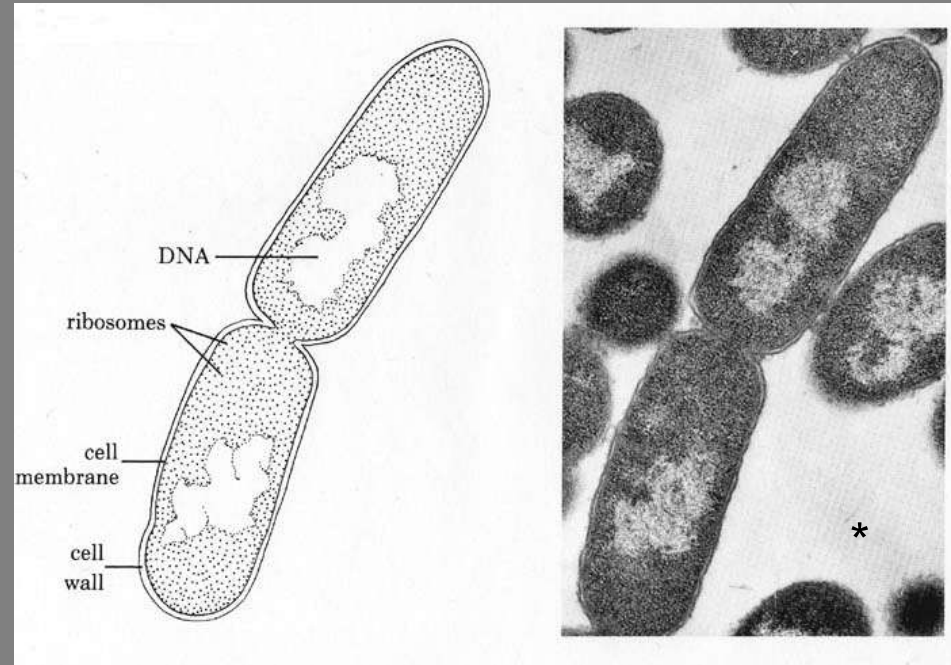
Prokaryotes

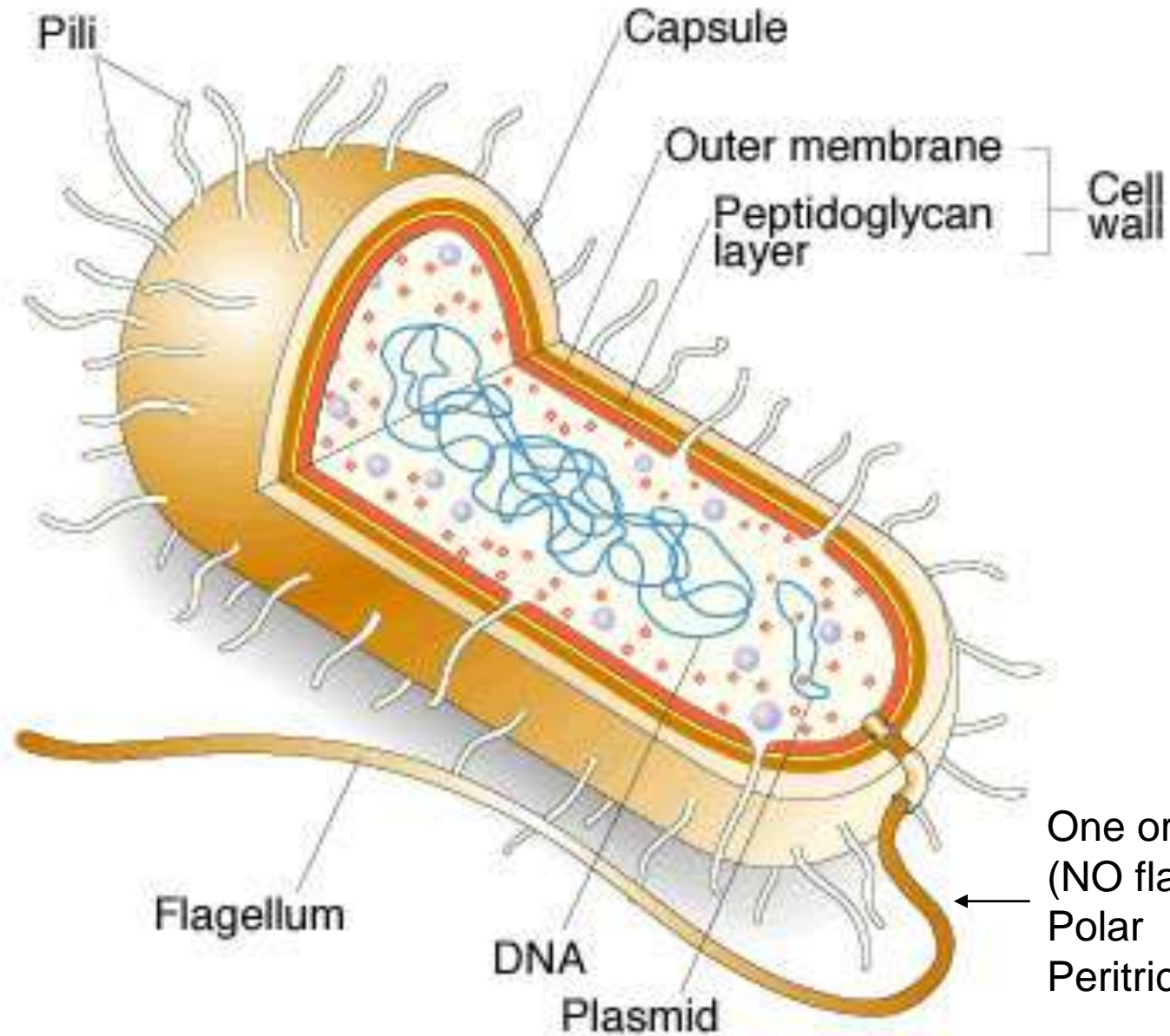
Unicellular

Variously shaped

Do not produce spores

Reproduce by binary fission





One or more flagella
(NO flagella)
Polar
Peritrichous

Genetic elements

Chromosome (single and circular)

Plasmids

Primary mechanisms of variation

Mutations

Loss/acquisition of plasmids and transposons

Recombination through:

Transformation: incorporation in chromosome or as a plasmid of exogenous DNA

Conjugation: incorporation of DNA from another bacterium

Transduction: incorporation through bacterial viruses called bacteriophages



Bacterial Leaf Scorch

Xylella fastidiosa



***Xylella fastidiosa*: The early years**

Anaheim vine disease

- 1882

- 30,000 - 40,000 acres lost

- 50 wineries closed

Pierce investigated viticulture, climate,
epidemiology

Vector and pathogen not known
-thought to be a virus

Isolated, identified as bacterium in 1978



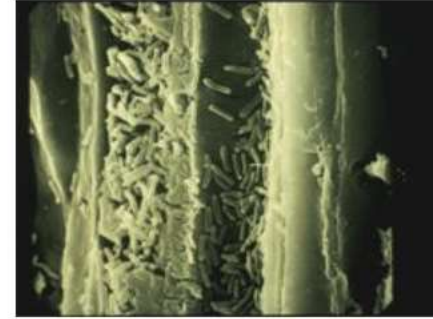
Xylella fastidiosa biology



Xylem-limited bacterium

Wide host range

- crops, native, ornamental, weedy plants
- disease severity differs among hosts



Substantial genetic variation

- host-specific strains
- pathogenicity varies among strains

Transmitted by xylem-sap feeders

- sharpshooters are most important vectors
- many sources of variation

No cure

Mechanism of pathogenicity

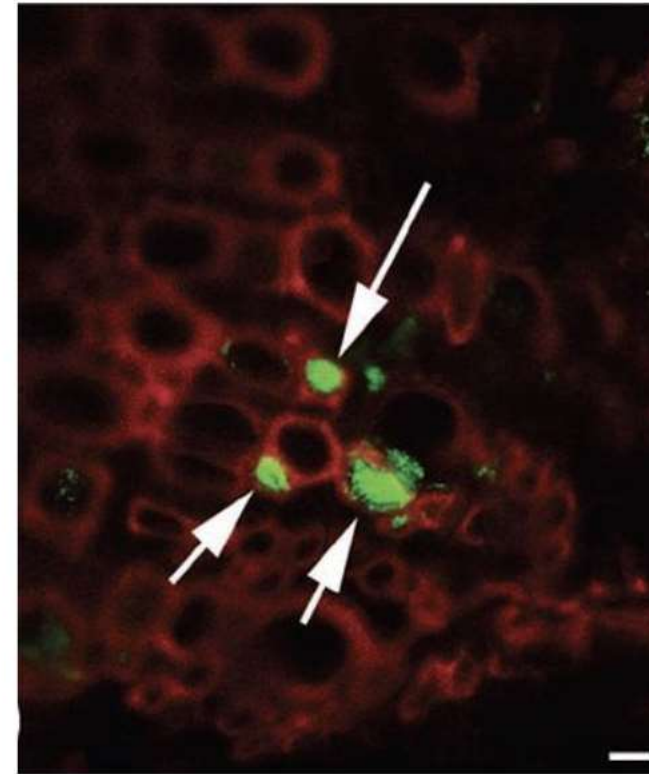
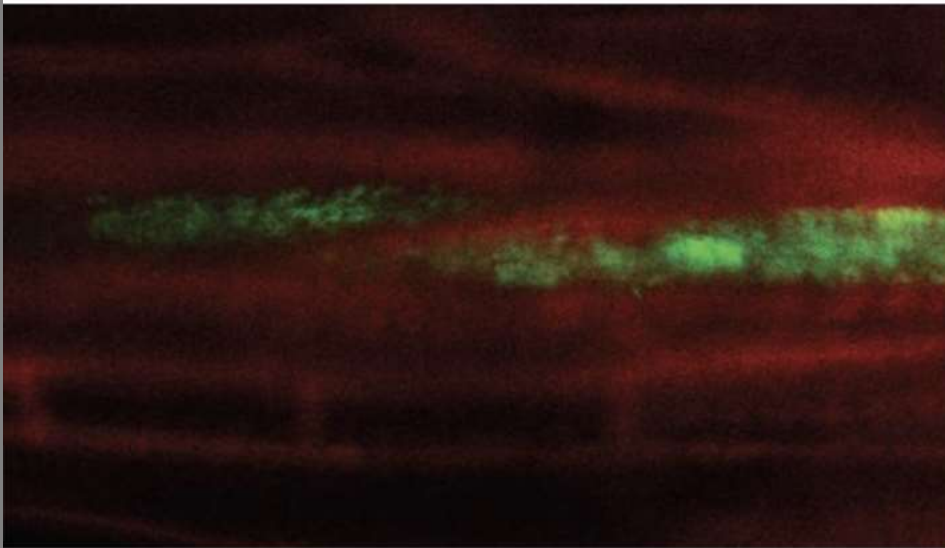


1. Vessel occlusion

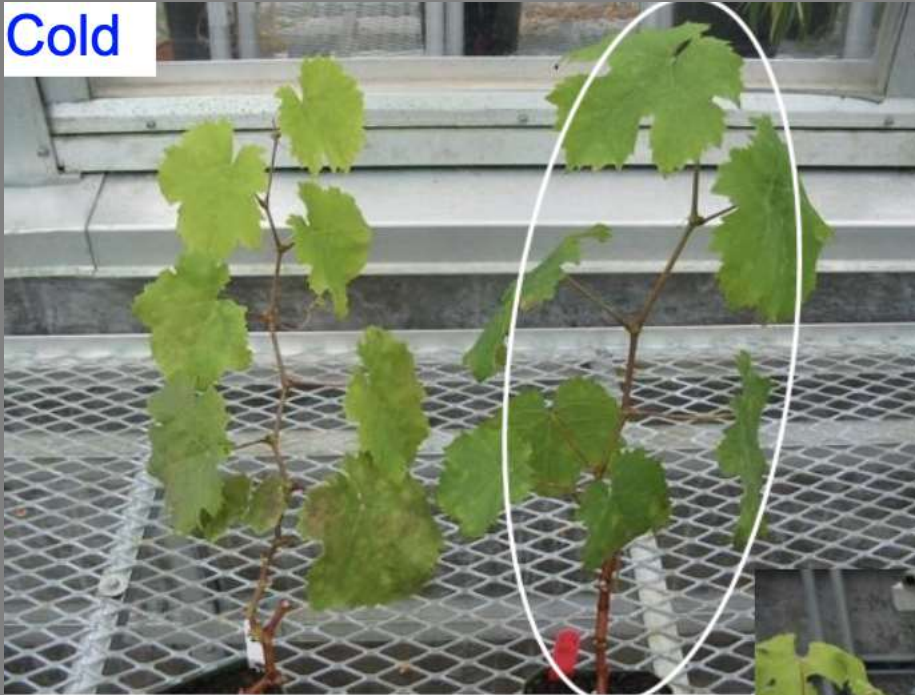
- bacterial aggregates
- restricted water flow
- water stress symptoms

2. "Phytotoxin"

- toxin not known



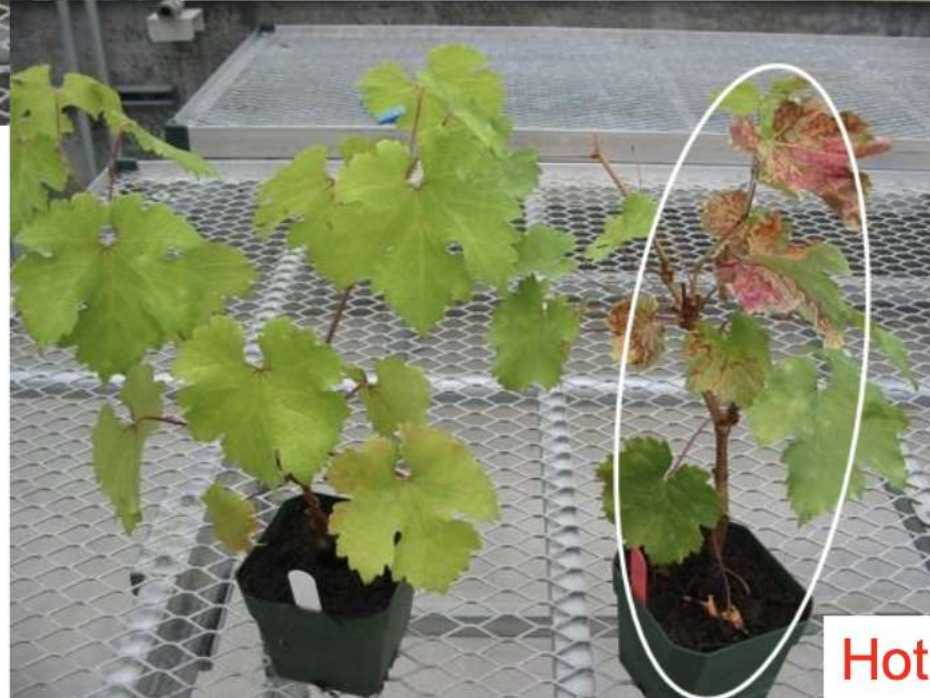
Cold



-mean daily min/max:
17/24°C



-mean daily min/max:
21/36°C



Hot

Hosts

Scientific	Common
<i>Acer</i> sp.	
<i>A. Rubrum</i>	Red maple
<i>A. negundo</i>	Boxelder
<i>A. saccharum</i>	Sugar maple
<i>C. florida</i>	Flowering dogwood
<i>C. occidentalis</i>	Hackberry
<i>L. styraciflua</i>	Sweet gum
<i>Morus alba</i>	Whitemulberry
<i>Platanus</i> sp.	
<i>P. occidentalis</i>	American sycamore
<i>P. x acerifolia</i>	London plane
<i>Ulmus americana</i>	American elm

Scientific	Common
<i>Quercus</i> sp.	
<i>Q. velutina</i>	Black oak
<i>Q. incana</i>	Bluejack oak
<i>Q. macrocarpa</i>	Bur oak
<i>Q. prinus</i>	Chestnut oak
<i>Q. laurifolia</i>	Laurel oak
<i>Q. virginiana</i>	Live oak
<i>Q. rubra</i>	Northern red oak
<i>Q. palustris</i>	Pin oak
<i>Q. stellata</i>	Post oak
<i>Q. coccinea</i>	Scarlet oak
<i>Q. imbricaria</i>	Shingle oak
<i>Q. shumardii</i>	Shumard oak
<i>Q. falcata</i>	Southern red oak
<i>Q. bicolor</i>	Swamp white oak
<i>Q. laevis</i>	Turkey oak
<i>Q. nigra</i>	Water oak
<i>Q. alba</i>	White oak
<i>Q. phellos</i>	Willow oak

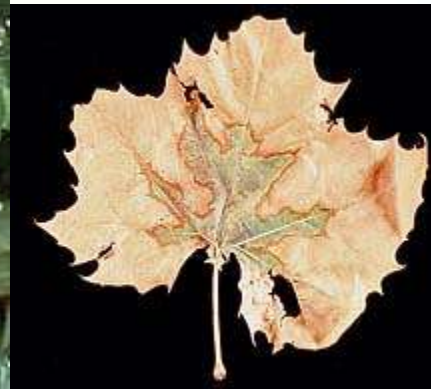
TABLE 1 | Partial list of the main plant hosts of *Xylella fastidiosa* and their *X. fastidiosa* subspecies.

Host scientific name	Type of infection	EPPOCode	Subspecies
<i>Acacia saligna</i>	Incidental	ACASA	pauca
<i>Acer rubrum</i>	Incidental	ACRRE	multiplex
<i>Carya illinoensis</i>	Minor	CYNL	multiplex
<i>Citrofortunella microcarpa</i>	Minor	CJFM	nd
<i>Citronchus</i>	Minor	1CJCG	nd
<i>Citrus</i>	Minor	1CIDG	pauca, fastidiosa
<i>Citrus sinensis</i>	Major	CDSI	pauca
<i>Coffea</i> sp.	Major	COFSS	pauca (BRA)
<i>Coffea</i> sp.	Major	COFSS	fastidiosa (C.Fila)
Cyperaceae	Wild/Wood	1CYPF	nd
<i>Fortunella</i>	Minor	1FOLG	nd
<i>Liquidambar styraciflua</i>	Incidental	LQST	multiplex
<i>Modiola sativa</i>	Minor	MEDSA	fastidiosa
<i>Morus alba</i>	Incidental	MORAL	morus, (former multiplex, sandy)
<i>Morus rubra</i>	Incidental	MORRU	fastidiosa
<i>Nerium oleander</i>	Major	NEROL	sandy
<i>Olea europaea</i>	Major	OLVEU	pauca (ITA, ARG, BRA)
<i>Olea europaea</i>	Major	OLVEU	multiplex (USA, FRA)
<i>Pennisetum americanum</i>	Incidental	PEBAM	nd
<i>Pistacia occidentalis</i>	Minor	PLTOC	multiplex
Poaceae	Wild/Wood	1GRAF	nd
<i>Polygala myrtifolia</i>	Major	POGMY	pauca (ITA)
<i>Polygala myrtifolia</i>	Major	POGMY	multiplex (FRA)
<i>Poncirus trifoliata</i>	Minor	PMTR	nd
<i>Prunus angustifolia</i>	Incidental	PRNAN	nd
<i>Prunus americana</i>	Minor	PRNAR	multiplex
<i>Prunus avium</i>	Minor	PRNAV	pauca (ITA)
<i>Prunus avium</i>	Minor	PRNAV	fastidiosa (USA)
<i>Prunus cerasifera</i>	Incidental	PRNCF	multiplex
<i>Prunus domestica</i>	Minor	PRNDO	multiplex
<i>Prunus dulcis</i>	Minor	PRNDU	multiplex-fastidiosa (USA)
<i>Prunus dulcis</i>	Minor	PRNDU	pauca (ITA)
<i>Prunus persica</i>	Major	PRNPS	multiplex, fastidiosa
<i>Prunus salicina</i>	Minor	PRNSC	multiplex
<i>Quercus palustris</i>	Minor	QUERPA	multiplex
<i>Quercus rubra</i>	Minor	QUERU	multiplex
<i>Sorghum halepense</i>	Wild/Wood	SORHA	nd
<i>Spartium junceum</i>	Incidental	SPJUJ	fastidiosa (USA)
<i>Spartium junceum</i>	Incidental	SPJUJ	multiplex (FRA), pauca (ITA)
<i>Ulmus americana</i>	Minor	ULMAM	multiplex
<i>Vaccinium corymbosum</i>	Minor	VACCO	multiplex
<i>Vaccinium vitigatum</i>	Minor	VACVG	nd
<i>Vitis minor</i>	Incidental	VNMI	pauca (ITA)
<i>Vitis</i>	Minor	1VITG	fastidiosa
<i>Vitis labrusca</i>	Minor	VITLA	fastidiosa
<i>Vitis vulpina</i>	Major	VITVI	fastidiosa
<i>Washingtonia frutescens</i>	Incidental	WESRO	pauca (ITA)
woody plants	Wild/Wood	2WCOF	multiplex

Source: EPPO Global Database (<https://gd.eppo.int/taxon/XYLFAAhosts>), and EFSA Journal database (EFSA, 2015). Major infections are indicated in bold.

Symptoms

- First appear in late summer /early fall
- Leaf scorching
- Limb death



Vector

- Not determined for each tree species yet
- Most likely Graphocephala, Oncometopia and Homalodisca species.



*Graphocephala
atropunctata*



*Oncometopia
orbona*



*Homalodisca
vitripennis*

Distribution Maps of Plant Diseases

Compiled by CABI in association with EPPO

Map No. 262

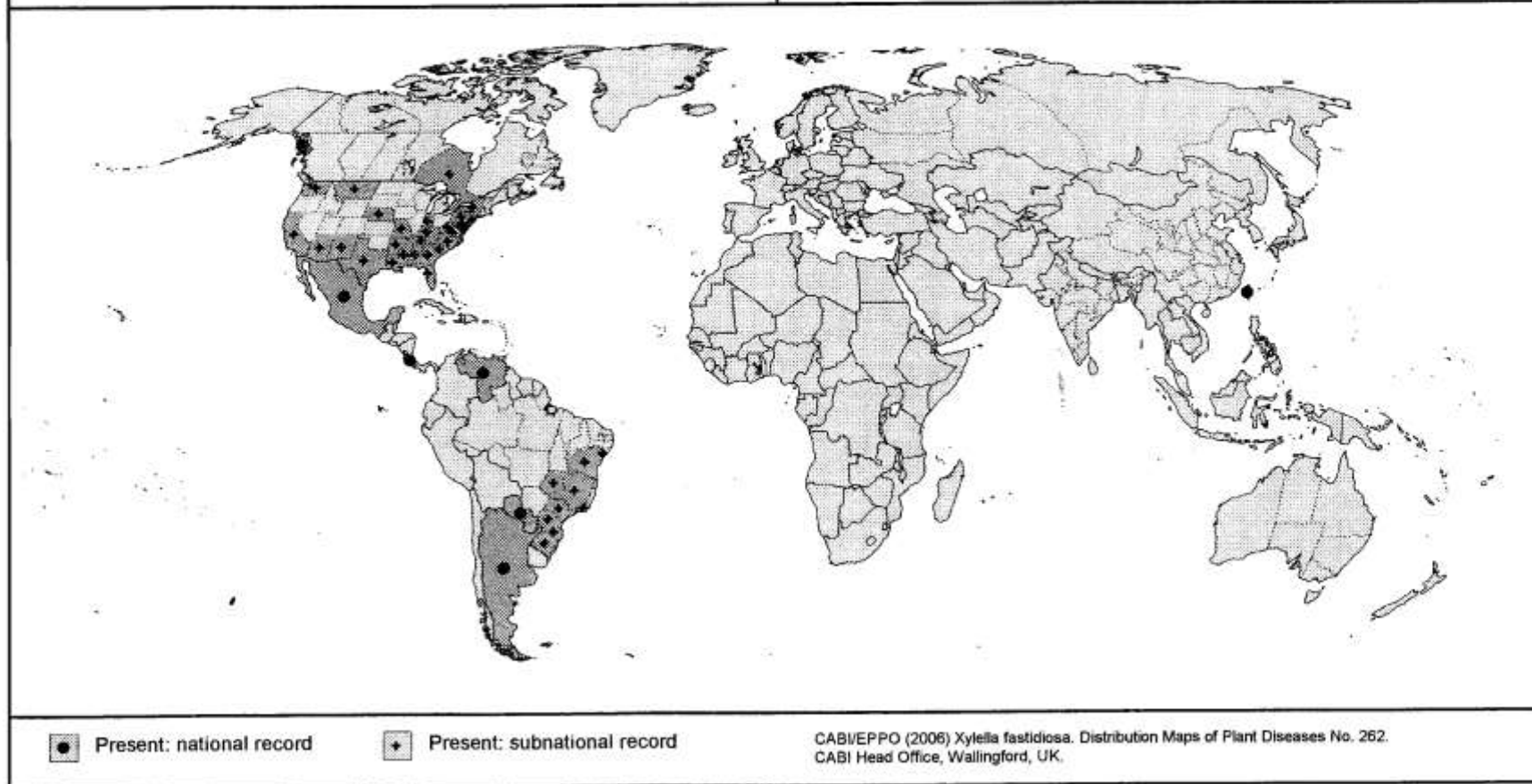
Edition 5

Issued April 2006

Xylella fastidiosa Wells et al.

Bacteria

Hosts: Grapevine (*Vitis vinifera* and others), peach (*Prunus persica*), Citrus, almond (*Prunus dulcis*), lucerne (*Medicago sativa*), some wild trees (incl. *Acer rubrum*, *Platanus occidentalis*, *Quercus rubra*, *Ulmus americana*), other wild plants and weeds.



Map No. 262

Major problem on maples in Pacific Northwest including Northern California



Phytoplasmas

Prokaryotes lacking a cell wall (MLOs - *Mollicutes*)

Usually vascular pathogens

Generally vectored by piercing insects

VIRUSES and VIROIDS

Submicroscopic particles always intracellular when in the host, infectious and pathogenic

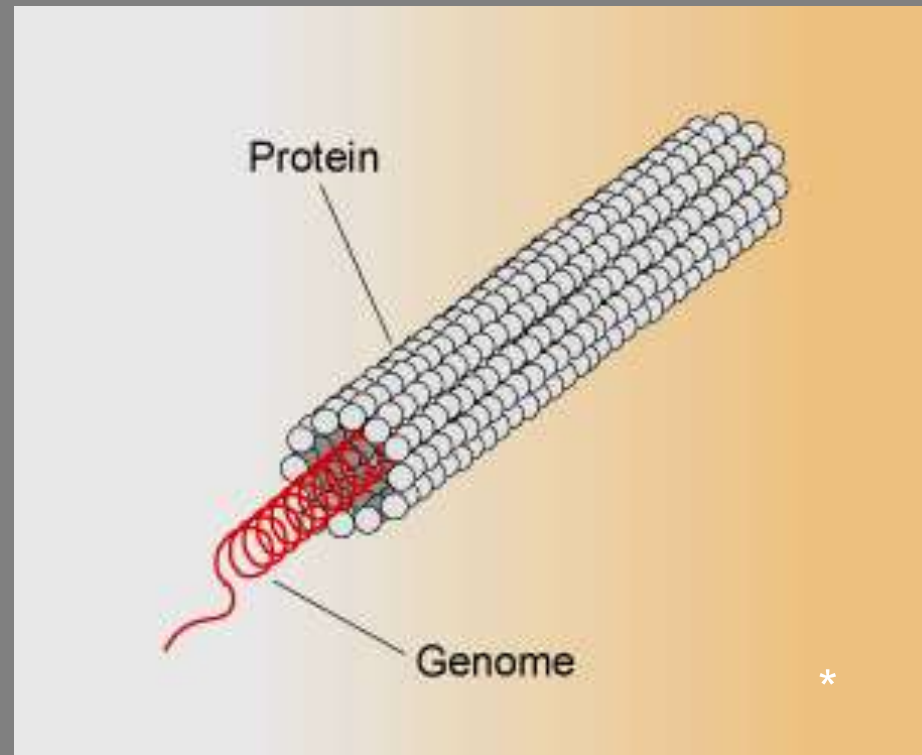
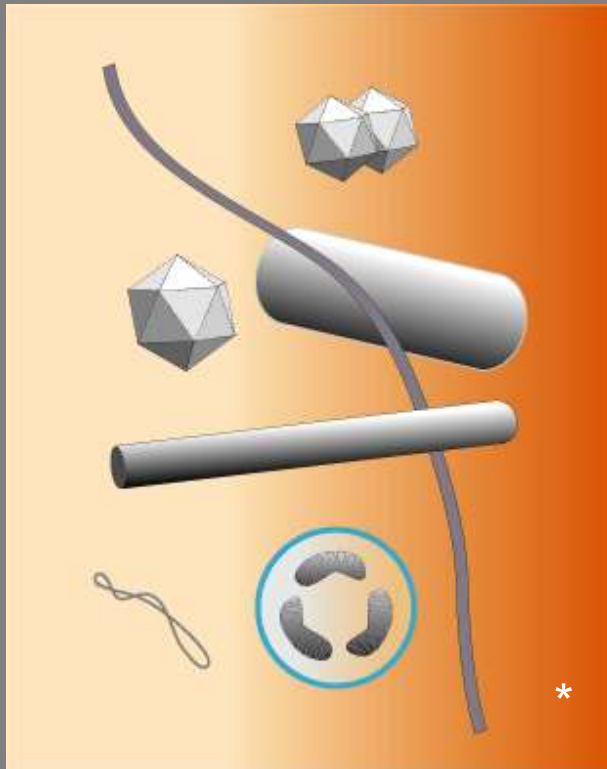
They comprise

Nucleic Acids (RNA or DNA) and a capsid protein

Viroids instead are simply constituted by a single RNA molecule, they do not code for or possess proteins

Nucleic acid: ssRNA, dsRNA, ssDNA, dsDNA

Protein capsid: protects virus during transport



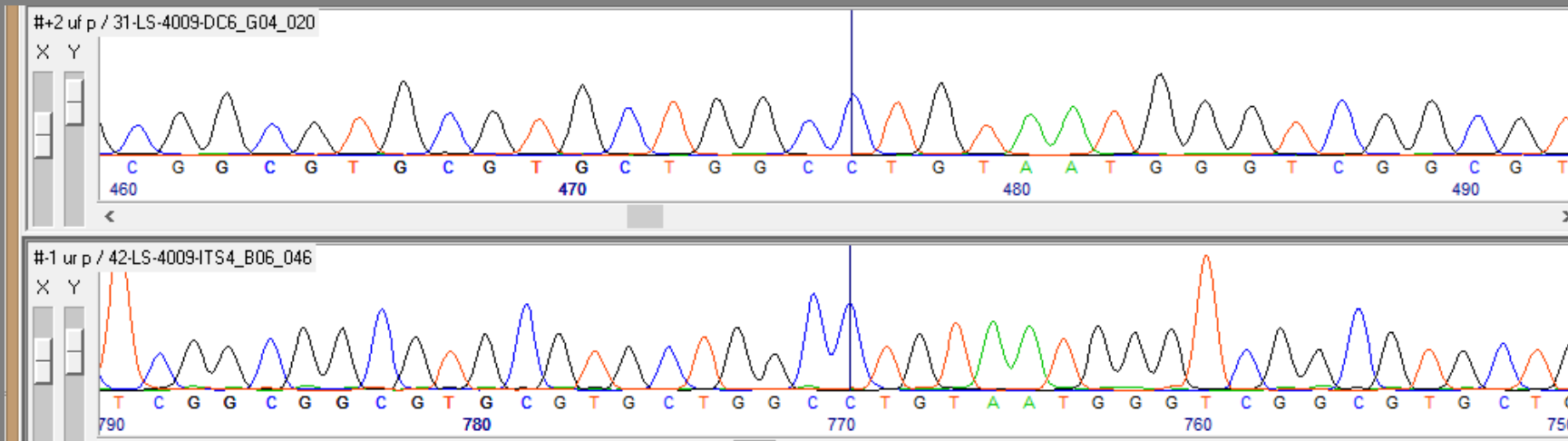
VIRAL REPLICATION

The genome of the virus codes for:

- 1- capsid protein;
- 2- polymerase;
- 3- protein for intracellular movement;
- 4- proteins involved in transmission and relationship with vectors

How does a virus code for all of the necessary proteins in such a small structure?

- 1- One virus may require multiple particles to successfully infect a host. Each particle codes for different genes
- 2- Same DNA or RNA strand, but coding is staggered, i.e. transcription starts at different points, thus one strand can code for multiple genes



→ Gene a

Gene b →

Some examples of bacteria, phytoplasmas and viruses present in forests

- Bacterial leaf scorch: *Xylella fastidiosa*
- Crown Gall: *Agrobacterium tumefaciens*
- Ash and elm yellows: Ca. *Phytoplasma alni* and *P. ulmi*
- Bacterial wetwood (*Enterobacter*, *Klebsiella*, *Erwinia* and *Pseudomonas*)
- Poplar mosaic virus, poplar potyvirus
- Cherry leaf roll virus (elms, dogwood)
- Tobacco Mosaic Virus (tanoak, oaks alders)



A. tumefaciens



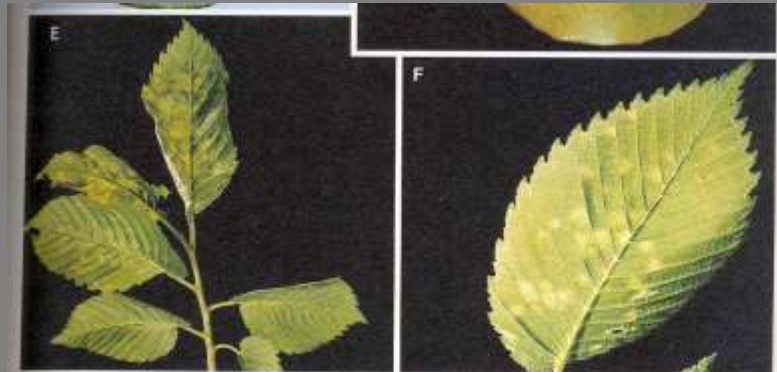
Elm yellows



Ash yellows

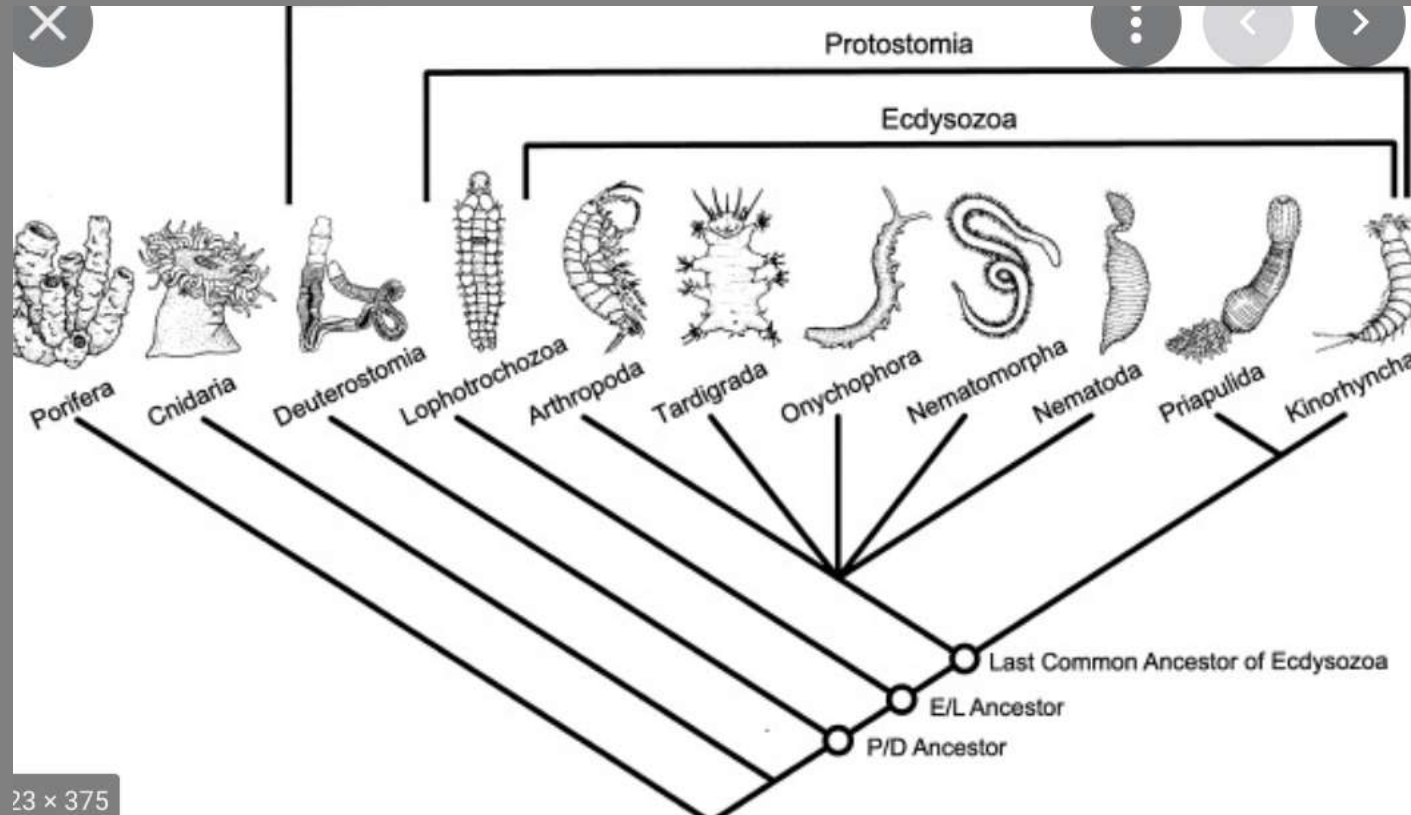


Bacterial wetwood



Cherry Leaf Roll virus

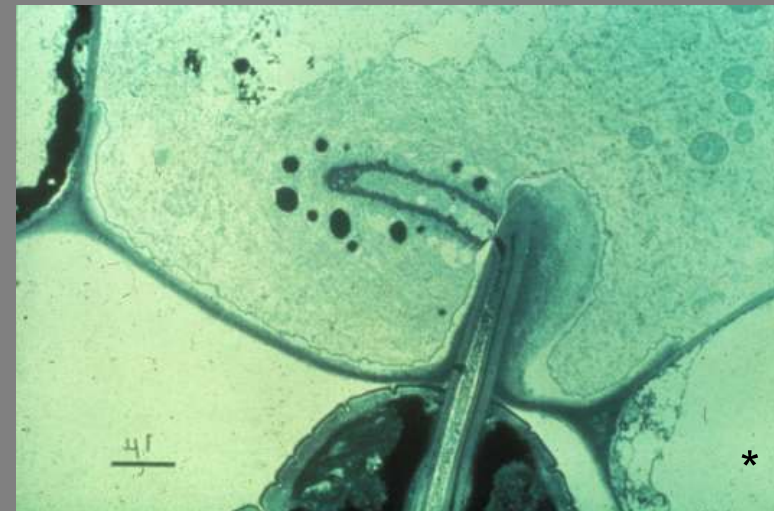
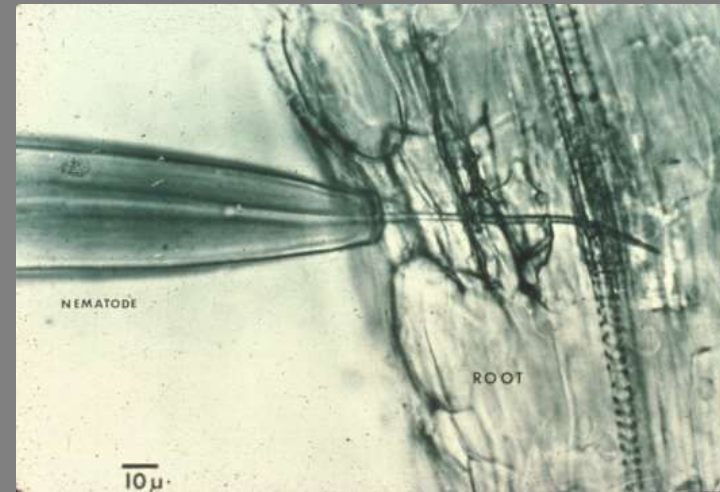
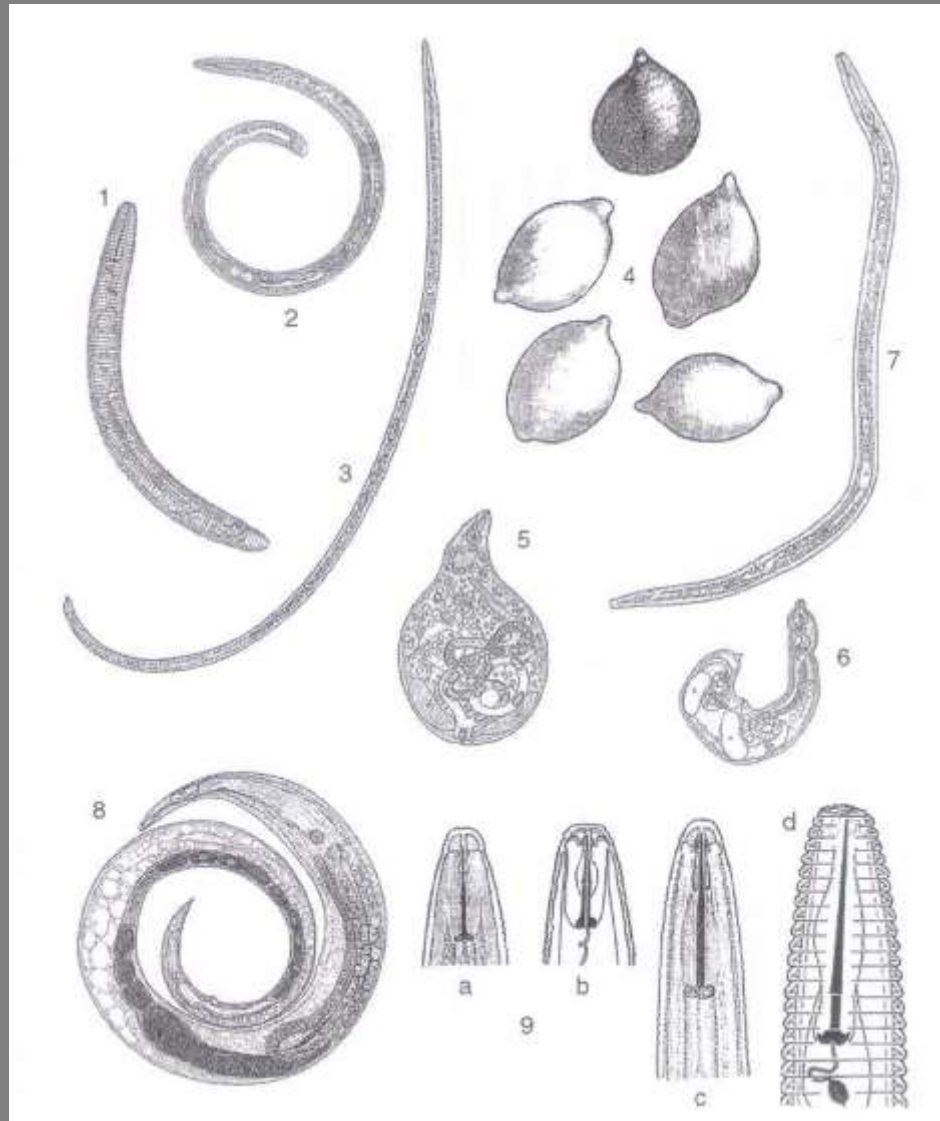
NEMATODES



Very common soil and root pathogens

Can also colonize xylem vessels: Pine Wood Nematode

NEMATODES

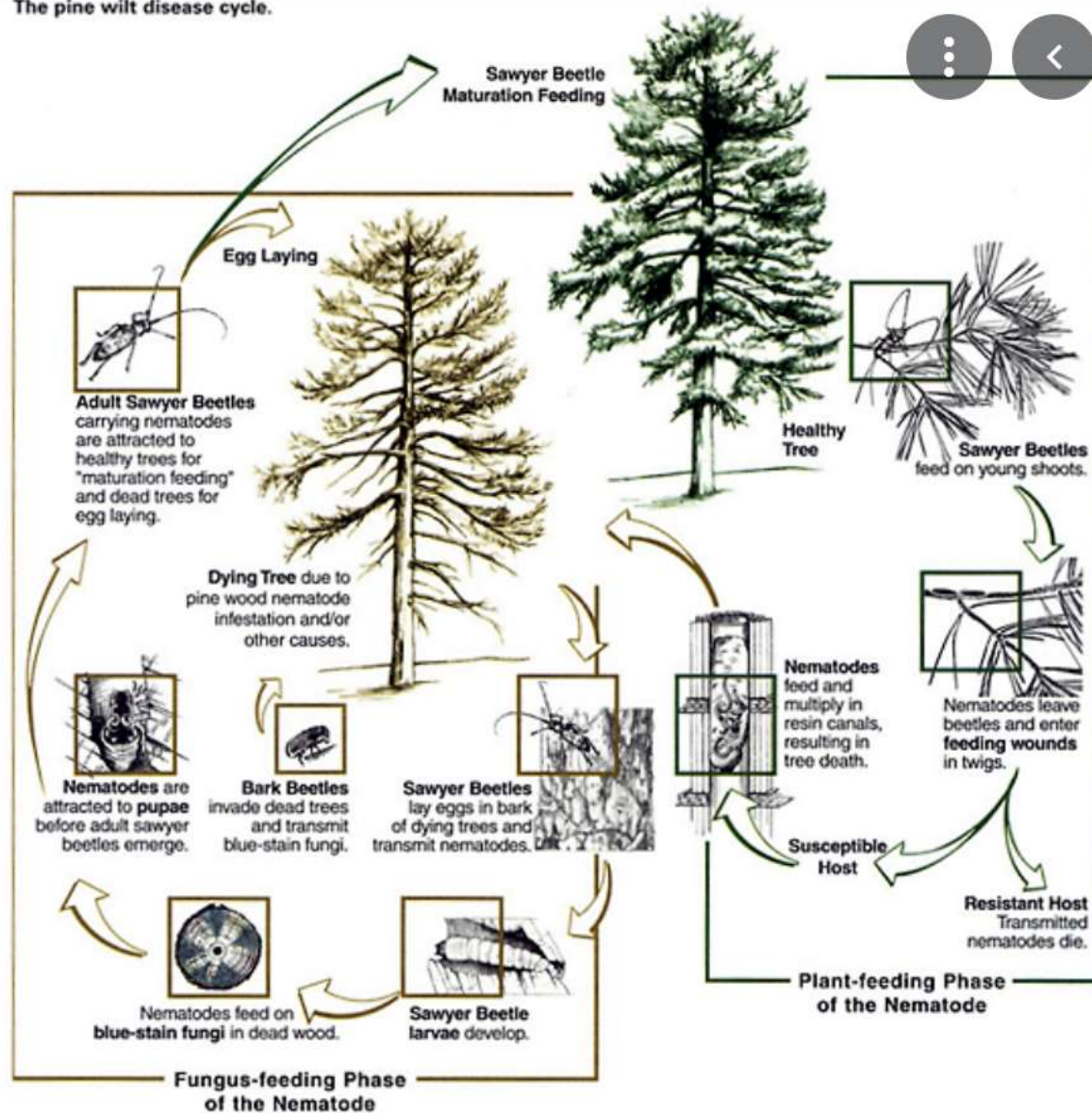


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The pine wilt disease cycle.



Interaction of the pine wood nematode with sawyer beetles to cause pine wilt.

Redrawn with permission from Wingfield, ed. (1987) *Pathogenicity of the Pine Wood Nematode*, APS Press, St. Paul, MN.

Nematode
Bursaphelenchus xylophilus

Longhorn beetles
Genus *Monochamus*

Bark beetles (Ips,
Dendroctonus)
Fungi: Blue Stain Fungi
Ceratocystis

Native to North America
Introduced in East Asia
and Iberian Peninsula



UGA1442032

Fungi

- Eukaryotic organisms, heterotrophs, characterized by chitin and B-glucans in the cell wall, feeding through absorption, reproducing by spores and producing a vegetative structure made up of tubular structures, branched, irregular, and indefinite in growth (modified from B. Kendrick 1992)